

Novel Perspectives for Progesterone in Hormone Replacement Therapy, with Special Reference to the Nervous System

Michael Schumacher, Rachida Guennoun, Abdel Ghomari, Charbel Massaad, Françoise Robert, Martine El-Etr, Yvette Akwa, Krzysztof Rajkowski, and Etienne-Emile Baulieu

Unité Mixte de Recherche 788, Institut National de la Santé et de la Recherche Médicale, and University Paris-Sud 11, 94276 Kremlin-Bicêtre, France

The utility and safety of postmenopausal hormone replacement therapy has recently been put into question by large clinical trials. Their outcome has been extensively commented upon, but discussions have mainly been limited to the effects of estrogens. In fact, progestagens are generally only considered with respect to their usefulness in preventing estrogen stimulation of uterine hyperplasia and malignancy. In addition, various risks have been attributed to progestagens and their omission from hormone replacement therapy has been considered, but this may underestimate their potential benefits and therapeutic promises. A major reason for the controversial reputation of progestagens is that they are generally considered as a single class. Moreover, the term progesterone is often used as a generic one for the different types of both natural and synthetic progestagens. This is not appropriate because natural progesterone has properties very

distinct from the synthetic progestins. Within the nervous system, the neuroprotective and promyelinating effects of progesterone are promising, not only for preventing but also for reversing age-dependent changes and dysfunctions. There is indeed strong evidence that the aging nervous system remains at least to some extent sensitive to these beneficial effects of progesterone. The actions of progesterone in peripheral target tissues including breast, blood vessels, and bones are less well understood, but there is evidence for the beneficial effects of progesterone. The variety of signaling mechanisms of progesterone offers exciting possibilities for the development of more selective, efficient, and safe progestagens. The recognition that progesterone is synthesized by neurons and glial cells requires a reevaluation of hormonal aging. (*Endocrine Reviews* 28: 387–439, 2007)

- I. Introduction
- II. The Recent HRT Trials
- III. Progesterone, Progestagens, and Progestins
- IV. Trophic and Protective Effects of Progesterone in the Nervous System

First Published Online April 12, 2007

Abbreviations: ADX, Adrenalectomized; ARK, aldo-keto reductase; BDNF, brain-derived neurotrophic factor; CBP, CREB-binding protein; CEE, conjugated equine estrogens; CNS, central nervous system; CREB, cAMP response element-binding protein; CSF, cerebrospinal fluid; DHEA, dehydroepiandrosterone; DHEAS, DHEA sulfate; ER, estrogen receptor (isoforms ER α and ER β); GABA, γ -aminobutyric acid; GFAP, glial fibrillary acidic protein; HERS, Heart and Estrogen/Progestin Replacement Study; HRT, hormone replacement therapy; HSD, hydroxysteroid dehydrogenase; hu-mPR α , human membrane progesterone receptor α ; MCAO, middle cerebral artery occlusion; MDN, mediodorsal thalamic nucleus; MPA, medroxyprogesterone acetate; mPR, membrane PR; MRI, magnetic resonance imaging; nAChR, nicotinic acetylcholine receptor; NBM, nucleus basalis magnocellularis; NMDA, N-methyl-D-aspartate; PAIRBP1, plasminogen activator inhibitor RNA binding protein-1; PBR, peripheral benzodiazepine receptor; PEPI, Postmenopausal Estrogen/Progestin Interventions trial; PGRMC1, progesterone membrane receptor component 1 (formerly 25-Dx); PNS, peripheral nervous system; PR, progesterone receptor (PR-A and PR-B); PREG, pregnenolone; RODH, retinol/sterol dehydrogenase; SDR, short-chain dehydrogenases/reductase; SRC, steroid receptor coactivator; SSRI, selective serotonin reuptake inhibitor; StAR, steroidogenic acute regulatory protein; TBI, traumatic brain injury; TSPO, translocator protein (18 kDa) (formerly PBR); WEST, Women's Estrogen for Stroke Trial; WHI, Women's Health Initiative.

Endocrine Reviews is published by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.

- A. Neuroprotective effects
- B. Promyelinating effects
- V. Neurons and Glial Cells in the Aging Nervous System
 - A. Aging neurons
 - B. Aging glial cells
- VI. Gender Differences and Sensitivity to Progesterone
- VII. The Sensitivity of the Aging Nervous System to Progesterone and Estradiol
 - A. Maintained sensitivity to ovarian steroids
 - B. Modified sensitivity to ovarian steroids
 - C. Antagonistic pleiotropy
- VIII. The Timing of HRT: A Therapeutic Window?
- IX. Effects of Progesterone in Peripheral Tissues
 - A. Blood vessels
 - B. Mammary glands
 - C. Bones
- X. Novel Perspectives for Progesterone in HRT: Multiple Signaling Mechanisms
 - A. Progesterone receptor isoforms and nuclear receptor coregulator proteins
 - B. The modulation of neurotransmitter receptors
 - C. Novel membrane receptors of progesterone
 - D. Dependence of steroid signaling on the physiopathological context
- XI. Novel Perspectives for Progesterone in HRT: Different Sources and Local Synthesis
 - A. Peripheral sources of progesterone
 - B. Synthesis of progesterone in the nervous system
 - C. Neurosteroids in the human nervous system

D. Regulation of the local synthesis of progesterone in the nervous system

XII. Conclusions

I. Introduction

A WIDELY USED therapeutic approach for relieving the symptoms, preventing the risks, and reversing some of the pathological changes related to the menopause is to compensate ovarian hormone deficiency by the administration of estrogens, alone or in combination with progestagens. Experimental research and observational clinical data have indeed provided evidence for the beneficial effects of postmenopausal hormone replacement therapy (HRT) on the aging nervous, vascular, and skeletal systems. However, more recently, the utility and safety of chronic hormone use in postmenopausal women has been seriously put into question by three large trials that showed no benefit and even potential hazards of postmenopausal HRT: the Heart and Estrogen/Progestin Replacement Study (HERS), the Women's Estrogen for Stroke Trial (WEST), and the Women's Health Initiative (WHI) (see *Section II*).

Their outcome has been extensively commented upon in the recent literature and has brought many fundamental issues of hormone therapies to light, but discussions were mainly limited to the effects of estrogens (1–5). In fact, progestagens are generally only considered with respect to their usefulness in preventing uterine hyperplasia and malignancy in response to estrogens. Thus, in a recent review on the clinical effects of progestagens, it was stated that “the only indication for the addition of progestins to estrogen-replacement therapy is endometrial protection” (6). There is even a debate about the real usefulness of progestagens in protecting the endometrium, and the possibility of omitting them from HRT has been considered (7). However, this may underestimate the potential therapeutic promises of progestagens, and in particular those of natural progesterone. These have been particularly well documented for the nervous system, where progesterone itself and its metabolites regulate vital neuronal and glial functions and, like estrogens, exert neuroprotective and neurotrophic effects (8–12). On the contrary, the effects of the natural hormone and its metabolites on peripheral tissues, including blood vessels, bone, and even classical targets such as the mammary glands, are still a matter of controversy and need to be studied further.

There are many excellent recent reviews on the effects of estrogens on the brain and on cognitive functions and their potential usefulness in HRT, but the effects of progestagens are surprisingly underrepresented in the literature. Two recent papers had the merit of at least calling attention to the potential usefulness of progestagens for HRT and of reminding readers that menopause is characterized by the concomitant loss of estradiol and progesterone (13, 14). The major aim of the present review is to discuss the pleiotropic effects of progesterone and its metabolites in the nervous system and their implications for preventing or treating age-dependent changes and dysfunctions of the brain and peripheral nerves.

Before discussing the neurotrophic, neuroprotective, and promyelinating actions of progesterone, a succinct description will be provided of the major recent HRT trials that have

stimulated so much debate and also created some confusion. The commonly used nomenclature for progestagens will then be clarified, stressing the differences between natural progesterone and its synthetic analogs. Promising neural targets of progesterone within the aging nervous system will be examined in detail, including neurons, glial cells, and the myelin sheaths, and the question of whether the aging nervous system remains sensitive to its actions will be discussed, as will the question of whether it is meaningful to attempt the treatment of age-related changes with progestagens. As a matter of fact, when addressing such an important and fundamental problem, it is necessary to also refer to the work on estrogens (15–23). Indeed, progesterone and estradiol often act in a concerted manner within target cells, and both steroids frequently exhibit similar properties. However, there are also opposing effects; whereas estrogens increase the excitability of neurons, progesterone and its reduced metabolites in general reduce their activity, an effect that may significantly contribute to their neuroprotective actions (24). The effects of progesterone in peripheral tissues will be examined before drawing attention to novel perspectives for the use of progestagens in HRT, resulting from the recent discoveries of their multiple signaling mechanisms and of their local synthesis by neurons and glial cells.

II. The Recent HRT Trials

Three major prospective clinical trials that have led to the questioning of the usefulness of HRT are the HERS, the WEST, and the WHI. The HERS trial compared the effects of conjugated equine estrogens (CEE) plus medroxyprogesterone acetate (MPA) treatment with placebo on cardiovascular functions in 2763 women with prior coronary disease. Results showed an increase in coronary heart disease during the first year of hormone treatment and no overall cardiovascular benefit with longer follow-up (25). The WEST trial was a randomized, double-blind, placebo-controlled trial of estradiol therapy (1 mg/d) in 664 postmenopausal women (mean age, 71 yr), who already had an ischemic stroke or transient ischemic attack. This large trial found no benefit of estrogen treatment on cerebral stroke incidence, but it found an increased risk of fatal stroke (26).

The WHI trial comprised two very large placebo-controlled arms: combined estrogen plus progestin and estrogen only. The combined estrogen plus progestin WHI trial involved more than 16,000 women with an intact uterus who received either placebo or CEE (0.625 mg/d) plus MPA (2.5 mg/d) (mean age, 63 yr). This arm of the WHI, designed to continue until 2005, was already terminated in 2002 because the overall risks from use of combined HRT outweighed the benefits: there was a slight increase in the risks of breast cancer and of cardiovascular complications, a significant increase in the levels of inflammatory biomarkers, and an increased risk of ischemic stroke (27–30). Within this arm of the WHI trial, the effects of the combined HRT on cognitive functions were examined in a subgroup of 4532 women aged 65 yr or older. This so-called “Women's Health Initiative Memory Study” (WHIM) found no improvement of cognitive functions and no protection against mild cognitive im-

pairment. Instead, the study revealed a very small increase in the risk of cognitive decline and dementia, including Alzheimer's disease (women with dementia: placebo = 22; CEE+MPA = 45 per 10,000 person-years) (31, 32). The explanation for the small increase in dementia is unknown, but it may result from vascular events (33).

The estrogen-alone arm of the WHI trial compared the effects of CEE alone (0.625 mg/d) *vs.* placebo in 10,739 postmenopausal women with prior hysterectomy. The use of CEE in the absence of MPA had no incidence on breast cancer or on coronary heart disease, but again an increased risk of cerebral stroke was observed (34). Thus, in all the trials, HRT was found to be associated with an increased risk of cerebral accidents. A recent retrospective analysis of 28 trials, involving a total number of 39,769 women, was consistent with this conclusion and revealed that among women who had a stroke, those taking HRT had a worse outcome (35). As with CEE+MPA, estrogen alone was also found to have adverse effects on cognition in a smaller recent study involving 2808 women aged 65 yr or older (36).

Subsequent to these trials, many medical organizations have recommended that HRT should not be used for the prevention of age-related diseases and, when used for treating acute climacteric symptoms, only at the lowest dose and for the shortest time. These recommendations have recently been renewed by the French Agency for Health Product Safety. Accordingly, estrogen alone or combined estrogen-progestagen HRT has been relegated to strictly short-term treatment of symptoms such as hot flushes at the beginning of the menopause (37–41).

III. Progesterone, Progestagens, and Progestins

Before discussing the activities of progestagens in the nervous system, it is important to clarify the terminology and to call attention to the fact that not all progestagens behave the same. They do indeed exhibit profound differences according to their structure, and it is certainly not correct to consider them as equivalent compounds, as unfortunately continues to be done. Thus, after the WHI trials, concern has been directed toward progestagens as a single class. Worse, the term progesterone has even been used as a generic one for the different types of natural and synthetic progestagens in recent papers. "Progesterone" should in fact only be used to designate the natural hormone, produced in the corpus luteum of the ovary after ovulation, in the placenta during pregnancy, in the adrenal glands and, as shall be discussed later, also in the central and peripheral nervous systems (CNS and PNS). The term "progestagen" (also sometimes wrongly spelled "progestogen") corresponds to a functional definition and refers to natural or synthetic steroids which, like progesterone, possess progestational activity: preparing and maintaining the uterus for pregnancy. This generally accepted definition may be too restrictive in the light of the pleiotropic actions of progesterone, and in particular of its close metabolites, which do not bind to the intracellular progesterone receptors (PRs), but exert important biological activities. This is the case of allopregnanolone (3 α ,5 α -tetrahydroprogesterone), which is a potent positive modulator of

γ -aminobutyric acid (GABA) type A (GABA_A) receptors and has been qualified as "neuroactive" (42). The multiple functions of allopregnanolone and its interactions with GABA_A receptors will be discussed in detail later.

Progesterone is indeed unidirectionally converted by steroid 5 α -reductases to 5 α -dihydroprogesterone, which also activates gene transcription via the intracellular PR (Fig. 1). These nicotinamide adenine dinucleotide phosphate (reduced form)-dependent enzymes convert a number of Δ 4–3-ketosteroids, including progestagens, glucocorticoids, mineralocorticoids, and androgens, into their 5 α -reduced metabolites. Two 5 α -reductase isozymes are encoded by distinct genes. The type 1 isoform is expressed throughout the rat brain at all stages of development, whereas the type 2 isoform shows a more restricted distribution: it is expressed in the brain almost exclusively around birth, and it is present in the adult spinal cord mainly within gray matter (43, 44).

The bidirectional metabolism of 5 α -dihydroprogesterone is catalyzed by two types of enzymes: the cytosolic nicotinamide adenine dinucleotide phosphate-dependent aldo-keto reductases (ARKs) and a subgroup of the membrane-bound nicotinamide adenine dinucleotide-dependent short-chain dehydrogenases/reductases (SDRs), the so-called retinol/sterol dehydrogenase (RODH)-like group of SDRs (45, 46) (Fig. 1). The four ARK1C1-ARK1C4 isoforms are frequently designated as hydroxysteroid dehydrogenases (HSDs), but also as hydroxysteroid oxidoreductases to insist on the supposedly bidirectional character of the enzyme reactions. However, although bidirectional *in vitro*, the ARKs may only function in the reductive direction in living cells, and they may thus be mainly responsible for the reduction of 3-ketosteroids to 3 α -hydroxysteroids, and more specifically of 5 α -dihydroprogesterone to allopregnanolone (47). On the other hand, the oxidation of 3 α -hydroxysteroids to 3-ketosteroids, and more specifically of allopregnanolone to 5 α -dihydroprogesterone, is thought to be catalyzed by the RODH-like SDRs (46). In humans, the RODH-like SDRs comprise four enzymes with 3 α -HSD activity. Interestingly, two of them also exhibit 3 α →3 β -hydrosteroid epimerase activity, as shown both *in vitro* and in living cells, and they may thus play a crucial role in the control of the local concentrations of biologically active allopregnanolone (48, 49). Indeed, as described in detail in Section X.B, some of the neuromodulatory and protective effects of progesterone are mediated by allopregnanolone, a very potent modulator of GABA_A receptor activity. On the contrary, the 3 β -epimer of allopregnanolone, iso-allopregnanolone (3 β ,5 α -tetrahydroprogesterone), is not only inactive at GABA_A receptors but is also known to antagonize the effects of allopregnanolone (50–52).

The term "progestin" is not used in a consistent manner. It designates both natural and synthetic progestational molecules, including natural progesterone, or exclusively synthetic ones. In the present review, the term progestin will only be used to designate synthetically produced progestagens, including both C19 testosterone derivatives (19-nortestosterone derivatives) and progesterone derivatives (17 α -hydroxyprogesterone derivatives and 19-norprogesterone derivatives) (Fig. 2 and Table 1). The pleonasm "natural progesterone" and "synthetic progestins" will be sometimes used to insist on the difference. The 19-norprogesterone de-

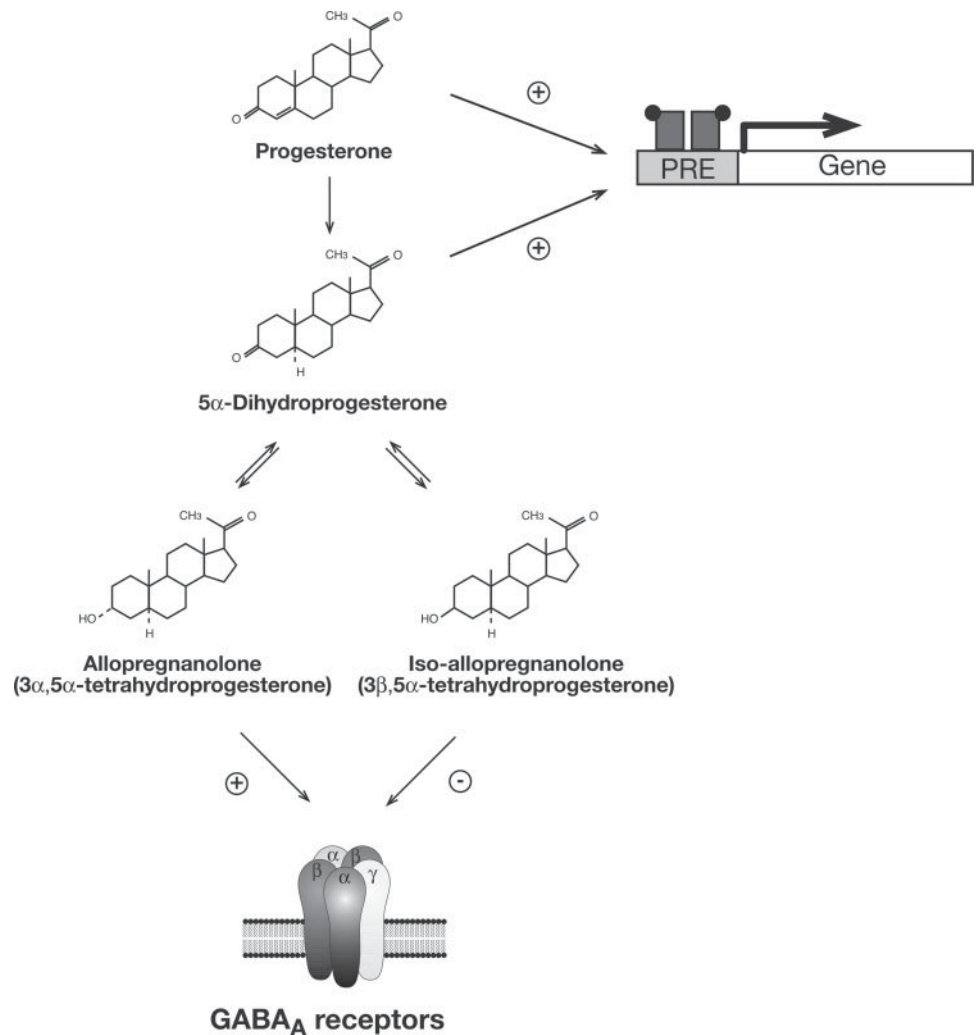


FIG. 1. Metabolism of progesterone. Progesterone is unidirectionally converted by the steroid 5 α -reductases to 5 α -dihydroprogesterone. Both progesterone and 5 α -dihydroprogesterone bind to the intracellular progesterone receptors, which activate gene transcription by interacting with progesterone response elements (PRE, also referred to as glucocorticoid/progesterone response elements, GRE/PRE) often located in the promoter regions of target genes. The bidirectional metabolism of 5 α -dihydroprogesterone is catalyzed by two types of enzymes: the cytosolic ARKs, which may only function in the reductive direction *in vivo*, and the membrane-bound RODH-like group of the SDRs, which oxidize allopregnanolone to 5 α -dihydroprogesterone or epimerize it to iso-allopregnanolone. Whereas allopregnanolone is a positive modulator of GABA_A receptors, iso-allopregnanolone is an inhibitor.

derivatives, such as 19-norprogesterone, promegestone (R5020), and nomegestrol acetate, are among the most selective agonists of the PR, and they are sometimes referred to as “pure” progestagens because as they do not in principle possess androgenic, estrogenic, or glucocorticoid activities (53–55). However, the other progestins bind to several steroid receptors and sometimes exhibit a wide range of non-progestagenic biological effects. Thus, the 17 α -hydroxyprogesterone derivative MPA, the most commonly prescribed replacement progestin in the United States and the one used in the recent large HRT trials, also has androgenic and glucocorticoid properties (56). The synthetic 19-nortestosterone-derived progestins, such as norethisterone acetate, a progestin commonly used in Europe, retain varying degrees of androgenic activity despite the removal of carbon 19 (57, 58).

In contrast to progesterone, progestins are not converted to the GABA_A receptor-active metabolite allopregnanolone, whose importance in mediating some of the biological effects of progesterone will be discussed later. Nevertheless, progestins are also extensively metabolized in various tissues, but their metabolites are not well characterized. Some of the 19-norprogestins may have the potential to be converted to neuroactive metabolites. Thus, 19-nortestosterone-derived

progestins including norethisterone, levonorgestrel, and gestodene are extensively converted to 5 α -, 3 α ,5 α - and 3 β ,5 α -reduced metabolites (59, 60). Interestingly, whereas the 5 α -reduction significantly increases the androgenic potency of testosterone, the 5 α -reduction of norethisterone results in a significant diminution of androgenicity (61). The 3 β ,5 α -reduced metabolites of norethisterone, levonorgestrel, and gestodene bind to ER α , although with a lower affinity than estradiol, and activate gene transcription via this receptor (60, 62, 63).

Whether A-ring-reduced metabolites of progestins act on GABA_A receptors needs to be clarified. Norethisterone acetate and MPA were shown to produce some anxiolytic-like effects when rats were tested in the “elevated plus maze” and the “shock-probe burying test”. In contrast, the norprogesterone derivative trimegestone only had little effect (64). However, these behavioral effects of the progestin metabolites do not necessarily result from the direct modulation of GABA_A receptors. Indeed, some observations suggest that the administration of progestins affects the concentrations of endogenous allopregnanolone in the brain and influences the activity of enzymes involved in the metabolism of progestagens (65, 66). For example, MPA (Provera), which does not

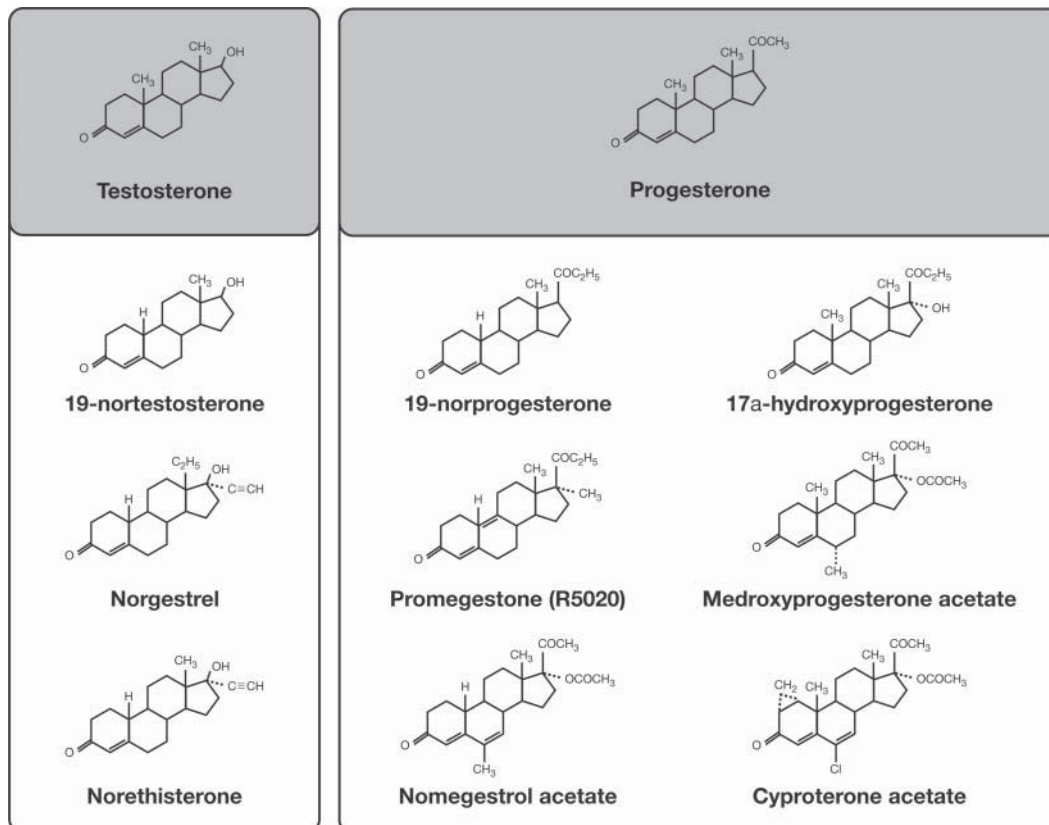


FIG. 2. Chemical structures of progestins. Comparison of the chemical structures of testosterone, progesterone, and progestins, comprising testosterone derivatives and progesterone derivatives (19-norprogesterone and 17 α -hydroxyprogesterone derivatives). Small structural changes account for important differences in the effects of progestins.

directly act on GABA_A receptors, enhances GABA_A receptor-mediated inhibitory neurotransmission in the rat hippocampus by inhibiting the metabolism of allopregnanolone (67). Another recent study has shown that 2-wk oral treatment with MPA increases allopregnanolone levels within the hippocampus, cerebral cortex, and hypothalamus of ovariectomized female rats (68). The effect of MPA on the endogenous levels of brain allopregnanolone may explain why the progestin is therapeutically beneficial for catamenial epilepsy

(69) and sometimes improves anxiety and mood in postmenopausal women (70, 71).

Only a few progestins have been tested for their effects on the nervous system, but concerns are particularly serious about the negative effects of MPA. Thus, MPA has been shown to antagonize the neuroprotective and promnesic effects of estrogen. Whereas progesterone and 19-norprogesterone, alone or in combination with estradiol, protected cultured hippocampal neurons against glutamate toxicity,

TABLE 1. Classification of synthetic progestins

19-Nortestosterone derivatives	19-Norprogesterone derivatives (C-20)	17 α -Progesterone derivatives (C-21)
Estranes (C-18)	Promegestone (R5020)	Medroxyprogesterone acetate (MPA)
Norethisterone (NET) (= norethindrone)	Nomegestrol acetate (NOMAC)	Cyproterone acetate
Norethisterone acetate (NETA)	Trimegestone (TMG)	Chlormadione acetate
Gonanes (C-17)	Nestorone (NES)	Megestrol acetate
Levonorgestrel (LNG)	Demegestone	Others
Desogestrel (DSG)	Medrogestone	Drospirenone (DRSP)
Gestodene (GES)		Dydrogesterone
Norgestrel (NG)		
Norgestimate		
Etenogestrel		
Norgestimate		
Dienogest (DNG)		

Progestins are derived from either testosterone (19-nortestosterone derivatives) or progesterone (19-norprogesterone derivatives and 17 α -hydroxyprogesterone derivatives). The 19-nortestosterone derivatives are further subdivided into estranes (with 18 carbons) and gonanes (with 17 carbons). The gonanes norethynodrel, lynestrenol, and ethynodiol are converted to norethisterone in the body. Other progestins are drospirenone, derived from spiro lactone, and dydrogesterone, a highly selective stereoisomer of natural progesterone (a retroprogesterone with an additional double bond between C-6 and C-7) (adapted from Refs. 53, 55, 826, and 827). For the pharmacological characteristics of progestins, see Refs. 54 and 828–830.

MPA not only failed to be effective but also attenuated the estrogen-induced neuroprotection. At the molecular levels, MPA blocked estrogen-induced expression of the antiapoptotic protein Bcl-2 and antagonized estradiol-induced attenuation of the glutamate-induced rise in intracellular calcium (72, 73). Thus, one of the most prescribed progestins for HRT and contraception opposes some of the beneficial effects of estradiol in the brain and may even exacerbate the excitotoxic death of neurons (74). *In vivo*, MPA has recently been reported to diminish the ability of CEE to reduce stroke damage in subcortical regions of the rat brain (75). In female monkeys, treatment with MPA reduced the increase in sexual initiation induced by estradiol treatment and increased aggressive behavior, which may represent a serious behavioral side effect (76). MPA has also been shown to directly inhibit the activity of steroidogenic enzymes, in particular of the human type II 3β -HSD, an enzyme that converts pregnenolone (PREG) to progesterone, and the progestin thus interferes with steroid biosynthetic pathways (77).

It is important to draw attention to differences in HRT regimens between countries. In the United States, the most commonly used progestin is MPA, generally combined with CEE, an association of more than 10 different estrogens. Most of them are sulfated and distinct from the predominant endogenous estrogens in women, that is, estradiol before and estrone after menopause (78). Nevertheless, estrogen components of CEE have recently been shown to have potent antioxidant and neuroprotective effects and also to reduce the cortical infarction volume in a rodent model of stroke (75, 79–81). In the United Kingdom, the progestins mainly used are 19-nortestosterone derivatives (norethisterone acetate, norgestrel, and levonorgestrel). In central and southern Europe, both 19-nortestosterone derivatives and a range of progesterone derivatives are used. In France, micronized progesterone and 19-norprogesterone derivatives are commonly prescribed in combination with oral or transdermal estradiol (82–84). It would certainly be worthwhile to attempt retrospective comparisons of the different HRT formulations.

Oral micronized progesterone has been widely used in Europe, and in particular in France, since 1980. Micronized progesterone is natural progesterone, whose average particle size has been reduced, leading to decreased destruction in the gastrointestinal tract, a longer half-life, and enhanced bioavailability. Before, the discovery of the micronization process, progesterone could not be taken orally because it is poorly absorbed and rapidly metabolized. The use of micronized progesterone is well tolerated, with mild and transient sedation as a side effect that can be minimized by taking the hormone at bedtime (85). Moreover, the elevation of circulating levels of progesterone by oral administration of the micronized hormone has been shown to be as effective as the administration of progestins for the control of endometrial growth (86, 87). Earlier studies have also reported that micronized progesterone may improve mood in patients with premenstrual mood disturbances and in postmenopausal women (88, 89). When compared with the MPA-containing regimen, micronized progesterone was found to improve significantly vasomotor symptoms, somatic complaints, anxiety, and depressive symptoms in postmenopausal women (90). However, work by Bäckström and col-

laborators (91, 92) has shown that treatment with progesterone can also result in adverse mood changes (tension, irritability, depression), and that the metabolite allopregnanolone may be the mediator of these effects. Thus, in two studies of postmenopausal women with climacteric symptoms, negative mood effects during treatment with vaginal progesterone implants were related to the blood concentrations of allopregnanolone. During the progesterone treatment period, women had increased negative mood symptoms when compared with the estradiol-only period, but only when serum concentrations of allopregnanolone were increased to those seen during the midluteal phase of the menstrual cycle, not when they were either higher or lower (91, 92). These observations suggest a bimodal association between allopregnanolone and adverse mood, and they point to the importance of a well-dosed HRT.

IV. Trophic and Protective Effects of Progesterone in the Nervous System

A. Neuroprotective effects

Neuroprotective effects of progesterone have been demonstrated in different lesion models, notably in populations of neurons that are particularly sensitive to excitotoxic and ischemic damage. Such vulnerable neurons, which are generally characterized by high metabolic activity and abundant excitatory afferents, include the pyramidal neurons of hippocampus and cerebral cortex, dopaminergic neurons of the midbrain, Purkinje cells of the cerebellum, as well as neurons of the dorsal striatum and the caudate nucleus (93, 94). Thus, the administration of progesterone reduced the loss of neurons in the CA1 and CA2 subfields of the dorsal hippocampus and within the caudate nucleus after experimentally induced ischemia in cats (93, 95). In rats, progesterone given before middle cerebral artery occlusion (MCAO), decreased the infarct size and neurological deficits (96, 97). A recent study on functional outcomes after MCAO in male mice showed a beneficial effect of progesterone on survival rate, weight recovery, and motor ability evaluated by the grid and rotarod tests. Noteworthy, the spatial memory performance of the mice evaluated in the Morris water maze was also preserved by the progesterone treatment (98).

Beneficial effects of progesterone have also been demonstrated in experimental models of traumatic brain injury (TBI). A much-studied system corresponds to bilateral contusion lesion of the rat medial prefrontal cortex, which produces cognitive deficits typically observed after human frontal lobe injury (99, 100). The medial prefrontal cortex receives glutamatergic and cholinergic afferents, respectively, from the mediodorsal thalamic nucleus (MDN) and from the nucleus basalis magnocellularis (NBM). TBI leads to edema, to secondary excitotoxic neuronal death in the vicinity of the lesion, and subsequently to retrograde neuronal degeneration in both MDN and NBM (101). Edema is a very important negative factor for the outcome of TBI. Therefore, the observation that progesterone treatment reduced both edema and secondary neuronal losses and improved behavioral recovery after TBI in male rats was particularly encouraging. Females are protected by their high endogenous levels of pro-

gestosterone, and their brains have much less water content after TBI when compared with males (102, 103). Following these important observations, a phase II, randomized, double-blind, placebo-controlled trial, named “ProTECT”, has been conducted in Atlanta to test the usefulness of progesterone as a treatment for moderate to severe TBI. In this study, which included 100 trauma patients, stable progesterone levels were rapidly achieved after TBI by its iv infusion (104). The very promising outcome of the trial has now been published. Progesterone-treated patients had a lower 30-d mortality rate than controls, and survivors of moderate TBI who received progesterone had better outcomes. However, the administration of progesterone had no effect on the disability of severe TBI survivors. It is important to note that no adverse events could be attributed to progesterone in this trial (105).

What makes progesterone a particularly attractive neuroprotective agent for the treatment of brain lesions is its surprisingly large therapeutic window. Even when administered as late as 2 h after the onset of MCAO, progesterone still provided therapeutic benefit (106), and the steroid was effective in reducing edema and in protecting neurons after TBI when treatment was delayed as much as 24 h after injury (107). Pretreating ovariectomized female rats with low physiological concentrations of progesterone also allowed hippocampal neuron loss in response to TBI to be reduced (108). With respect to the duration of the progesterone treatment and its mode of administration, available experimental data show that both prolonged and continuous administration of the hormone leads to more complete behavioral recovery after TBI (109, 110).

An important finding was that administration of the enantiomer of progesterone (*ent*-progesterone) also decreased cerebral edema, neuron death, inflammatory cytokines, and reactive gliosis (111). Enantiomers of steroids indeed have a therapeutic potential for treating lesions and age-dependent dysfunctions of the nervous system (112, 113). An enantiomer is a mirror-symmetric, non-superimposable image of a molecule, with identical physical properties, except for the different rotation of polarized light, but with different biological actions (114). The protective effects of *ent*-progesterone were not mediated by the intracellular PR because the compound did not activate PR-mediated gene transcription, and its mechanisms of action, which may involve membrane receptors, need to be clarified. A previous study had shown that *ent*-progesterone is a potent competitive inhibitor of human enzymes involved in steroid metabolism, namely, the cytochromes P450c17 and P450c21 (115).

Neuroprotective effects of progesterone have also been demonstrated for midbrain dopaminergic neurons. Both progesterone and estradiol were found to protect dopaminergic neurons against degeneration induced by 1-methyl-4-phenyl-1,2,3,6 tetrahydropyridine, a finding consistent with a possible role of these hormones in the deterioration of dopaminergic functions with age and in the development of Parkinson’s disease (116). There is indeed a higher incidence of Parkinson’s disease in men when compared with women, and the risk of its occurrence is increased in women with an early onset of menopause (117).

In the spinal cord of male rats, chronic treatment with

progesterone for 5 d reduced the size of the lesion and prevented secondary neuronal loss after contusion injury (118). Progesterone is indeed an important neuroprotective factor for spinal motoneurons, as has been shown after spinal cord lesion and in a genetic model of motoneuron disease, the Wobbler mouse (11, 119, 120). After spinal cord transection, progesterone treatment preserved the Nissl bodies of the ventral horn motoneurons, restored choline acetyltransferase levels, normalized the expression of the Na,K-ATPase, and increased growth-associated protein of 43 kDa (GAP-43) and brain-derived neurotrophic factor (BDNF) message and protein (121, 122).

The Wobbler mouse is a particularly useful model for the study of motoneuron diseases, including amyotrophic lateral sclerosis. The Wobbler phenotype is due to a missense mutation in the gene encoding the vacuolar-vesicular protein sorting factor Vps54 (123). The first manifestations of the disease are already observed at 2–3 wk of age (124, 125). When 2-month-old, symptomatic Wobbler mice presenting tremor, ambulatory difficulty, and diminished muscle strength received sc implants of progesterone that produced constant high physiological levels of the hormone for only 2 wk, the neuropathological changes of spinal motoneurons were less severe, motoneuron vacuolation was reduced, and there was better preservation of the endoplasmic reticulum and of the mitochondria. Most importantly, progesterone treatment also had beneficial effects on muscle strength of the animals (126–128). The link between the mutant Vps54 and the severe impairment of Wobbler motoneurons needs to be explored, but it is likely that the mutation results in altered axonal transport. It was thus a significant finding that retrograde axonal transport is indeed impaired in Wobbler motoneurons, as shown by the injection of fluorogold into the limb muscles and its retrograde tracing, and that it can be restored by treating the animals for 8 wk with progesterone (119).

Only a few experimental studies have reported an absence of effect or negative actions of progesterone in the injured nervous system (attention has already been drawn in *Section III* to the disruptive effects of progestins such as MPA). Thus, one study did not find a beneficial effect of progesterone after ischemic insult in senescent female rats (129), and two dose-response studies have raised concerns about the possibility that high doses of progesterone (30–60 mg/d) may exacerbate the outcome of MCAO in ovariectomized female rats or of TBI in male rats (130, 131). The influence of sex hormones may also be dependent on the type and the severity of a brain lesion. Thus, sex differences in the outcome of TBI favoring female rats have been consistently observed after diffuse weight-drop-induced TBI, but these results could not be confirmed after more severe focal impact injury, qualified as “focal TBI” and characterized by a very fast evolution of neurodegeneration (132, 133). One study has reported that progesterone inhibited the neuroprotective effects of estradiol in the rat hippocampus observed after systemic kainate administration, which induces excitotoxic neuron death (134). Although very sparse, these negative results indicate that one should remain cautious and demonstrate the need for more research on the modes of action of progesterone and of its metabolites in the nervous system.

The mechanisms by which progesterone promotes morphological and functional recovery after brain injury are indeed not well understood, and they seem to involve multiple actions, some of which may be particularly relevant to the potential role of progesterone in the aging nervous system. In this regard, the above-mentioned observation that progesterone may improve impaired axonal transport is particularly interesting. There is indeed strong evidence that the slowing of axonal transport may be a key event in the aging process of the brain and of peripheral nerves (135–138). Reduced axonal transport has also been proposed to play an early and causative role in the development of Alzheimer's disease. That is, deficits in axonal transport are characteristic of the early stages of the disease in mouse models and in patients, and they may lead to aberrant amyloid- β peptide formation and subsequently to neurodegeneration (139).

Another important consequence of progesterone treatment is the reduction of lipid peroxidation, but the underlying mechanisms still need to be specified (140). The peroxidation of lipids is indeed a complex phenomenon, involving distinct enzymatic pathways as well as nonenzymatic mechanisms, such as free radical mediated peroxidation, and it is reduced by the actions of different antioxidant enzymes (141). During aging, there is an increase in the concentration of lipid peroxidation products, and the oxidative damage of lipids may play an important role in mediating or even initiating specific aspects of age-dependent changes (142). In addition, the oxidation of neuronal lipids resulting from an oxidative imbalance has been proposed to play a significant role in Alzheimer pathogenesis and, consequently, may represent an interesting therapeutic target at early stages of the disease (143). The increased exposure of aging tissues to oxidative stress partly results from decreased activity of antioxidant enzymes such as the superoxide dismutases. The prolonged treatment of middle-aged and old acyclic female rats (12, 18, and 24 months of age) with low doses of progesterone, estradiol, or a combination of both steroids increased superoxide dismutase activity and reduced lipid peroxidation (144, 145).

The age-dependent accumulation of oxidative damage is also a consequence of a decline in mitochondrial function, another major component of the normal aging process and of neurodegeneration (146–150). Many of the reactive oxygen species involved in oxidative stress are indeed toxic byproducts of the mitochondrial energy production pathway, and they damage not only lipids, but also proteins and nucleic acids. Over the past few years, attention has indeed focused on the role of the mitochondria in brain aging and neurodegeneration (151–153), and this cellular organelle is another important target for the actions of progesterone. One of the prevalent neuropathological changes found in spinal motoneurons of Wobbler mice is damaged mitochondria with severe vacuolation, and treatment with progesterone allowed the restoration of a normal appearance of the mitochondria (120). Progesterone can protect neurons against apoptotic cell death by increasing the expression of anti-apoptotic proteins residing in the outer mitochondrial membrane, such as Bcl-2, and by down-regulating proapoptotic gene expression (bax and bad) and the caspase-3 enzyme (73, 154, 155). The expression of Bcl-2-family proteins is regulated

by nuclear steroid actions, and the newly synthesized proteins are translocated to the outer mitochondrial membrane (156).

Some studies have suggested direct actions of steroids on mitochondria. Thus, estrogen and glucocorticoid receptors have been detected in mitochondria, and the mitochondrial genome contains nucleotide sequences with high similarity to known steroid-responsive elements (157–160). It has also been proposed that estradiol may protect against the formation of mitochondrial reactive oxygen species by directly acting on mitochondrial estrogen receptors (ERs) (161). Whether progesterone may also exert direct effects on mitochondria remains to be explored. In the low physiological range, progesterone has been shown to completely reverse postinjury alterations in mitochondrial respiration (108). The mitochondria is also a target for other steroids, such as dehydroepiandrosterone (DHEA), and it is the site where the first step in steroid hormone biosynthesis takes place, the conversion of cholesterol to PREG by the cytochrome P450_{scc}, as shall be discussed later. There obviously exist complex relationships between steroids, mitochondrial activity, oxidative stress, the aging of brain cells, and neurodegenerative events. Unfortunately, the available data are still much too fragmentary for an integrated picture.

Progesterone exerts many other actions, which can be related to its neuroprotective effects and may also have implications for the aging brain. Thus, progesterone regulates the expression of aquaporin 4 in the injured brain, a membrane-channel protein involved in water homeostasis, which is largely distributed throughout the brain and may play a significant role during edema formation (162). It is puzzling that some of the actions of progesterone resemble those of the estrogens. For example, like estradiol, progesterone up-regulates the expression of antiapoptotic proteins such as Bcl-2 (163–165), reduces inflammation by repressing the activation of microglial cells and by inhibiting the production of proinflammatory cytokines (101, 166–168), up-regulates the expression of neurotrophins such as BDNF (122, 169, 170), and protects neurons against glucose deprivation and the toxicity of glutamate, FeSO₄, and β -amyloid peptides (72, 171, 172).

B. Promyelinating effects

Progesterone is also known to have a role in myelination and remyelination. By preserving or restoring the integrity of myelin sheaths, which insulate the large axons and are required for the efficient and rapid conduction of electrical impulses along nerve fibers, progesterone may not only play an important role in the efficient communication between neurons but also promote their viability. In fact, myelin has neuroprotective functions, and myelin-associated proteins influence the caliber of axons (173). Moreover, after injury, secondary demyelination of spared axons contributes to neuronal degeneration, the extension of the lesion, and functional deterioration (174).

A role of progesterone in myelination was first demonstrated in the regenerating mouse sciatic nerve after lesion as well as in explant cultures of rat dorsal root ganglia composed of sensory neurons and Schwann cells, which are the myelinating glial cells of the PNS (175). Progesterone also

enhanced the rate of myelin formation in dissociated cocultures of neurons and Schwann cells (176). Whether progesterone promotes myelination of peripheral nerves directly by acting on Schwann cells, or indirectly by acting on neurons, needs to be clarified because there are contradictory reports. Whole-cell radioligand binding assays suggested the presence of specific and saturable progesterone binding sites in Schwann cells (177). The PR has also been detected in Schwann cells either grown in culture or within the rat sciatic nerve by immunocytochemistry (178). On the contrary, in cocultures of dorsal root ganglia neurons and Schwann cells, the presence of PR mRNA and protein was only detected in the neurons, not in the Schwann cells (179). Also, in a recent study, purified rat Schwann cells and various Schwann cell lines were found to only express extremely low amounts of PR mRNA (180). In the mouse Schwann cell line MSC80, progesterone did not activate the transcription of a progesterone-sensitive reporter gene. Demonstration that the PR was the limiting factor was provided by the positive transcriptional responses obtained when exogenous receptors were transiently expressed (180). Whether present in neurons or glial cells, the PR in the peripheral nervous system is a potential pharmacological target for the therapy of inherited and acquired peripheral neuropathies. In a transgenic rat model of Charcot-Marie-Tooth disease overexpressing the peripheral myelin protein PMP22, the culprit of the disease, treatment with a progesterone antagonist reduced PMP22 expression and had a beneficial influence on the evolution of the disease (181). In a model of diabetic neuropathy, induced in rats by an injection of streptozotocin, prolonged treatment with progesterone or its reduced metabolites had beneficial effects on peripheral nerves at the neurophysiological, functional, and neuropathological levels (182).

In the CNS, brain, and spinal cord, axons are myelinated by oligodendrocytes (183, 184). That progesterone also promotes myelination by oligodendrocytes has been demonstrated in explant cultures of cerebellar slices taken from 7-d-old rats and mice (185). These organotypic cultures closely reproduce developmental events and provide a unique model for examining neuronal survival and maturation, as well as the myelination of axons (186, 187). In these explants, myelination is very intense during the second postnatal week, exactly at a time when endogenous levels of progesterone are elevated in the cerebellum (188, 189). A stimulatory effect of progesterone on myelination was observed in cerebellar slices of both sexes and involved the classical PR: 1) it could be mimicked by the selective progestin promegestone; 2) it was completely abolished by the PR antagonist mifepristone (RU486); and 3) it was not observed in cerebellar slice cultures from 7-d-old PR knockout mice (185). In these slices, progesterone was shown to stimulate the proliferation and maturation of oligodendrocyte progenitor cells (190). An earlier study had already shown that adding progesterone to cultures of glial cells isolated from neonatal rat brains increased the number of oligodendrocytes (191). More recently, the addition of progesterone to the medium of cultured oligodendrocytes has been shown to increase their branching, whereas estradiol stimulated myelin membrane formation (192). Progesterone also promotes remyelination by oligodendrocytes *in vivo*. After toxin-induced demyelination, the sys-

tematic administration of progesterone promoted the slow endogenous remyelination of axons within the cerebellar peduncle of aging male rats (193). In the lesioned rat spinal cord, treatment with progesterone was found to increase the density of NG2⁺ oligodendrocyte progenitor cells and the expression of myelin basic protein (194).

The promyelinating effects of progesterone are particularly relevant when discussing the significance of the hormone in the aging nervous system. Indeed, it is less well appreciated that overall loss of myelin and altered integrity of myelin sheaths are among the most reliable markers of the aging nervous system, correlating with chronological age and cognitive decline (195, 196). Age-dependent changes in brain myelin, which have been extensively studied in rhesus monkeys, include alterations in oligodendrocytes, abnormalities and breakdown of the myelin sheaths, and loss of white matter (195, 197–199). Based on these observations, it has been proposed that myelin changes may significantly contribute to age-related cognitive decline by altering conduction velocities along axons (196). A postmortem study has provided evidence that aging in humans is also accompanied by the loss of myelinated fibers (200). By using the method of diffusion tensor magnetic resonance imaging (MRI), it has been shown that myelin disruption occurs in men even during normal aging. Most importantly, alterations of cortical myelin correlated with declined cognitive ability (201). In parallel with these morphological and functional studies, only a few reports have dealt with biochemical changes in myelin during aging. Increase in water and decrease in cholesterol within white matter have been described (202), and decreased expression of the major peripheral myelin proteins has also been reported in peripheral nerves of aged rodents and humans (203–206).

Importantly, remyelination continues to take place in the brains of aged monkeys, but the newly formed myelin sheaths are thin and have short internodes (207). Age is indeed a negative factor for the capacity to regenerate myelin sheaths, as has been demonstrated in the rodent CNS after demyelination induced by a gliotoxin; in old rats, the process of remyelination takes much longer than in young animals (208–210). The reasons for the age-associated slowing down of myelin repair are not well understood, but impaired recruitment of progenitor cells and their delayed differentiation into myelinating oligodendrocytes, as well as delayed expression of growth factors, may be responsible (211, 212). In humans, differences in the speed of remyelination could explain the much slower functional recovery in older patients after demyelinating diseases such as optic neuritis (213). In addition, a reduced capacity for myelin repair with age is consistent with the observation that the prognosis of multiple sclerosis is mainly age-dependent (214).

In conclusion, a substantial number of animal studies have documented neuroprotective effects of progesterone or its reduced metabolites in the lesioned or diseased nervous system of young adult rodents. Particularly promising for the treatment of traumatic lesions is the large therapeutic window of progesterone. Progesterone may exert neuroprotective effects and promote neuroregeneration by a dual action: by directly acting on neurons and increasing their survival, and by accelerating the formation of new myelin sheaths.

Progesterone and its metabolites may exert similar beneficial effects in the aging brain and peripheral nerves. The significance of progesterone in aging will be further explored when discussing the question of whether the aging nervous system remains sensitive to the beneficial effects of steroids.

V. Neurons and Glial Cells in the Aging Nervous System

A. Aging neurons

Early studies describing massive loss of neurons during nonpathological aging of the brain have been largely refuted by the use of more accurate stereological techniques for the precise counting of cells in histological sections (215). There is indeed no extensive loss of neurons during aging, as previously thought, even within vulnerable brain regions such as the cerebral cortex and the hippocampus (216). Stereological studies have also shown that there is no significant loss of neurons within the hypothalamic nuclei involved in the control of reproductive functions in older women, but rather a substantial remodeling of neuronal circuits and changes in neuropeptide expression (217, 218). Consequently, normal age-associated neuronal impairment is more likely to be mediated by synaptic alterations, which may be reversible, making the treatment of age-related dysfunctions of the brain a therapeutic possibility (219). Even in old rats with impaired spatial learning, no significant neuron loss was observed within the hippocampus (220). Also in patients with Alzheimer's disease, the loss of forebrain cholinergic neurons may not be as important as previously thought. Indeed, within the NBM, only a small subset of the neurons was found to die, but the large cholinergic neurons underwent atrophy and lost their markers (221). Neuronal death is indeed a relatively late stage event in Alzheimer's disease associated with dementia, and alteration of synapses is one of the early pathogenic processes (222–224).

However, brain structures may differ in the involvement of neuron loss, and some populations of neurons may be more affected by the aging process than others. For example, within subregions of the rat hippocampal formation, the number of neurons may significantly decrease at advanced ages, in particular within the subiculum and the hilus of the dentate gyrus (225). Among the most vulnerable cells of the nervous system are the cerebellar Purkinje cells, and there is consistent evidence for their significant loss during normal aging in rodents and humans. The age-dependent loss of Purkinje cells correlates with decreased eye-blink conditioning, a reflex pathway mediated by these neurons, and elderly people with very slow eye-blink conditioning may have an increased risk of becoming demented (226, 227).

B. Aging glial cells

Neurons have long been the main focus of studies on brain aging, and glial changes have been largely neglected. As already pointed out, there are significant deteriorations of the myelin sheaths with age, which may reflect age-dependent changes in the myelinating glial cells, oligodendrocytes in the CNS and Schwann cells in the PNS. However, the for-

mation and maintenance of myelin sheaths are also dependent on neuronal signals, and there are complex reciprocal interactions between axons and myelinating glial cells in both compartments of the nervous system (228–231). Consequently, any age-dependent alterations of myelin sheaths may result from impaired neuronal functions, from changes in the myelinating glial cells themselves, or from both events.

Another type of glial cell also plays an essential role in brain aging. The general assumption was that the increased number of astrocytes (astrogliosis) during aging may be a consequence of neuron degeneration. However, there is now strong experimental evidence provided by Finch and collaborators (232) that changes in astrocytes are in fact a very early event in the aging process, and that increased glial fibrillary acidic protein (GFAP) expression by astrocytes may contribute to decreased synaptic functioning and plasticity in the aged brain. Indeed, in cocultures of neurons and old astrocytes, diminishing GFAP levels by RNA interference restored neurite outgrowth, whereas overexpression of GFAP in young astrocytes modeled the effects of aging by reducing neurite outgrowth (233). Consistent with these *in vitro* findings is the observation that inactivation of the GFAP gene in mice improves both neuronal survival and neurite growth (234, 235).

Important for the present discussion is the observation that astrocytes can mediate some of the effects of progesterone and estradiol on neuronal plasticity, and that steroids are a critical component of the cross-talk between neurons and glial cells (236, 237). Thus, enhanced neuronal sprouting after lesion in response to estradiol is mediated in part by the repression of GFAP expression in astrocytes (238). Astrocytes are also a target for the actions of progesterone; after a penetrating brain injury, treatment with progesterone decreased astrocyte accumulation in both female and male rats (239, 240). Progesterone was also shown to reduce astrocytic hypertrophy after TBI close to the lesion site (154). However, in two other models, progesterone was not found to modify astrocyte accumulation in rats, either after spinal cord transection or after medial frontal cortex contusion (241, 242).

VI. Gender Differences and Sensitivity to Progesterone

When studying the effects of progesterone on the nervous system, it is important to be vigilant about the possible contribution of structural and biochemical sex differences (243, 244). There is indeed increasing recognition that gender differences may influence the incidence and development of diseases and the responses to therapies (245). Accordingly, the effects of steroids may also differ between females and males, and data obtained for one gender may not necessarily apply to the other. Since the pioneering studies of Raisman and Field (246), important differences in brain structure between males and females have been widely recognized. In rodents, they arise in part through the permanent "organizational effects" of androgens secreted by the testis during sensitive periods in early life (247–249). Sex differences affecting brain structures and functions, including its asymmetry and functional lateralization, are also observed in hu-

mans (250–253). In addition, MRI analysis has revealed that age-specific changes within the human brain are also sexually differentiated (254). The sexual phenotype of brain cells is not determined exclusively by the exposure to steroid hormones during early development, but also by their genetic sex because some sex differences are already established before the maturation of the embryonic gonads (255, 256). More recently, the development of transgenic mouse models has allowed it to be shown that XX and XY brain cells are not equivalent, even when they have developed in a similar hormonal environment (257, 258).

Only a few studies have investigated the possible influence of gender on the response of the adult nervous system to the trophic and protective effects of ovarian steroids, and there are surprisingly few observations of sex differences. A recent study has revealed that the effects of progesterone and its 5α -reduced metabolites on the expression of peripheral myelin protein genes differs between males and females. This was shown by using sex-specific cultures of Schwann cells prepared from neonatal rats (259). On the other hand, the differential sensitivity of the male and female rodent brain to injury appears to be largely determined by the presence of different levels of progesterone. Thus, the more favorable outcome after cerebral stroke or TBI in female rats when compared with males mainly results from the presence of high endogenous levels of progesterone in the females. Similarly, treatment with progesterone provides similar neuroprotection in males and in females after TBI (102, 260).

It is worth mentioning here that a few studies have reported that some brain responses to estrogen are sexually dimorphic. In one study, estradiol was found to improve neurological outcome after TBI in male rats, but to exacerbate brain injury in females (261). In another study, although estrogen therapy protected both male and female brains against ischemic insult, the responses differed between sexes: acute exposure to estrogen was sufficient to ameliorate ischemic brain injury in males, whereas females required longer-term replacement (262). As will be discussed later, the response of aged hippocampal synapses to estrogen is also sexually dimorphic (263).

Very few studies have addressed the question of the effects of gender on the outcome of injury in the human nervous system, and in TBI patients the role of gender is still controversial. One clinical study has reported that female TBI patients have a better outcome than male patients (264). More recently, another group has shown that female patients have lower levels of cerebrospinal fluid (CSF) lipid peroxidation and oxidative damage products, consistent with the already discussed antioxidant properties of ovarian hormones (265, 266). However, other studies have not shown a beneficial effect of female gender on TBI outcome (267, 268).

Gender is also an influential factor in the incidence and progression of multiple sclerosis, a demyelinating disease that selectively affects the brain and spinal cord (214, 269–271). The questions of why more women have multiple sclerosis than men and why it affects women differently from men have been mainly addressed experimentally by examining hormonal influences on autoimmune responses (269, 272, 273). However, the recent observation showing that myelin is sexually dimorphic casts a new light on the role of

gender and hormones in the maintenance, alterations, and diseases of myelin (274). In this study, gender differences in myelin components of white matter tracts of young and aged rodents were found to be so dramatic that it was possible to determine the sex of an animal from blind sections immunostained for oligodendrocyte-specific markers. Gonad-derived steroids appeared to be a major contributor to these sex differences because castration of adult males produced a female phenotype (274). Consistent with clinical observations showing that the course of multiple sclerosis is mainly age-dependent, and that women reach disability milestones at older ages than males (214, 275), the extent of oligodendrocyte remyelination after a demyelinating lesion was found to be significantly reduced in aging rats, and middle-aged males and females (12 months of age) differed in their capacity to remyelinate axons. This sex difference was not influenced by castration, suggesting a more stable sex difference (210).

VII. The Sensitivity of the Aging Nervous System to Progesterone and Estradiol

Two fundamental questions need to be addressed when discussing the usefulness of HRT: 1) does the aging nervous system remain sensitive to the actions of ovarian hormones; and 2) do these hormones continue to exert beneficial effects on the aging nervous system? Indeed, the majority of studies documenting beneficial effects of ovarian steroids have been carried out in young adult animals or in cultured cells isolated from embryonic or neonatal tissues. Data on aged animals or cells are rare, and only a few laboratories have examined the question as to whether responses of target tissues to steroids are preserved during the aging process. There is as yet no conclusive answer to this question, and the extent to which mechanisms of neuroprotection are similar in young adults and reproductively senescent animals remains to be clarified. There is however some experimental evidence that the aging nervous system remains, at least to some extent, sensitive to ovarian steroids, and that their administration may even allow reversal of some of the age-dependent structural abnormalities and dysfunctions. On the other hand, there are also indications that some responses of neural cells to hormones may change, even during the normal aging process. Such changes in hormone effects appear to be dependent on the steroid, brain region, and nervous function examined. The reader will notice that in this section explicit reference will also be made to the effects of estradiol. Indeed, a large number of the studies concerning steroid sensitivity of the aged nervous system has tested the effects of estradiol, and some of them are quoted here to exemplify the problem.

A. Maintained sensitivity to ovarian steroids

1. *Progesterone.* Beneficial effects of progesterone on the aging nervous system have been particularly well demonstrated for myelinated nerve fibers. In *Section IV.B*, progesterone plays an important role in peripheral nerve myelination, and recent studies have shown that treatment with progesterone or its 5α -reduced metabolites allows reversal of age-related

myelin abnormalities. Thus, in the sciatic nerves of aged male rats, a significant decrease in myelin-associated activity of the 5 α -reductase, the enzyme that converts progesterone to 5 α -dihydroprogesterone (or testosterone to 5 α -dihydrotestosterone), is associated with a reduction in myelin gene expression. Treatment of the aged rats for 1 month with progesterone, 5 α -dihydroprogesterone, or allopregnanolone allowed reversal of the age-dependent decline in peripheral myelin protein expression, whereas repeated injections of androgens were without effect (204, 205). The administration of progesterone not only counteracted the drop in myelin protein expression, it also allowed reversal of age-related structural abnormalities of the peripheral myelin sheaths. Indeed, the prolonged treatment of old male rats (22–24 months) with progesterone or its 5 α -reduced metabolites significantly decreased the percentage of fibers with myelin abnormalities as well as the number of fibers with irregular shapes, and it increased the number of small myelinated fibers. Again, as previously observed for the normalization of myelin gene expression, the effects were specific for progesterone and its metabolites because the administration of androgens was inefficient (276).

As already mentioned, the capacity to repair myelin in the brain decreases with age; spontaneous remyelination after gliotoxin-induced demyelination is very rapid in the brain of young rats, but it is very much delayed in middle-aged rats. Whereas no beneficial effect of progesterone on central myelin repair could be observed in young males (10 wk old), because spontaneous remyelination was too rapid, the implantation of sc progesterone pellets stimulated a slow remyelination of axons in middle-aged animals (9 months old) (193, 277).

It has been proposed that the disappearance of the protection against ischemic brain injury in females after reproductive senescence may be a consequence of ovarian hormone deficiency. However, aging female rats remain responsive to the protective actions of ovarian hormones, at least until a certain age. Thus, in middle-aged female rats (16 months), the administration of either progesterone or estradiol alleviated cerebral stroke (278).

Animal models also support the anxiolytic effects of progesterone, which are mediated by its conversion to allopregnanolone, a potent positive modulator of GABA_A receptors (see Section X.B) (279–281). In fact, the anxiolytic actions of progesterone do not require the intracellular PR because as they are still observed in PR knockout mice, which even exhibit a greater anxiolytic response than their wild-type littermates (282). However, progesterone does not enhance anxiolytic behavior in mice deficient of the type 5 α -reductase (283). Most importantly, middle-aged (between 9 and 12 months of age) and old (between 18 and 24 months of age) wild-type and PR knockout mice continue to respond to the anxiety-reducing effects of progesterone (284). That the brain of senescent mice continues to be responsive to progesterone and its metabolites was also demonstrated by the results of another study showing that middle-aged and old female mice primed with estradiol show lordosis behavior after the intraventricular injection of progesterone or allopregnanolone (285). Lordosis is a stereotypic posture adopted by

a sexually receptive female rodent in response to a mount by a male.

Another consequence of ovarian hormone deficiency is a decline in cognitive performance. Data concerning the effects of steroids on cognition, and in particular on memory, need to be interpreted cautiously because they are not always consistent. This may reflect the fact that complex behaviors are under the influence of multiple factors, and that behavioral testing procedures differ between laboratories. In particular, the behavioral effects of progestagens can be difficult to interpret, because progesterone and its metabolites can influence cognitive processes by multiple mechanisms. Thus, the rapid neuromodulatory effects of progesterone and allopregnanolone are expected to transiently impair memory performance: 1) progesterone behaves as an antagonist of σ 1 receptors and inhibits the promiscuous effects of σ 1 agonists (286); 2) allopregnanolone is a positive modulator of GABA_A receptors, involved in the inhibition of memory processes within the hippocampus and at the level of cholinergic forebrain neurons (287–291) (see Section X.B). However, under stressful conditions, the effects of allopregnanolone may be beneficial for memory tasks because of its anxiolytic properties (292, 293). Moreover, although the immediate effect of progestagens may be a diminution of memory functions, because of the rapid modulation of neurotransmitter receptors, in the long run, they may have beneficial effects on cognitive performances because of their trophic and protective actions (294). Two studies have reported an influence of progesterone on cognition in aged female rats. In females ovariectomized at the age of 13 months, the prolonged weekly administration of estradiol and progesterone was found to be slightly more efficient in enhancing acquisition of a spatial memory task than treatment with estradiol alone (295). However, two other studies have reported negative effects of progesterone on cognition and working memory in aged female rats (296, 297).

2. *Estradiol.* As already mentioned, the administration of either progesterone or estradiol improved the outcome of cerebral stroke in middle-aged female rats (278). In both young (3–4 months) and middle-aged (9–12 months) female rats, replacement with physiological doses of estradiol decreased the extent of ischemic injury in the cerebral cortex (165, 298). It is important to note that these protective effects of the ovarian steroids were not mediated by changes in cerebral blood flow. In another study, the pretreatment of reproductively senescent, female rats (14–18 months) with estradiol alone or combined estradiol plus progesterone also reduced cortical infarct volume after MCAO (129).

Memory performance can also be restored in senescent female rats by estradiol if administered in an appropriate manner, even at an advanced age and after long-term hormone deficiency (299). Consistent with these behavioral findings, cholinergic pathways involved in memory processes can still be activated by estrogen in the brain of aged female rats (24 months) (300). In middle-aged nonhuman primates, multiple cognitive functions also remain sensitive to estrogen (301). Thus, in perimenopausal rhesus monkeys around 22 yr of age, which is at a time when cognitive functions start to decline, estrogen treatment improved memory (302). Even

after many years (up to 16) of estrogen deprivation, estradiol was still able to enhance some aspects of working memory in aged female rhesus monkeys (21–24 yr) (303, 304).

B. Modified sensitivity to ovarian steroids

1. *Progesterone.* In comparison with the estrogens, examples of age-related changes in the sensitivity to progestagens are sparse. This does not necessarily mean that the aging brain remains more sensitive to progesterone than to estradiol, although this a possibility, but it may simply reflect the fact that fewer studies have been devoted to the effects of progestagens. Among the few studies that have reported a loss in the sensitivity of the aging brain to progesterone, two have shown that acute administration of progesterone either before or after MCAO reduced cortical infarct in young, but not in reproductively senescent female rats (14–18 months old) (129, 305). It has been proposed that changes in the tissue metabolism of progesterone may be one means by which the effectiveness of progesterone decreases during aging (306).

2. *The hypothalamic-pituitary-ovarian axis.* An age-dependent reduction in the sensitivity to ovarian steroids has been well documented for hypothalamic nuclei involved in reproductive functions. The hypothalamic-pituitary-ovarian axis does indeed become less responsive to the positive feedback effects of ovarian hormones in middle-aged females, but the underlying mechanisms are only partly understood (307, 308). One possibility is that the hypothalamus does not respond to steroids in the same way in young and old females because of changes in the expression or functionality of steroid receptors, but discrepancies between studies make a general conclusion difficult. At least, there appears to be no global and marked decline in brain PR and ER with age (309–313). A recent stereological analysis has even reported an increase in ER α -immunoreactive neurons within specific hypothalamic nuclei of aged females (between 12 and 24 months of age) (314).

Within the hypothalamus, PRs involved in the regulation of reproductive functions are induced by estradiol (315, 316). Again, the ability of estradiol to induce hypothalamic PR seems not to be attenuated with age (309, 317). In a recent study, the induction of PR mRNA by estradiol within distinct nuclei of the hypothalamus was found to be at least as strong in 15-month-old as in young 3-month-old female rats (310). Obviously, decreased hypothalamic ER and PR expression does not provide a satisfactory explanation for reduced steroid sensitivity of the hypothalamus and reproductive senescence in female rats. However, binding and expression studies do not reveal very much about the functionality of steroid receptors. As will be discussed in *Section X.A*, work over the past 10 yr has revealed that the transcriptional efficiency of liganded steroid receptors is determined by nuclear coregulator proteins (318, 319). So far, there is little information concerning the effects of aging on the recruitment of coregulators by steroid receptors, and this is certainly a line of research worth exploring. One study has reported that expression of two coactivators, namely, steroid receptor coactivator-1 (SRC-1) and CREB-binding protein (CBP) are decreased in androgen-sensitive spinal motoneu-

rons of aged rats (320). Another study has shown that low levels of SRC-1 within the olfactory bulb and forebrain of reproductively senescent female rats correlated with the reduced sensitivity of neurotrophins and their receptors to estrogens (321).

Other explanations than reduced steroid receptor expression have been proposed for the decreased ability of estradiol to generate a LH surge in aged females, such as alterations in circadian rhythmicity of the suprachiasmatic nucleus (308, 322). The age-dependent loss of astrocyte plasticity in the rostral preoptic area, where a subgroup of GnRH neurons resides, may also have an impact on the ability of estrogens to activate GnRH neurons because dynamic changes in the coverage of neurons by astrocytes play a key role in synaptic plasticity and efficacy (323, 324).

3. *Estradiol.* There is a series of studies which document that the response to estradiol of brain regions other than the hypothalamus is modified during the aging process. Within the medial septum and NBM, levels of choline acetyltransferase and of nerve growth factor receptor TrkA mRNA were substantially reduced in old long-term ovariectomized female rats (ovariectomy at the age of 13 months, histological analysis of the brains around 30 months of age). These changes could not be reversed by estradiol plus progesterone replacement, despite the fact that the cholinergic neurons were still present and that their number and size were apparently not affected (325). Other studies have provided evidence that the ability of estradiol to enhance cholinergic and cognitive functions declines with age in rats (299, 326). Thus, estrogen replacement can reduce memory deficits induced by the muscarinic receptor antagonist scopolamine in young and in middle-aged female rats with irregular cyclicity (12–13 months of age), but not at a more advanced age characterized by consistent estrus (20 months of age) (326). It has also been reported that estradiol replacement is not effective in improving working memory performance in female rats after long-term hormone deprivation by ovariectomy. However, when initiated immediately after ovariectomy, estradiol replacement significantly improved memory, even at 17 months of age (327). Based on these experimental findings, the existence of a “window of opportunity” after the loss of ovarian function has been proposed, during which hormone treatments may be most efficient for preventing a decline in cognitive functions (295). This hypothesis will be more closely addressed in *Section VIII*.

In the rodent hippocampus, estradiol regulates multiple aspects of synaptic plasticity involved in memory processes, such as an increase in the number of dendritic spines bearing excitatory *N*-methyl-D-aspartate (NMDA) synapses (328, 329). With age, there is a loss of CA1 synapses, which is not reversible by estrogen replacement: in contrast to young female rats (3–4 months old), estradiol failed to increase the number of dendritic spines in old females (23–24 months old) (330). However, although estradiol did not increase the number of dendritic spines in the hippocampus of aged female rats, it increased the number of NMDA receptors per synapse, which may be understood as a compensatory response (331, 332). It would be interesting to examine whether there is a relation between the increase in synaptic NMDA recep-

tors and the well-known increase in vulnerability of the aged hippocampus to excitotoxicity (333). In female rhesus monkeys, the number of dendritic spines on CA1 pyramidal neurons is also highly responsive to estrogen, but unlike aged female rats, hippocampal neurons of aged female rhesus monkeys (19–23 yr old) retain their capacity for spine induction in response to estrogen (334). Estradiol treatment also increased the density of dendritic spines of pyramidal neurons within the dorsolateral prefrontal cortex of both young and aged female rhesus monkeys, thus demonstrating a maintained responsiveness to estradiol and capacity for plasticity (5, 335).

Interestingly, the effects of steroids on the plasticity of aged synapses can differ between sexes, as has been reported for granule neurons of the dentate gyrus. Male and female rats were gonadectomized at 2 months of age. When examined at 16–20 months of age, the old females had a paucity of dendritic spines on granule neurons, but males showed no decrease in dendritic spines with age. A short-term treatment with estradiol allowed the density of dendritic spines in the old females to be increased, but on the contrary, decreased spine density in males (263).

Another example of reduced sensitivity of the aging nervous system to estrogens is the decrease in their ability to attenuate neuronal damage in response to unilateral entorhinal cortex lesion (336). After this type of lesion, which is used as a model of Alzheimer's disease-like deafferentation of the dentate gyrus, estradiol stimulated compensatory synaptic sprouting in young (3 months old), but not in middle-aged (18 months old) female rats (337). In the same study, it was shown that increased GFAP expression by hippocampal astrocytes in response to the lesion was reduced by estradiol in the young, but not in the middle-aged females. As already discussed, elevated GFAP expression in astrocytes contributes to the reduction in the neuronal sprouting response during aging (233). The increased expression of GFAP in astrocytes, which is progressive and begins before midlife, may thus play an important role in neuron atrophy and impaired synaptic plasticity (233).

Age-dependent impairment has also been documented for the regulation of neurotrophin signaling by estrogen. Estradiol increased the expression of BDNF and of the neurotrophin receptors TrkA and TrkB within distinct brain regions in young adult, but not in reproductively senescent female rats (321). However, another study has reported a comparable increase in TrkA mRNA expression by estrogen in the NBM of young (3 months old) and old (24 months old) rats, suggesting that the sensitivity of neurotrophin signaling to estradiol may be preserved in the aged brain (300).

C. Antagonistic pleiotropy

Some nervous functions may not only lose their sensitivity to steroids with progressing age, but may even become negatively affected by them. According to the “antagonistic pleiotropy” theory of senescence formulated by Williams in 1957, some genes with positive effects upon fitness early in life may become deleterious late in life. Antagonistic pleiotropic effects have been documented for various classes of genes, including those encoding growth factors, hormones,

heat shock proteins, and apoptosis regulator proteins (338, 339).

With respect to steroid effects on the nervous system, this concept has only received support from a limited number of experimental and observational studies on estrogens (2), and there is no example of adverse effects of progestagens on the aging nervous system. Whether this situation is due to the smaller number of studies with progesterone or whether it reflects a better toleration of progesterone by the aged brain needs to be clarified. Indeed, in *Section VII.A*, the nervous system of even very old rodents remains surprisingly responsive to the beneficial effects of progesterone.

In contrast to young female rats, the treatment of reproductively senescent females with estradiol exacerbated neural injury and worsened the inflammatory response within forebrain circuits after an excitotoxic lesion (340). Estradiol, known to attenuate cytokine responses, was also shown to increase inflammatory cytokine expression by immune cells in acyclic rats (341). Similarly, effects of estradiol on neurotrophin expression can be opposite in young and aged females. Thus, estradiol increased BDNF expression in young female rats but decreased it in reproductively senescent females (321). The same group has also recently shown that the effects of estradiol on the blood-brain barrier are also dependent on age. Estrogen replacement to surgically castrated young female rodents reduced the permeability of the blood-brain barrier, but conversely made it more permissive in senescent females (342). Together, these findings point to the risk of estrogen replacement in the elderly. Within this line of thinking, an extreme view proposes that decreased estrogen levels may even become beneficial for some populations of neurons in the aged brain (221).

Concerning the cardiovascular system, ovarian steroids may become dangerous by inducing inflammatory responses and by having prothrombotic effects once atheromas and narrowing of blood vessels are established. This view is supported by mouse models of atherosclerosis, which have revealed that estradiol has an atheroprotective effect by acting on healthy endothelium, but induces inflammatory-immune responses once atheromatous plaques are formed (343). In monkeys, estradiol has been shown to inhibit atherosclerotic plaque formation when given directly after ovariectomy, but not 2 yr later (344).

In conclusion, the results of animal studies are rather encouraging because they show that some structures and functions of the aging nervous system remain sensitive to the beneficial actions of ovarian steroids, and in particular to those of progesterone, providing an experimental ground for the usefulness of HRT. Neuronal networks appear to remain in place during the aging process, and age-dependent structural changes and dysfunctions can be improved by the administration of ovarian steroids. However, there is also evidence that some responses to steroids are substantially modified with progressing age, and that aging and long-term deprivation of ovarian steroids may result in the insensitivity of specific brain functions to either progesterone or estradiol. Moreover, there is some concern about possible deleterious effects of estradiol on the brain and blood vessels in aged animals.

VIII. The Timing of HRT: A Therapeutic Window?

There may be therapeutic windows during which HRT is particularly efficient and during which the beneficial effects of hormones may prevail (345). Arguments in favor of this hypothesis have mainly been provided by the outcomes of estrogen-only or combined estrogen-progestin therapies. As in rodents, brain regions involved in the regulation of ovarian activity may also become less sensitive to estradiol during aging in women. Thus, data from the Women's Health Across the Nation (SWAN) study, a survey of women going through the menopause transition, suggest that secretion patterns of estradiol and LH in middle-aged women may reflect an increasing insensitivity to estradiol (346). In premenopausal women, estrogen levels are equivalent or higher than those observed in younger women, but conversely, LH pulse frequency is decreased, consistent with an altered positive estrogen feedback in the brain (347–349).

In particular, at very advanced ages or after very long hormone deprivation, the steroid responsiveness of specific neuronal circuits and cognitive functions may be altered in women. For them, the time when HRT starts relative to the onset of reproductive senescence may be a crucial factor. An accumulating body of evidence indeed suggests that the immediate postmenopausal period may constitute a window of opportunity for HRT to protect against cognitive decline and to reduce the risk of Alzheimer's disease (350, 351). In fact, a major difference between studies that found a protective effect of estrogens on cognitive functions and those that reported negative results was the time when the treatment started. This would be consistent with results of a series of animal studies suggesting that estrogens may be more efficient in preserving memory functions if treatment is started soon after the deprivation of ovarian steroids (304, 352, 353).

A large prospective trial conducted in Utah, named the Cache County Study, has examined the incidence of dementia among 1889 women (mean age, 74 yr) and 1357 men (mean age, 73 yr). Results for women showed that when HRT was initiated at the beginning of the menopause and lasted for at least 10 yr, the incidence of Alzheimer's disease was significantly decreased, and the age-dependent increase in the risk of Alzheimer's disease was abolished. On the contrary, in women who began HRT 10 yr or later after the onset of menopause, there was a tendency for an increased risk in developing Alzheimer's disease (354). The latter observation is thus consistent with the findings of the WHIM showing that HRT, initiated as late as 10 to 20 yr after the onset of menopause, increased the incidence of dementia and Alzheimer's disease (32, 355). Results of another recent study, involving a small number of postmenopausal women, confirm that women with few years since the onset of menopause benefit more from HRT with respect to cognitive functions than women starting later (356). The follow-up of another cohort of 343 women has shown that short-term HRT in the early phase of the menopause and lasting for only 2–3 yr may provide long-term protection against cognitive impairment, still observable 15 yr later (357). All these data are consistent with the view that initiation of HRT early in menopause may have cognitive benefits and reduce the risk of

dementia, whereas hormone therapy initiated decades after the onset of hormone deficiency may be without benefit, and may even become unsafe.

It is noteworthy that a similar conclusion has been reached by examining steroid effects on the cardiovascular system. The time when HRT is started relative to the onset of menopause seems indeed to be crucial for its effectiveness on the vascular system (358, 359), and the effects of estrogens and progestagens on the vasculature may also depend on the stage of existing atherosclerosis (360). Moreover, longer exposure to endogenous ovarian hormones as a result of delayed onset of menopause may be protective: for each year's delay in the onset of menopause, the cardiovascular mortality risk was shown to decrease by 2% (358, 359). Thus, duration may also be a critical factor for HRT. In the Rancho Bernardo cohort, women who had used HRT for at least 10 yr had significantly less plaque burden than shorter term users (361). The analysis of data of the Nurses Health Study, which has followed 120,000 female nurses between 1976 and 2000, also suggested that starting HRT earlier may indeed make the difference; women who began HRT near menopause had a significantly reduced risk of coronary heart disease. On the contrary, no significant relation was found between HRT and coronary heart disease among women who initiated therapy at least 10 yr after menopause (362). A recent cohort study has shown that women who receive 2–3 yr of HRT after menopause do not have increased all-cause mortality, but have prolonged cardiovascular benefits (363). Windows of opportunity for the therapeutic benefits of HRT could explain the increased risk of dementia and of cerebrovascular events observed in the HERS, WEST, and WHI trials, involving women with a mean age over 65 yr. That is, once atheromas and luminal narrowing of blood vessels are established, adverse effects of HRT may prevail, including vascular inflammation and prothrombotic effects (364, 365).

The importance of starting HRT early after the onset of menopause may also provide an explanation for some discrepancies between previous observational studies, most of which have reported beneficial effects of HRT on the nervous and cardiovascular systems, and the more recent prospective studies. In fact, in most observational studies, women started HRT early, during the perimenopause or at the beginning of the postmenopause period, for the relief of climacteric symptoms, whereas in the recent large trials, women had started HRT as late as 10 to 20 yr or even longer after the onset of menopause.

However, although appearing convincing, the hypothesis proposing a "window of opportunity for HRT" is still awaiting definitive proof. In fact, not all beneficial effects of hormones may be limited to a given time window. There are indeed experimental and clinical observations suggesting that even a delayed initiation of hormone treatment may still have beneficial effects on particular brain functions (366). This view is consistent with the animal studies described in Section VII.A, providing evidence that the nervous system of old animals remains at least to some extent sensitive to the beneficial effects of ovarian steroids, in particular of progesterone, and receives further support from recent imaging studies investigating the effects of HRT on the human brain.

Data obtained by high-resolution MRI indeed show that even in elderly postmenopausal women (age range, 57–79 yr), HRT attenuates the shrinkage of both gray and white matter within brain regions known for their sensitivity to age-related decline, such as the prefrontal, parietal, and temporal cortex, and the hippocampus (367). Most importantly, HRT had significantly greater effects on different brain structures with increasing age of the women. This finding is coherent with recent experimental studies in rodents showing that hormone treatments can reverse some age-related structural abnormalities of the nervous system. The MRI study also showed that the longer the duration of HRT, the better the sparing of brain tissue. Women had received unopposed estrogen or estrogen plus MPA, but because of the small number of women in the latter group, it was not possible to draw conclusions on the possible effects of the progestin. Unfortunately, these MRI results have not been related to cognitive measures (367).

Other recent MRI studies have reported significant effects of HRT on regional brain volumes. In elderly women (age range, 60–83 yr), HRT was shown to protect against hippocampal atrophy (368) and to have a positive effect on gray matter volume of cerebellum, parietal, and occipital cortex in women older than 50 yr (369). In another study of postmenopausal women (mean age, 60 yr), HRT was associated with greater gray matter volumes in cerebellum, the amygdaloid-hippocampal complex, and the frontal, temporal, parietal, and occipital cortex. Interestingly, women who underwent HRT sooner after menopause had greater volumes of gray matter compared with women under current treatment (369). This observation agrees with the already mentioned results of the Cache County Study, showing greater cognitive performance and reduced risk of Alzheimer's disease in women with earlier HRT (354). However, the fact that there are also MRI studies which did not find an effect of HRT on gray and white matter volumes, as was the case for a recent cross-sectional study involving 213 postmenopausal women aged 60–64, should not be passed over (370). On the other hand, most of the functional imaging studies, based on the measure of changes in regional cerebral blood flow by positron emission tomography, have consistently shown beneficial effects of HRT on brain activity, in particular within brain regions sensitive to neurodegenerative changes (371, 372).

In conclusion, clinical data indicate that there may be a therapeutic window for optimal efficiency and that it may be important to start HRT early after the onset of menopause. Once age-dependent alterations or diseases of the nervous or cardiovascular systems are established, steroids may lose their efficacy and may even develop adverse effects. However, some beneficial effects of HRT seem to persist even at advanced ages. Whether there is an optimal time interval after the loss of ovarian steroids for efficient hormone replacement intervention needs to be clarified by carefully designed animal and clinical studies. Two clinical trials, the Kronos Early Estrogen Prevention Study (KEEPS) and the Early *vs.* Late Intervention Trial with Estradiol (ELITE), have been launched to collect information on the usefulness of HRT in women at the beginning of their menopause (373, 374). KEEPS is testing whether starting HRT 6 months to 3

yr after the last menstrual period will prevent the progression of atherosclerosis. ELITE is comparing the effects of estrogen treatment started in early menopause with estrogen treatment begun 10 or more years after menopause. Another important factor to be taken into account is the duration of HRT, and it is important to clarify whether longer term HRT users gain more benefits than shorter term HRT users. However, the safety of long-term hormone use in younger women is a very important issue and is still difficult to appreciate.

IX. Effects of Progesterone in Peripheral Tissues

When considering the usefulness of progesterone for protecting aging neurons and for reversing age-related structural changes and dysfunctions of the nervous system, it is necessary to also examine its effects on peripheral tissues, especially on blood vessels, mammary glands, and bones. It is rather surprising for a steroid hormone isolated more than 70 yr ago (375–379) that our knowledge concerning the actions of progesterone in these target tissues is still very limited. Nevertheless, available data are rather encouraging because they point to the safety and also to beneficial effects of progesterone.

A. Blood vessels

In a study using video microscopic recording of blood flow, blood vessel morphology, and activities of various blood cells in live animals, the administration of progesterone did not result in vascular toxicity (380). Moreover, in other studies, progesterone and 19-norprogesterone derivatives were found not to interfere with estrogen protection against vasoconstriction (360, 381–383). A nongenomic endothelium-independent vasorelaxing action of the 19-nortestosterone derivative norethisterone and of its reduced metabolite 3 α ,5 α -norethisterone have recently been demonstrated (384).

However, there has been some concern with high doses of progesterone (385, 386). Also, a recent study has suggested that progesterone treatment may exacerbate the cerebrovascular inflammatory response to lipopolysaccharide (387). This finding contrasts with the reports of the beneficial effects of progesterone in experimentally induced cerebral ischemia, where inflammation of the cerebral vasculature is a key process (see Section IV.A). Obviously, more research on the effects of natural progesterone on the vascular system is needed (388, 389).

However, as for the nervous system, serious concerns have been raised about the use of synthetic progestins such as MPA, for which vascular toxicity has been reported (380). In both peripheral and cerebral vasculature, synthetic progestins were shown to cause endothelial disruption, accumulation of monocytes in the vessel wall, and platelet activation (380). There are many other reports demonstrating negative effects of MPA on the vascular system (390–394). Whereas progesterone and nomegestrol acetate increased nitric oxide synthesis by human endothelial cells, MPA lacked such an effect. Moreover, neither progesterone nor nomegestrol acetate interfered with the effects of physiological concentrations of estradiol, whereas MPA impaired estradiol signaling

(395). In comparison with MPA, norethisterone acetate caused less cerebrovascular tension in rabbits (396). Progestins indeed differ with respect to their influence on intracranial hemodynamics (397). Results of the Postmenopausal Estrogen/Progestin Interventions (PEPI) trial, involving 596 nonhysterectomized postmenopausal women, showed that MPA attenuates the beneficial effects of estrogen on plasma lipoprotein levels (86).

B. Mammary glands

The prevailing opinion is that progesterone may contribute to the development of breast cancer, but the role of the natural hormone in breast epithelial proliferation is not completely understood (398). The assumption that progesterone may be an important breast mitogen initially came from the observation that the proliferation of breast epithelial cells is maximal during the luteal phase of the menstrual cycle. However, studies on normal human breast tissue implanted *sc* into adult female athymic nude mice found that progesterone had no mitogenic effect. Only estradiol at high levels, equivalent to those measured during the luteal phase, was found to stimulate the proliferation of breast epithelial cells (399–401).

However, in transgenic mouse models, PR-induced mammary epithelial proliferation has been shown to play a role in the initiation and progression of carcinogen-induced mammary tumors. In PR knockout mice, there was indeed a marked reduction in mammary tumor incidence when compared with wild-type mice after treatment with a carcinogen (402). Subsequently, the selective inactivation of either PR-A or PR-B has shown that the B isoform is required for a normal proliferative response of breast epithelial cells to progesterone (403). Alterations of progesterone signaling may play a significant role in breast cancer, as is strongly suggested by two observations: 1) the equilibrium of PR-A and PR-B isoform expression observed in normal breast tissue is disrupted in breast tumors; 2) the dissociation between steroid receptor expression and cell proliferation observed in normal mammary glands is lost after exposure to carcinogens and in breast tumors (401, 404–407). The conversion of progesterone to its 5 α - and 20 α -reduced metabolites, including allopregnanolone, may also play a role in regulating cell proliferation within breast tissue (408).

Results of a recent experimental study have revealed that exposure to progesterone pellets strongly increased mammary gland volume in mice deficient in the breast cancer susceptibility gene *BRCA1*, whereas the same treatment had little effect in wild-type mice (409). It was then shown that PRs are overexpressed in the mutant mammary gland because of a defect in their degradation by the proteasome pathway, and that treatment of the *BRCA1*-deficient mice with the PR antagonist mifepristone prevented mammary tumorigenesis (410).

What do clinical trials tell us about the role of progesterone in the development of breast cancer? Recent studies of large cohorts of premenopausal women strongly suggest a protective role for the endogenous hormone. Indeed, the risk of developing breast cancer was decreased in women with high luteal levels of progesterone (411, 412). Also, progesterone

seems not to potentiate, but rather to protect against, the carcinogenic effect of estrogens during HRT (413). The recent analysis of a large French cohort of 54,548 postmenopausal women, named the E3N cohort, which is the French arm of the large European Prospective Investigation into Cancer and Nutrition (EPIC), has shown that both oral and transdermal estrogen use were associated with increased breast cancer risk after only 2 yr when combined with a synthetic progestin, but not with micronized progesterone (414). These results have been recently updated; even after 8 yr of treatment, there was no significant increase in breast cancer in women taking a combination of estrogen and micronized progesterone (415). A recent preclinical study performed in rhesus monkeys has also shown that estradiol plus micronized progesterone does not stimulate breast epithelial proliferation (416). However, in the same study, estradiol plus MPA was mitogenic for the cells (416).

The tumor-promoting role of synthetic progestins in general has indeed been demonstrated by recent studies (83). Earlier studies had failed to show such an effect, but this may be explained by the fact that the percentage of women who received combined HRT was relatively low (417). Results of the Million Women Study, which was set up to examine the effects of HRT on breast cancer incidence and mortality and involved 1,084,110 UK women aged 50–64, confirmed the suspicion that current and recent users of HRT may be at increased risk of breast cancer, and also showed that the relative risk rises with increasing duration of HRT (418). Moreover, the magnitude of the associated risk was significantly greater for estrogen-progestin than for estrogen-only treatment. Breast cancer risk was significantly increased for users of MPA, norethisterone, or norgestrel/levonorgestrel (418). These results are consistent with those of the WHI trial, showing a greater breast cancer incidence for the use of estrogen-MPA combination when compared with estrogen-only preparations (419). In addition, MPA has also been suspected of being the culprit for the recurrence of breast cancer associated with HRT (420, 421). These results have received further support from two recent reviews of the literature on breast cancer (422, 423). Because micronized progesterone has no harmful effects, it is likely that the increased risk of breast cancer found with progestins may be the consequence of their non-progesterone-like effects (84). Thus, interactions with the glucocorticoid receptor and disruption of protective androgen receptor signaling may both contribute to the increased breast cancer risk in response to MPA (424, 425).

C. Bones

It is now well established that the deficiency of ovarian steroids leads to the development of osteoporosis, one of the most serious age-related disturbances affecting more than 30% of postmenopausal women (426–428). Women have an increased risk of osteoporosis after a premature or surgical menopause and in the absence of HRT (429), and the most consistent beneficial effect of estrogen alone or estrogen plus progestin therapy concerns the bones.

As for most target tissues of ovarian hormones, the roles of progesterone in bone physiology and in preventing bone

loss are less well studied than those of estradiol, and most discussions have again focused on the role of estrogen administration. In addition to the estrogens, androgens have also been shown to have a profound influence on bone physiology (427, 429). In 1990, experimental, epidemiological, and clinical data supporting an important role of progestins in bone remodeling were reviewed (430). The author proposed that progesterone may play a role in the coupling of bone resorption with bone formation; whereas the main action of estrogens would be to decrease bone resorption, the predominant effect of progestins would be to promote bone formation. Progesterone may stimulate bone formation directly by regulating the expression of target genes and also indirectly by antagonizing the effects of glucocorticoids, well known to reduce bone formation (398, 430, 431).

Since the publication of Prior (430), data relating to the specific effects of progesterone and progestins on bone metabolism have remained relatively sparse, and their significance in bone physiology remains poorly understood. Again, this can be explained by the facts that: 1) many types of progestins with different pharmacological properties are used in HRT, and it is not always clear whether their effects can be ascribed to their actions on the PRs or their interference with other steroid receptors; 2) the positive effects of progesterone and progestins could be masked by the simultaneous use of high doses of estrogen, frequently together with other bone-promoting supplements such as vitamin D and calcium (432, 433); and 3) progestagens may act in synergy with estrogens and have no effect on their own. In one clinical study, the combination of a low-dose of ethinyl estradiol with the progestin norethisterone acetate (norethindrone acetate) had a greater bone-preserving effect than a higher dose of estrogen alone (434).

Concerning the actions of progesterone, intracellular PRs are present in cultures of human osteoblasts and osteoclasts (435–437). In addition, membrane actions of progesterone have been demonstrated in rat osteoblasts (438, 439). Progesterone stimulated the proliferation of human osteosarcoma cells and osteoblasts and up-regulated the transcription of early response genes, osteocalcin, and growth factors (437, 440). In rodents, progesterone has been shown to stimulate the proliferation of osteoprogenitors. Most importantly, progesterone stimulated the growth of osteoprogenitors and the expression of PR in these cells with a comparable efficiency in young and old female rats (441, 442).

There is also evidence that progesterone may act on bone cells in concert with estradiol and potentiate its actions. Thus, in cultured human osteoblasts, estradiol has been shown to increase the specific nuclear binding of progesterone (443), and PR expression was enhanced by the phytoestrogen genistein and by the selective estrogen receptor modulator raloxifene (444, 445). Both A and B isoforms of PR (PR-A and PR-B) were induced by estrogen in human osteoblasts (446). The progestin Org 2058 had no effect on the proliferation of human osteosarcoma cells and of primary rat osteoblasts when added alone to the culture medium, but strongly potentiated the mitogenic effect of estradiol. The authors of this study proposed the existence of a specific class of progesterone-sensitive osteoprogenitors, which may only proliferate in the presence of both estradiol and progesterone (435, 447).

In human osteoblasts, a synergistic effect of estradiol and progesterone in up-regulating the expression of the insulin receptor substrate-2 has recently been reported (448). Insulin receptor substrate-2 is one of the substrates of receptor tyrosine kinases, and it plays an important role in bone formation (449).

There is so far no clear clinical evidence for a beneficial effect of progesterone or of synthetic progestins on bone in postmenopausal women, and estrogens continue to be considered as the primary bone-active agent in HRT. In the PEPI and in another recent double-blinded and placebo-controlled trial, the intake of micronized progesterone had no significant bone-protective effect, and in these studies, MPA was also found to be inefficient (450, 451). In the WHI trial, a reduction in hip fractures was observed, either after combined estrogen plus progestin or after estrogen-only treatment (27, 34). Lower doses of CEE with or without MPA than those administered during the WHI trial have also been shown to increase bone mineral density and bone mineral content (452). A prospective study of more than 138,000 postmenopausal women aged 50 to 69 yr and recruited within the above-mentioned Million Women Study revealed that all types of HRT studied, estrogen alone or in combination with MPA, norethisterone, or norgestrel/levonorgestrel, conferred substantial protection against fracture (453). Consistent favorable effects of HRT on bone density have also been confirmed by a large meta-analysis of 57 studies (454). One study has suggested that the progestin norethisterone, when administered alone, may prevent bone loss in postmenopausal osteoporosis by decreasing bone turnover (455).

However, not all studies have demonstrated an efficacy of HRT in the management of bone fractures, such as the HERS study, but women in this trial were not osteoporotic (25). Also, bone loss has been associated with MPA treatment, suspected to result from its glucocorticoid activity (456, 457), but in the WHI trial, combined CEE and MPA administration was found to reduce fractures (see previous paragraph). Based on the recent evidence, HRT has been recommended for the prevention and treatment of osteoporosis (458). However, because of the potential risks of HRT, its recommendation for osteoporosis prevention has been limited to the shortest possible time (usually 2 yr) and only in women with climacteric symptoms (428).

In conclusion, the significance of progesterone in peripheral tissues such as blood vessels, mammary gland, and bone still remains a matter of controversy. It is important to be aware that the choice of a progestin for HRT is very important with respect to efficiency and side effects. Micronized progesterone and the more selective 19-norprogesterone derivatives offer promising perspectives for more efficient and safe HRT. There is evidence that progesterone may not have cancer-promoting effects on normal breast tissue or deleterious effects on the blood vessel wall, and that it may eventually be protective.

X. Novel Perspectives for Progesterone in HRT: Multiple Signaling Mechanisms

Research over the past few years has revealed a multitude of signaling mechanisms of steroids, and in particular of

progesterone and estradiol (459–462). They offer exciting possibilities for the development of new ligands with more selective actions on specific targets, but they also point to the limitations and possible side effects of steroid compounds used at present in HRT. The still poor knowledge of the molecular mechanisms of steroid actions is hampering the development of more efficient therapies for the diseased, injured, and aging brain. The multiple mechanisms of action of progesterone and its metabolites comprise the regulation of gene transcription, the modulation of neurotransmitter receptors, and the activation of signaling cascades via new, recently identified membrane receptors. In particular the recognition of the significance of nuclear coregulator proteins in regulating the transcriptional activities of steroid receptors, as well as the cloning of new membrane steroid receptors, can be expected to completely change our vision of steroid actions on specific target tissues during the next few years.

A. Progesterone receptor isoforms and nuclear receptor coregulator proteins

The effects of progesterone on gene expression, the so-called “genomic effects,” are mediated by at least two intracellular receptor isoforms (PR-A and PR-B), which are generated from a single gene (463, 464). They only differ by an

additional 164 amino acid segment in the N-terminal region of PR-B, called the B-receptor upstream segment (465). It has been proposed that the PR isoforms undergo continuous nucleocytoplasmic shuttling, resulting from their active transport into the nucleus and their diffusion out into the cytoplasm (466). Binding of progesterone causes conformational changes of the PR, resulting in the dissociation of a chaperone protein complex, receptor dimerization, increased receptor phosphorylation, binding of the receptor dimer to specific hormone-responsive DNA elements located on target genes, and interactions of the receptor complex with nuclear coactivators (460, 467) (Fig. 3).

In vitro studies have provided evidence that PR-A and PR-B display distinct transactivational properties and that PR-B is a more active transactivator than PR-A (468–472). Moreover, in a promoter- and cell-specific manner, PR-A can even repress PR-B-mediated gene transcription, as it also does with the transcriptional activity of the estrogen and the other steroid receptors (469, 473, 474). The term “transrepression” has been coined to refer to the suppression of the transcriptional activities of other steroid receptors by PR-A, which involves the N-terminal domain of the receptors (475).

Engineered cells and transgenic mice have been used to demonstrate that PR-A and PR-B regulate the expression of different subsets of genes (476, 477). The study of transgenic

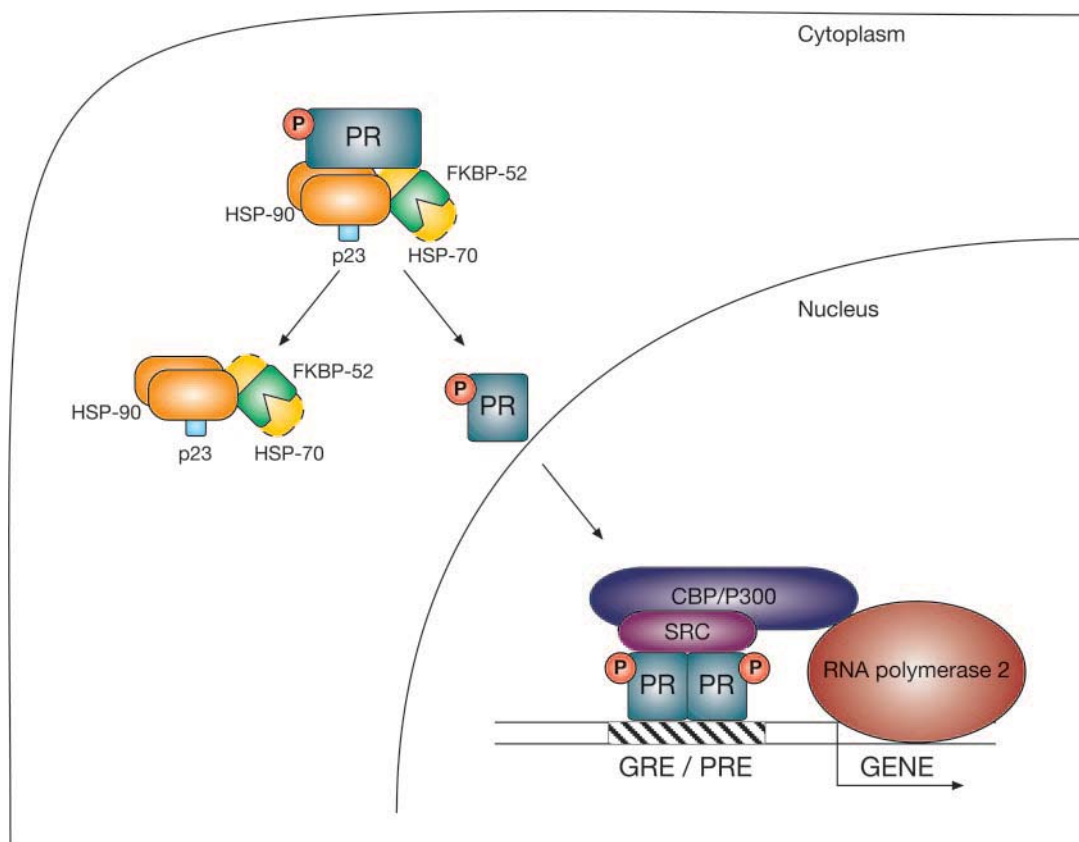


FIG. 3. Genomic actions of progesterone. Binding of progesterone (P) causes conformational changes of the PRs, resulting in the dissociation of a chaperone protein complex, composed of the heat-shock proteins (HSP), protein p23, and the immunophilin FKBP-52. The liganded PRs bind as homodimers to glucocorticoid/progesterone response elements (GRE/PRE), generally located in the promoter regions of target genes, and recruit nuclear coactivator proteins. The best characterized groups of coactivators are the p160 (SRCs) and the CBP/p300 families. Unfortunately, a heterogeneous nomenclature is actually in use for the p160 coactivators (see text).

mice overexpressing either the A or the B form of the PR has shown that regulated expression of both isoforms *in vivo* is required for normal mammary gland development (478, 479). Subsequently, the creation of mice selectively lacking either PR-A or PR-B has revealed that the two PR isoforms function as distinct transcription factors, and that they mediate different but partially overlapping reproductive responses to progesterone (403). Inactivation of PR-B resulted in reduced pregnancy-associated mammary ductal morphogenesis, but did not affect ovarian and uterine development, consistent with the observation that PR-A is sufficient for the establishment and maintenance of pregnancy. On the contrary, female mice lacking PR-A have normal mammary glands but display severe ovarian abnormalities and uterine hyperplasia and are infertile (407, 480).

Because PR-B and PR-A show different responses to progestagens, have different transactivation functions, differently influence the actions of other steroid receptors, and also activate the transcription of different progesterone-responsive genes, their coordinated expression determines the effects of progestagens (406, 481, 482). However, their respective functions have so far only been explored in reproductive tissues. In the nervous system, their biological properties and their role in the cell-specific actions of progesterone are not well defined. Many studies performed over the past decade have documented that regulation of PR-A and PR-B expression varies between different brain regions according to sex, hormonal status, and age (483–488).

In addition to PR-B and PR-A, other transcripts are generated from the PR gene. First identified in malignant progesterone target tissues or cloned from cDNA libraries, their biological functions have not been studied very much (489–497). There is, however, evidence for a possible involvement of other PR isoforms, resulting from alternative splicing or the use of additional exons, in mediating the actions of progesterone (489, 498–500). The presence and biological significance of such additional PR variants has never been explored in the nervous system.

In the mid-1990s, the discovery of nuclear steroid receptor coregulators, comprising coactivators and corepressors, has added to the complexity of steroid signaling and has opened new avenues of research into the genomic effects of steroid hormones. The number of coregulators that can be recruited by steroid and other nuclear receptors is continuously increasing, and their cell-, promoter-, and ligand-specific actions have been extensively reviewed (318, 501–508). Agonist binding is believed to increase the affinity of steroid receptors for coactivators, providing the conditions for efficacious transcriptional activation. Coactivators include molecules that facilitate the access of the basal transcriptional machinery to gene promoters, such as histone acetyltransferase coactivators. The acetylation of histone lysines indeed disrupts molecular interactions, which maintain gene promoters in a closed state. The best characterized groups are the p160 (SRC-1a, SRC-1e, SRC-2, and SRC-3) and the CBP/p300 families, which act in concert with other factors and bring histone acetyltransferase activity to the vicinity of steroid receptor complexes (Fig. 3). Unfortunately, a heterogeneous nomenclature is actually in use for the p160 coactivators (SRC-1 = NCoA-1; SRC-2 = NCoA-2 or GRIP1 or TIF-2; SRC-3 =

p/CIP, ACTR, TRAM-1 or RAC-3) (509–511). For reasons of clarity, the SRC nomenclature will be exclusively used here. It should also be emphasized that numerous other coactivator complexes are involved in bridging the RNA polymerase II complex with basal transcription factors, in providing enzymatic activities modifying DNA structure, and in allowing interactions with other nuclear proteins, which have been reviewed elsewhere (506, 512).

Important functions of the SRCs in PR-mediated physiological processes have been revealed by the study of SRC knockout mice. Thus, the ability of the uterus to mount a decidual response is markedly impaired in SRC-1 knockout females, whereas in SRC-3 knockout mice, parity-associated development of the mammary gland was compromised (513, 514). These results suggested an important role for SRC-1 in the uterus and for SRC-3 in PR-mediated effects on the mammary gland. This model is also supported by the results of a study in which transgenic PR activity indicator mice, in which cell-specific PR activation is reflected by a fluorescent reporter gene, were crossed with SRC-1 knockout and SRC-3 knockout mice (515). A mouse model, in which SRC-2 function was selectively abrogated in PR-expressing cells by using a *cre-lox* engineering strategy, has allowed the demonstration that SRC-1 and SRC-2 cooperate in the progesterone-dependent decidualization of the mouse uterus (516).

Only a few studies have examined the functional relationship between steroid receptors and their coregulators in the nervous system. However, the still fragmentary findings show that coactivators of the p160 family are critically involved in the amplification of nuclear receptor actions within the brain in a temporally and spatially coordinated manner (512, 517, 518). Knockout mouse models for the three members of the p160 coactivator family show no major phenotypic defects, despite the fact that these coactivators are largely expressed throughout the nervous system. This lack of effect suggests adaptive compensations and functional redundancy between coactivators. Thus, SRC-1^{-/-} mice are viable and only show partial hormone resistance and slight loss in reproductive functions, presumably thanks to the compensation by SRC-2 (519). Disruption of SRC-1 also slightly delays the development of cerebellar Purkinje cells, leading to moderate motor dysfunctions in adulthood (520).

However, the stereotaxic injection of antisense oligodeoxynucleotides, allowing the expression of specific coactivators to be decreased in a time- and brain region-specific manner, has allowed the demonstration of the critical role of nuclear coactivators in steroid receptor signaling within the brain. Thus, infusion of SRC-1 antisense oligodeoxynucleotides into the rat brain impaired the process of steroid-dependent sexual differentiation of reproductive behavior and brain morphology during development and inhibited progesterone-facilitated sexual receptivity in adult females (521, 522). The process of sexual differentiation of the brain and of behavior could also be disturbed by using CBP antisense oligodeoxynucleotides (523). Antisense oligodeoxynucleotides to SRC-1 and SRC-2, but not to SRC-3, inhibited lordosis behavior and the induction of the PR by estrogen within the ventromedial hypothalamus in rats and mice, consistent with the presence of SRC-1 and SRC-2 and the absence of SRC-3 within this brain region (524). That

coactivators are indeed limiting factors in steroid responses in the brain has been demonstrated in a recent elegant study. Decreasing SRC-1 expression in the preoptic area/hypothalamic region of the male quail by injecting antisense significantly blocked the activation by testosterone of both estrogen- and androgen-dependent male sexual behaviors, decreased the size of the preoptic medial nucleus, and reduced expression of the preoptic aromatase, responsible for the local conversion of androgens to estrogens. Most importantly, when injections of SRC-1 antisense were stopped, the coactivator became overexpressed in the hypothalamus, accompanied by an increase in sexual behavior and in the volume of the preoptic medial nucleus (518).

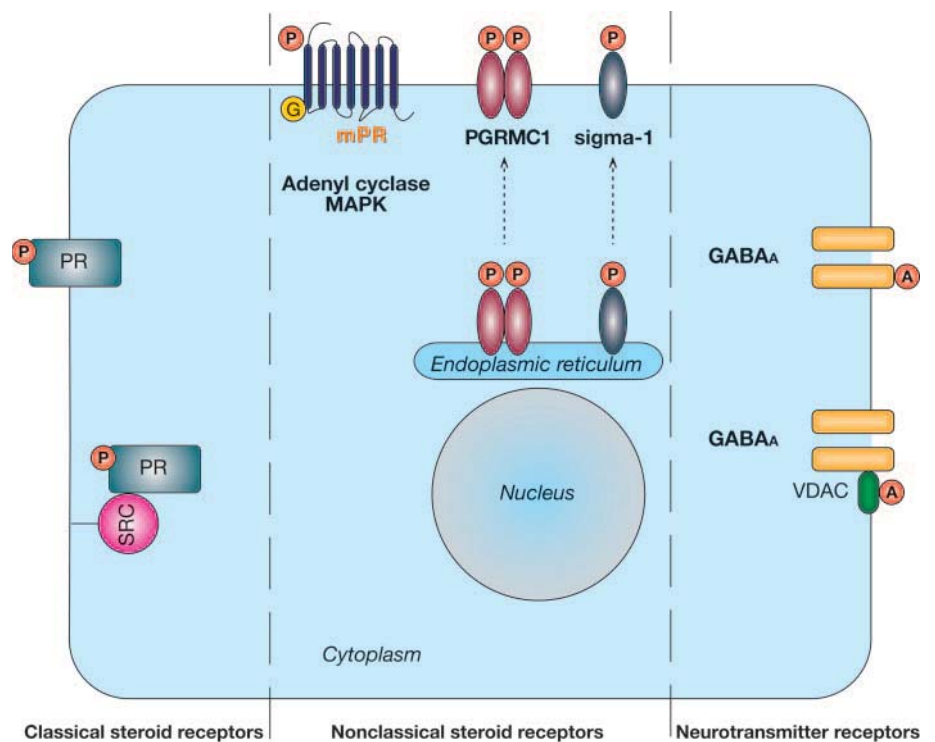
Recent studies on the interactions between steroid receptors and nuclear receptor coactivators of the p160 and CBP/p300 families in different cell types of the nervous system have revealed a novel gene- and cell-specific recruitment (525–527). These findings open new opportunities for the selective regulation of steroid actions within the nervous system. Indeed, an important concept that has emerged from recent steroid receptor and nuclear coregulator research is the influence of the cellular environment on the outcome of a particular ligand being bound. This has spurred research to develop novel synthetic ligands for steroid receptors, which would activate only a subset of the functions of natural hormones and ideally produce unique physiological effects in a particular cell type without undesired side effect activities (502, 503, 528). Such ligands have been referred to as “selective steroid receptor modulators,” including the selective estrogen receptor modulators (SERMs) and the selective progesterone receptor modulators (SPRMs), which exhibit agonistic or antagonistic activities in a cell- or tissue-specific manner (529–532).

Steroid receptor actions have even become more complex with the discovery that they are targeted not only to the nucleus, where they function as ligand-dependent transcription factors, but also to the cell membrane, where they interact with cellular signaling pathways (533) (Fig. 4). Thus, the N-terminal domain common to PR-A and PR-B indeed contains a SH3 domain interaction motif, which was shown to selectively interact with Src tyrosine kinase family members. Interactions of the PR with the SH3 domain of these signaling molecules was found to be transient and ligand-dependent and corresponds to a unique feature of the PR (534, 535). A functional consequence of PR interaction with the SH3 domain of Src kinase is the activation of the Src/Ras/MAPK signaling pathway, as has been documented in MCF-7 breast cancer cells (536, 537).

B. The modulation of neurotransmitter receptors

The inhibition of neuronal excitability is an important component of the neuroprotective effects of progesterone and its metabolites. In this regard, they differ from the estrogens, which in general have excitatory effects by potentiating the actions of excitatory neurotransmitters (24). On the contrary, progesterone and its metabolites inhibit excitatory neurotransmitter receptors and stimulate inhibitory neurotransmitter receptors (538). Thus, the $3\alpha,5\alpha$ -reduced progesterone metabolite allopregnanolone is a potent positive modulator of GABA_A receptors at physiological nanomolar concentrations (539, 540), which explains some of its psychopharmacological actions, in particular its anesthetic, analgesic, and anxiolytic effects, as well as its role in stress, depression, memory, seizure susceptibility, and alcohol dependence (541, 542) (Figs. 1 and 4).

FIG. 4. Membrane actions of progestagens. Membrane actions of progestagens comprise: 1) The regulation of neurotransmitter receptors, in particular of GABA_A receptors, by the progesterone metabolite allopregnanolone (A) and of the nAChR by progesterone (P) (right). Allopregnanolone directly binds to GABA_A receptors, but some effects of allopregnanolone may be mediated by GABA_A receptor-associated membrane proteins. 2) The binding to classical PRs after their translocation to the plasma membrane or their interaction with other membrane signaling components such as SRC (left). 3) The binding to novel G protein-coupled membrane receptors of progesterone (mPR) (middle). 4) The binding to σ_1 receptors or to PGRMC1 (also called 25-Dx) (middle). Part of the σ_1 receptors and of PGRMC1 may reside on membranes inside the cell and may translocate to the vicinity of the plasma membrane upon activation.



How neurosteroids interact with GABA_A receptors has for long been an enigma. The existence of specific steroid binding sites on GABA_A receptors has always been suspected because of the enantioselectivity of the effects of allopregnanolone (543). Moreover, a selective inhibitor of the GABA-modulatory effects of 5 α -pregnane steroids has recently been developed (544). The expression of recombinant GABA_A receptor subunits coupled with site directed mutagenesis had allowed the identification of structural motifs involved in steroid modulation (42, 538). However, it was only last year, precisely 20 yr after the discovery of the modulation of GABA_A receptors by steroid hormones, that two discrete steroid binding sites were identified on GABA_A receptors. Allopregnanolone potentiates GABA responses by binding to a cavity located between the transmembrane domains M1 and M4 of the α -subunits, whereas it directly activates GABA_A receptors by binding to interfacial residues between the α - and β -subunits (545). Previously, it had been proposed that allopregnanolone may also modulate GABA_A receptor activity by binding to associated membrane proteins, and this of course remains a possibility (546, 547).

It is important to note that allopregnanolone does not indiscriminately enhance neuronal inhibition via GABA_A receptors throughout the nervous system. Indeed, accumulating evidence shows that the steroid-GABA_A receptor interactions are specific to individual brain regions and different neurons. Thus, the modulation of GABA_A receptors by allopregnanolone has been shown to be influenced by receptor subunit composition, phosphorylation, and the local steroid metabolism (42).

As was already pointed out, some of the biological effects of progesterone may be mediated via its conversion to allopregnanolone. Thus, allopregnanolone has antiapoptotic and antiastrogliotic effects, and its administration improves cognitive recovery after TBI (101, 154). The two brain nuclei where secondary neuron loss takes place after TBI, the MDN and NBM, are indeed under the control of GABAergic innervation, and its potentiation by allopregnanolone may preserve neurons from the effects of excessive excitatory neurotransmitter release in response to injury. Most importantly, the 3 β ,5 α -reduced isomer epiallopregnanolone, which is inactive at GABA_A receptors, neither reduced secondary neuronal loss nor improved behavioral recovery in this model. After transient MCAO in male rats, allopregnanolone was even more potent than progesterone in attenuating cortical damage (548). Allopregnanolone also protects neurons in the hilus of the hippocampus from kainic acid excitotoxicity (101, 549). In this lesion model, it was also observed that progesterone administration increased levels of 5 α -dihydroprogesterone and allopregnanolone within the hippocampus, and that the neuroprotective effects of progesterone could be blocked by the administration of 5 α -reductase inhibitors. In contrast, MPA did not increase hippocampal levels of allopregnanolone and did not prevent kainic acid-induced neuronal loss (550).

In vitro studies have provided further evidence for the neuroprotective actions of allopregnanolone. Thus, when grown in culture, mouse P19 neurons were protected by allopregnanolone against NMDA-induced apoptosis (551). Exposure of cultured rat Purkinje cells to increasing concen-

trations of progesterone during oxygen-glucose deprivation revealed a dose-dependent protection by progesterone. The neuroprotective effect of progesterone could be mimicked by allopregnanolone and blocked either by the 5 α -reductase inhibitor finasteride or by the GABA_A receptor antagonist picrotoxin (552).

Most experimental studies have reported beneficial effects of allopregnanolone on brain functions, and to our knowledge, there is only one report that allopregnanolone caused neurite regression in cultures of hippocampal neurons (553). However, as clarified by the results of a more recent study, allopregnanolone-induced neurite regression may reflect a mitogenic effect of the steroid on still immature neuronal cells. Indeed, allopregnanolone has been shown to stimulate the proliferation of cerebral and hippocampal neuroprogenitor cells via GABA_A receptor-activated voltage-gated L-type calcium channels (554). An autocrine regulatory loop involved in the regulation of progenitor cell proliferation involving allopregnanolone and GABA_A receptors has recently been described in expressing the polysialylated form of the neural cell adhesion molecule (PSA-NCAM⁺) neural progenitors and isolated from neonatal rat brain. These multipotent progenitors, which spontaneously differentiate into oligodendrocytes, not only express GABA_A receptors but also synthesize significant amounts of allopregnanolone and GABA. Both GABA and allopregnanolone were shown to increase progenitor proliferation through a GABA_A receptor-mediated mechanism (555–557). The preclinical proof of concept for the therapeutic potential of allopregnanolone to promote neurogenesis has also been provided by a recent study (558). Neurons and neural progenitors are not the only cells in the nervous system that are directly influenced by the interactions between allopregnanolone and GABA_A receptors. Thus, Schwann cells have been shown to contain the messengers of several GABA_A receptor subunits, and it has been shown that allopregnanolone increases the expression of specific peripheral myelin proteins by acting on Schwann cell GABA_A receptors (178).

Thus, progesterone could become part of the neuroprotective strategies that target GABA_A receptor activity, some of which have proved to be quite efficacious (559, 560). Due to its hyperpolarizing properties, GABA protects hippocampal and cortical neurons against glutamate-induced neuronal loss both *in vitro* and in animal models of ischemia and epilepsy (559, 561). However, one should be aware that GABA does not always function as an inhibitory neurotransmitter. In the immature brain, when glutamatergic synapses are still quiescent, GABA functions as an excitatory neurotransmitter, and it only gradually converts to an inhibitory neurotransmitter. This switch from depolarizing to hyperpolarizing actions of GABA can take place very late during postnatal development for some types of neurons. Among them are the GnRH neurons in female rats, which only become hyperpolarized by GABA at the time of puberty (562). It is the transmembrane Cl⁻ ion gradient that determines whether the opening of GABA_A receptors results in inhibition or excitation. GABA_A receptors are indeed freely permeable to Cl⁻ ions, and GABA is excitatory during early development because of high intracellular Cl⁻ concentrations. During brain maturation, GABA progressively be-

comes inhibitory by the delayed expression of a transporter, which reduces intracellular Cl^- concentrations in neurons (563).

The excitatory effects of GABA during early life may explain its neurotrophic effects during brain development (564, 565). Since the pioneering studies by Wolff *et al.* (566), other laboratories have demonstrated that GABA acts as an important signaling molecule in neuronal proliferation, migration, and differentiation by causing membrane depolarization and by raising intracellular Ca^{2+} (567–569). In addition, GABA has been shown to inhibit proliferation of neuronal precursor cells, to influence the migration of neurons, and to promote morphological maturation of postmitotic neurons (570–573). However, excessive stimulation of GABA_A receptors may become toxic for immature neurons (574). Thus, the GABA_A receptor agonist muscimol can kill immature hippocampal neurons, and this neurotoxic effect is exacerbated by estradiol, consistent with the neuroexcitatory effects of this steroid (575, 576).

It is important to be aware that the depolarizing actions of GABA are not limited to the developing nervous system, because they can still take place in the adult brain, under either natural or pathological conditions (577). For instance, GABA has excitatory effects in adult dorsal root ganglia neurons and also in neurons of the cerebral cortex, depending on the resting membrane potential and on spatiotemporal interactions with excitatory amino acid inputs (578). In the suprachiasmatic nucleus, GABA acts as an excitatory transmitter during the day but is inhibitory at night (579). Most importantly, GABA can also cause the depolarization of hippocampal neurons under pathological conditions, as has been shown in temporal lobe epilepsy (580). Thus, in response to seizures, intense GABA_A receptor activation can result in a transient accumulation of Cl^- ions within neurons, changing the GABAergic effect from inhibition to excitation (581). Increased concentrations of intracellular Cl^- ions can also result from axonal injury, because of alteration of the cation Cl^- cotransporter KCC2 in the lesioned neurons. As a consequence, the activation of GABA_A receptors becomes excitatory; opening of the GABA_A receptor channel results in Cl^- efflux and depolarization of the neuronal membrane (582). Under such particular circumstances, the stimulation of GABA_A receptors could be expected to result in increased neuronal damage, and these recent observations raise the question of the safety of therapies aimed at increasing GABA concentration in the injured adult brain and during development.

However, all the trophic and protective effects of allopregnanolone are not necessarily mediated by GABA_A receptors, as shown by the following findings. In the brain of the NP-C mouse, an animal model of Niemann-Pick type C disease characterized by neurological deficits and Purkinje cell loss, levels of 5α -dihydroprogesterone and allopregnanolone are decreased, as are the expressions of the enzymes involved in their synthesis, especially in the cerebellum. Most importantly, the systemic administration of allopregnanolone significantly delayed the onset of neurological symptoms and prolonged Purkinje cell survival (583). These neuroprotective effects of allopregnanolone could be blocked by bicuculline, an antagonist specific for GABA_A

receptors. Thus they seem to be mediated, at least in part, by GABA_A receptors. However, it was then shown that the beneficial effects of allopregnanolone could be mimicked by its enantiomer (*ent*-allopregnanolone), which is inactive at GABA_A receptors. Moreover, the efficacy of micromolar concentrations of allopregnanolone and *ent*-allopregnanolone correlated with their ability to activate pregnane X receptor-dependent gene expression (584). These findings point to a role for the pregnane X receptor in mediating protective effects of elevated levels of allopregnanolone in the nervous system.

In addition to GABA_A receptors, other neurotransmitter systems are targets for the rapid modulatory effects of progestagens. Progesterone itself inhibits the activity of the neuronal nicotinic acetylcholine receptor (nAChR), but only at high micromolar concentrations (585, 586). However, it is not impossible that such concentrations are reached by locally synthesized progesterone (see Section XI), and a series of experimental observations are indeed compatible with the negative allosteric modulation of nAChR by progesterone (587). Unfortunately, even the most sensitive assay methods that are actually available do not allow a precise estimation of very localized steroid concentrations, for example at the level of synaptic clefts. The 19-norprogesterone derivative promegestone also behaves as an antagonist of the nAChR (588).

C. Novel membrane receptors of progesterone

More than 25 yr ago, a rapid action of progesterone at the level of the cell membrane, not requiring gene transcription, was described: the reinitiation of *Xenopus* oocyte meiosis (589, 590). This membrane effect involved a rapid increase in intracellular Ca^{2+} , inhibition of the adenylate cyclase/protein kinase A system, and activation of the MAPK cascade (591, 592). Recently, the amphibian homolog of the mammalian PR has been cloned and proposed to mediate nongenomic progesterone signaling in *Xenopus* oocytes after translocation into the cell membrane (593–595). However, this so-called *Xenopus* PR is not the only one mediating rapid membrane effects of progesterone on oocytes, as a recently cloned membrane progesterone receptor (mPR) is also involved in initiating the resumption of meiosis.

The characterization of membrane binding sites for progesterone has always been tentative and has usually been limited to immunostaining and molecular weight determinations. Another common approach for their study was the use of progesterone coupled to a protein or a polymer as a ligand, which supposedly does not enter cells and only acts at the cell surface. Thus, progesterone conjugated to radiolabeled BSA has been used to identify and to characterize binding sites on brain cell membranes (596). It was only in 1996 that a first putative membrane receptor of progesterone, distinct from the classical PR isoforms and comprising 194 amino acids with a single membrane-spanning domain, was isolated and cloned from porcine liver (597, 598). Binding of [^3H]progesterone to this new membrane protein was found to be reversible, saturable, and selective. Subsequently, homologous proteins were cloned in rat (named 25-Dx), cattle, and humans (599). The name 25-Dx has been used in recent

studies describing the expression and regulation of this membrane receptor of progesterone in the rat brain and spinal cord (600–603). However, in recent studies on the ovary, the protein has been referred to as “progesterone membrane receptor component 1” (PGRMC1) (604), and this nomenclature will be adopted here (Fig. 4).

In hepatocytes, the binding of progesterone to PGRMC1 was found to be associated with endomembranes rather than with plasma membranes. Moreover, expression of the cDNA of PGRMC1 in CHO (Chinese hamster ovary) cells resulted in increased microsomal progesterone binding (605). According to these earlier studies, PGRMC1 would rather qualify as an endomembrane progesterone binding protein. However, much progress has been made over past 2 yr in understanding the subcellular localization and the significance of this membrane receptor by studying the actions of progesterone in the rat ovary. There is now strong evidence that PGRMC1 mediates the antiapoptotic actions of progesterone in both rat granulosa and luteal cells. In the granulosa cells, PGRMC1 localizes to the nuclei, but after treatment of the cells with human chorionic gonadotropin, it is almost exclusively present at the plasma membrane (606). There, PGRMC1 interacts with another membrane protein, the plasminogen activator inhibitor RNA binding protein-1 (PAIRBP1; also known as RDA288 or SERBP1) and forms a complex required for transducing the antiapoptotic actions of progesterone in the ovary (607). The two proteins are also expressed in human granulosa and luteal cells, where they colocalize near the plasma membrane (608). The elucidation of the signal transduction pathway of the PGRMC1-PAIRBP1 progesterone membrane receptor complex has just begun. Available data strongly suggest that in the ovary, the membrane complex increases cGMP, which in turn activates protein kinase G (608). The cytoplasmic domain of PGRMC1 also has several potential Src homology 2 and Src homology 3 domains, through which progesterone activation could transduce an intracellular signal (606). Interestingly, PGRMC1 tends to form aggregates that can be as large as 200 kDa, although Western blots often detect PGRMC1 as a 56-kDa dimer or a 28-kDa monomer.

The presence of PGRMC1 has also been described on the cell surface of hypothalamic and spinal neurons (named 25-Dx in these studies) (600, 601). However, its signaling mechanisms have still not been studied in the nervous system. The distribution and regulation of PGRMC1 within different compartments of the nervous system may provide some clues concerning its functions. In the ventromedial hypothalamus of female rats, expression of PGRMC1 was shown to be increased by estrogen treatment, and the protein may thus play a role in the activation of female sexual behavior (600). A detailed immunohistochemical study of the distribution of PGRMC1 in the rat brain has confirmed the presence of the protein in the hypothalamus and has demonstrated its expression in circumventricular organs and ependymal cells of the ventricular walls as well as in vasopressin neurons of the paraventricular, supraoptic, and retorchiasmatic nuclei. Together with the observations that PGRMC1 was up-regulated in neurons and induced in astrocytes after TBI, these findings strongly suggested a role of the progesterone-binding protein in the maintenance of the

water balance after injury (603). A role of PGRMC1 in mediating protective effects of progesterone in the nervous system is also supported by the observation that its mRNA and protein were up-regulated by progesterone treatment in dorsal horn neurons of spinal cord-injured male rats (601). In the cerebellum, PGRMC1 is present in Purkinje cells and in the external granule cell layer. The protein is particularly abundant during early postnatal life, suggesting a role in developmental processes. In the Purkinje cells, PGRMC1 immunoreactivity was found to be associated with membrane structures of the endoplasmic reticulum and the Golgi apparatus (602). Whether PGRMC1 undergoes translocation from intracellular membranes to the plasma membrane and associates with other membrane proteins such as PAIRBP1 in the nervous system needs to be clarified.

Another potential progesterone binding protein, which may also translocate from intracellular compartments to the cell membrane and which is distinct from PGRMC1, is the sigma-1 (σ 1) receptor (Fig. 4). This receptor was first defined by its ability to bind with high affinity a variety of pharmacologically active drugs, named “sigma ligands” (609, 610). The molecular nature of the σ 1 receptor remained enigmatic until the purification and cloning of the 223-amino acid receptor from guinea pig liver microsomes (611). Subsequently, the cDNAs for the orthologs of this orphan receptor were cloned from a human placental cell line, from mouse kidney, and from rat brain cDNA libraries (612–614). It has been proposed that this endoplasmic reticulum-anchored protein may, upon activation, translocate to the vicinity of the cell membrane, where it may regulate plasma membrane-bound signal transduction (615). Physiological roles of the σ 1 receptor involve the modulation of intracellular Ca^{2+} levels and of various neurotransmitter systems (286, 616). Recent observations show that σ 1 receptors continue to be expressed in the aging brain and that they may offer interesting ways to attenuate the progressive decrease of cognitive performance during normal and pathological aging and to protect neurons against β -amyloid-induced neurotoxicity (617–619).

The endogenous ligands of the σ 1 receptor are unknown, but progesterone may be one of them because it acts as a competitive inhibitor of agonist binding (611, 620, 621). A role for the inhibition of σ 1 receptor functions by progesterone has been documented in the dorsal hippocampus. There, the potentiation of the NMDA response of hippocampal neurons and the NMDA-evoked norepinephrine release from preloaded hippocampal slices by σ 1 ligands were both strongly reduced in the presence of progesterone (622–624). Furthermore, progesterone has been shown to influence the behavioral efficacy of σ 1 receptor ligands in mice (625, 626). A recent study has explored the influence of progesterone on σ 1 receptor function during the aging process of the nervous system. Decreased levels of progesterone were measured in the hippocampus and cerebral cortex of aged, senescence-prone male SAMP/8 mice, correlating with an enhanced behavioral efficacy of σ 1 ligands (617). Again, this observation is consistent with an inhibition of σ 1 receptors by progesterone.

An important event was the cloning in 2003 of mPR from fish oocytes by the Thomas laboratory (627, 628) (Fig. 4). More than 20 genes closely related to the fish mPR have been

cloned from several vertebrate species including human, mouse, and pig. In humans, three mPR subtypes have been named mPR α , mPR β , and mPR γ (628, 629). They are all unrelated to known nuclear steroid receptors and encode proteins with seven transmembrane domains, with the characteristics of G protein-coupled receptors, and belong to a large and ubiquitous family of proteins found in both prokaryotes and eukaryotes, termed “progesterone and adiponectin receptors” (630, 631).

The mPRs are for the first time meeting the criteria of true membrane receptors: structure of a membrane-spanning protein, plasma membrane localization, expression in steroid target tissues, selective steroid binding, regulation of intracellular signaling pathways, regulation by hormones and biological functions. In the spotted sea trout, the first mPR gene cloned was found to be selectively expressed in reproductive endocrine tissues and in brain. Computer modeling predicted a protein with seven transmembrane domains, characteristic of G protein-coupled receptors. In fact, the fish mPR was shown to activate a pertussis toxin-sensitive inhibitory G protein, to inhibit adenylate cyclase activity and to activate the MAPK pathway, thus resembling the membrane actions of progesterone in *Xenopus* oocytes first described more than three decades ago (591). In fact, *Xenopus* oocytes express a mPR β ortholog at the level of the plasma membrane, and the *Xenopus* mPR β was shown to fulfill all the criteria for a progesterone receptor involved in oocyte maturation (632). The human mPR α (hu-mPR α) has also recently been shown to inhibit cAMP production in a pertussis toxin-sensitive manner and to inhibit membrane-bound adenylyl cyclase activity when transfected into the human breast cancer MDA-MB-231 cells (633).

The expression and regulation of these new membrane receptors have begun to be explored in the mammalian reproductive tract but have still not been explored in the nervous system. A recent study has provided evidence for the expression and regulation of mPRs in the rat corpus luteum, a tissue that interestingly does not contain detectable levels of intracellular PRs, but where actions of progesterone may be mediated by membrane receptors (634). Very recently, the presence of both mPR α and mPR β has been demonstrated in human myometrium, where the activation of the mPRs leads to transactivation of PR-B and to a decrease in SRC-2 expression, thus suggesting a cross-talk between membrane and nuclear receptors of progesterone (635). Both mPR α mRNA and protein are also present in MCF-7 and SK-BR-3 human breast cancer cells. Interestingly, mPR α expression was found to be higher in breast tumor biopsies than in normal breast tissue, pointing to a possible role of membrane receptors in the development or progression of breast cancer (636).

The study of membrane receptors of progesterone, including PGRMC1 and the mPRs, is a very recent field, and it is thus not surprising that some controversies are being raised (637). However, we certainly are at the beginning of a great adventure. The novel membrane receptors may indeed mediate particular functions of progesterone, and they are likely to provide exciting opportunities for the development of novel receptor ligands specifically targeting the plasma membrane of cells. A very recent study has indeed shown

that the binding characteristics of the hu-mPR α are very distinct from those of the classical PRs. In stably transfected human MDA-MB-231 cells, hu-mPR α localized to the plasma membrane and bound progesterone with high affinity and selectivity ($K_d \approx 7$ nM). However, in contrast to the intracellular PRs, the recombinant hu-mPR α did not bind progestins commonly used as contraceptives or in HRT: 19-norprogesterone derivatives (promegestone, demegestone) and 19-nortestosterone derivatives (norethisterone, norgestrel). The hu-mPR α also had no affinity for the PR antagonist mifepristone (RU486) and for the very selective PR agonist Organon-2058. Thus, hu-mPR α showed a very specific pharmacological profile, very distinct from that of the classical PRs. Estradiol and cortisol did not bind to hu-mPR α , but interestingly, testosterone was able to displace the binding of [3 H]progesterone ($IC_{50} = 390$ nM) (633). At the functional level, G protein activation by hu-mPR was also specific for progesterone because promegestone and cortisol were ineffective (633). The binding characteristics of PGRMC1 are still less well characterized. The binding of [3 H]progesterone to microsomal and solubilized membrane fractions of porcine liver could be displaced by testosterone ($IC_{50} = 3$ μ M), corticosterone ($IC_{50} = 2$ μ M), cortisol ($IC_{50} = 12$ μ M), and a higher dose of promegestone ($IC_{50} = 20$ μ M), but not by estradiol, dexamethasone, or aldosterone. It is important to note that these membrane binding sites had much lower affinity for progesterone (K_d in the 100 nM range) than mPRs (10-nM range) and PRs (1-nM range) (638).

Like the progestagens, estrogens also exert direct effects on the cellular membrane (599, 639). It has been proposed that the orphan G protein-coupled receptor GPR30 may be an estrogen membrane receptor, unrelated to the nuclear ERs and regulated by progesterone (640). However, a recent study has cast some doubt by showing that estradiol fails to activate intracellular signaling pathways in cells that lack classical ER even when GPR30 is present (641). ER-X is another recently identified putative estrogen membrane receptor, which is enriched in caveolar-like microdomains of cellular membranes and interacts with kinases of the MAPK cascade and other signaling pathways (642). It is interesting that ER-X shows a particular pharmacological profile because it binds the 17 α -isomer of estradiol and also progesterone at low micromolar concentrations (642, 643).

Thus, the binding specificities of steroid membrane receptors are very distinct from those of the classical intracellular receptors, perhaps requiring the creation of new categories of receptors. This is not further surprising because there is no homology in any region of the membrane receptors with the ligand binding domain of intracellular receptors. More work is needed to better characterize the binding and also the functional characteristics of the new steroid membrane receptors.

D. Dependence of steroid signaling on the physiopathological context

Recent observations strongly suggest that steroid actions may involve different signaling mechanisms depending on the physiopathological context. It has already been mentioned that GABA $_A$ receptors may become excitatory in re-

sponse to seizures or injury, and as a consequence, that the actions of the GABA-active steroid allopregnanolone on neuronal activity may shift from inhibitory to excitatory. Expression of the potential progesterone membrane binding protein PGRMC1 is increased after spinal cord transection in progesterone-treated rats. On the contrary, the classical intracellular PR was found to be down-regulated under these conditions (601). In response to TBI, expression of PGRMC1 was up-regulated in neurons and induced in astrocytes (603). Thus, membrane receptors of progesterone may play an important role in the neuroprotective effects of progesterone.

Another example illustrating alternative receptor signaling pathways in the injured nervous system concerns the putative estrogen membrane receptor ER-X, which is present in cortical and uterine plasma membranes of postnatal but not of adult animals, suggesting important functions in developmental processes. However, ER-X is again expressed after ischemic brain injury and may thus play a role in mediating the neuroprotective and neurotrophic effects of estrogen in the adult nervous system (642).

In conclusion, the variety of mechanisms by which progesterone and its metabolites exert their effects in the nervous system, including their genomic and rapid membrane actions, offer exciting new possibilities for the development of more efficient and safe steroid treatments, and in the future, it will be necessary to take into account this increasing complexity. An important emerging concept is that steroids may exert different actions and use different signaling mechanisms in the normal, injured, and perhaps also in aged nervous tissues. For example, membrane receptors of progesterone and estradiol are induced in response to lesion, which may mediate their protective and trophic effects. Also, the transcriptional effects of steroids mediated by classical intracellular receptors may differ between normal, lesioned, and aged nervous tissues, either because of changes in receptor expression or because of interactions with particular coregulator proteins or cooperative signaling pathways (644). In this regard, it is significant that some genes of the CNS involved in neuronal functions only become sensitive to progesterone after injury (10, 120).

XI. Novel Perspectives for Progesterone in HRT: Different Sources and Local Synthesis

Menopause is characterized by the rapid arrest of both progesterone and estradiol secretion by the ovaries, resulting in a marked decrease of their circulating levels (645). The idea of compensating for this loss is thus straightforward. However, this decline in circulating hormones does not necessarily reflect changes in steroid levels within specific target tissues, and even after menopause, women are not completely deprived of endogenous progesterone and estradiol. Indeed, these so-called “sex steroids” are not only derived from the ovaries; progesterone and its metabolites are also produced by the adrenal glands, and both progesterone and estrogens can be locally synthesized within hormone-sensitive tissues, including the nervous system. This concept is very important for fully appreciating the consequences of the decline in ovarian activity and steroidal aging in general.

A. Peripheral sources of progesterone

The adrenal glands are an important source of progesterone in rodents and in humans. Progesterone synthesized in the adrenal glands is indeed not only a precursor for the synthesis of gluco- and mineralocorticosteroids, but is also secreted into the bloodstream, stimulated by ACTH (646). Thus, progesterone continued be present in arterial plasma of female rats long after ovariectomy, but it rapidly became undetectable after combined ovariectomy and adrenalectomy (647). The secretion pattern of progesterone by the adrenal glands even shows a diurnal pattern in female rats, with peak values in the early morning hours (648). In ovariectomized and estrogen-primed rats and guinea pigs, the adrenal secretion of progesterone in response to ACTH stimulation can even become sufficient to facilitate sexual receptivity (649). Because progesterone production by the adrenal glands is regulated by ACTH, it is sensitive to stress; whereas in undisturbed female rats, adrenal progesterone secretion is about one fifth of the ovarian progesterone secretion measured during metestrous, it can become nearly as high as ovarian progesterone secretion in response to stress (650). Estradiol treatment has also been shown to stimulate progesterone secretion by the rat adrenal glands (651, 652).

Reproductive aging in female rats, one of the most commonly used animal models for studying the effects of hormone treatments, shows some particularities. When middle-aged female rats (9–10 months) enter an acyclic, persistent estrous state, levels of ovarian hormones show a marked decrease, but they still continue to have significant concentrations of circulating progesterone and estradiol provided by the adrenal glands, which again increase between 18 and 24 months of age (145, 647, 653, 654). When exposed to a male, early acyclic female rats display a mating-induced increase in progesterone from the adrenal glands along with small gonadotropin surges (655). Irregular estrous cycles can also be reestablished in old acyclic female rats by stimulation of the adrenal glands by ACTH or stress or by the administration of progesterone (656). Another means of restoring cyclic ovarian activity in aged female rats (15–20 months old) is to treat them with a nasal spray of male urine. This stimulus causes the release of progesterone from the adrenal glands, necessary for the increase in gonadotropins (657). It is noteworthy that even after several months of anovulation, old female rats can spontaneously resume ovulatory activity at irregular intervals, with the formation of functional corpus lutea, which are maintained for prolonged periods and secrete progesterone (654).

In fertile women, at least part of serum late follicular progesterone is derived from the adrenal glands (658), whereas in men plasma progesterone is exclusively of adrenal origin (659). The adrenal glands continue to be a source of progesterone in postmenopausal women. Progesterone released by the adrenal glands in response to mild inflammatory stress or to ACTH infusion was even sufficient to stimulate LH increase in postmenopausal women with estrogen replacement (660). Both the ovaries and adrenal glands contribute to serum allopregnanolone, whose levels do not change in women with age, in contrast to what is observed in men (661).

Concerning the postmenopausal ovary, it has been suggested that some steroid production may continue, in particular the synthesis of androgens, which can then be converted within target tissues to estrogens. Thus, the postmenopausal ovary has been reported to be a significant source of plasma testosterone and androstenedione, and there may be no abrupt decrease of ovarian androgen production at the time of menopause (662–664). The results of the Rancho Bernardo Study are consistent with these earlier findings; circulating levels of testosterone and androstenedione were lower in oophorectomized women when compared with unoperated postmenopausal women (665). However, the view that the climacteric ovary may be a major source of androgens has been challenged by a study showing that women averaging 12 yr after menopause with complete adrenal insufficiency had no detectable circulating androgens, strongly suggesting that the adrenal glands may be the major source. This finding was corroborated by the observation that levels of testosterone and androstenedione are very low in postmenopausal ovarian tissue, as is the expression of steroidogenic enzymes. Consistent with an adrenal origin of androgens was the observation that plasma androgen levels were strongly decreased after dexamethasone administration in postmenopausal women with normal adrenal glands (666). A recent analysis of steroidogenic enzymes in the postmenopausal ovary suggested a particular pattern of expression, which would favor the formation of $\Delta 5$ steroids (PREG, DHEA) over $\Delta 4$ steroids (progesterone, androgens). Expression of the type II 3β -HSD mRNA was found to be greatly reduced, and the enzyme could not be detected by Western blot analysis (667).

In addition to the steroidogenic endocrine glands, enzymes required for the synthesis and metabolism of progesterone are also expressed and functional in many peripheral tissues. Local synthesis and intracrine actions of progesterone have been proposed to take place in various tissues, including mammary gland, uterus, adipose tissue, bone, skin, kidney, liver, heart, blood vessels, brain, and peripheral nerves (9, 10, 457, 668–676). However, although expression of the 3β -HSD enzymes has been studied in a variety of tissues at the mRNA and protein levels, their activities and substrates have only been measured in a limited number of studies. This represents a serious gap because the 3β -HSDs can catalyze the conversion of a variety of steroid substrates; they not only convert PREG to progesterone, but they are involved in the biosynthesis of the major classes of steroid hormones by catalyzing the conversion of $\Delta 5$ - 3β hydroxysteroids to their corresponding $\Delta 4$ - 3 -ketosteroids. As a consequence, the type of steroid produced by the 3β -HSDs in a given tissue will depend on the available substrates.

The 3β -HSD gene family has recently been reviewed in detail (677). The genes of two 3β -HSD isoenzymes have been cloned in humans: the type I 3β -HSD gene (*HSD3B1*), predominantly expressed in placenta and peripheral tissues including the mammary gland and skin; and the type II 3β -HSD gene (*HSD3B2*), expressed in the gonads, adrenal glands, and brain (677–679). Multiple 3β -HSD isoenzymes have been cloned from several other species: four members of the rat and six members of the mouse 3β -HSD family (680, 681). Unfortunately, for each of these species, the different

3β -HSD isoforms have been numbered according to the chronology of their discovery, and the isoforms of different species do not correspond by number.

Estradiol can also be synthesized in significant amounts from circulating precursor androgens in extragonadal sites such as the breast, adipose tissue, bone, and brain. In these tissues, the aromatase enzyme, which converts testosterone to estradiol or androstenedione to estrone, is present and becomes the major source of estrogens after the menopause (682). The significance of local estrogen formation in peripheral tissues is well documented by the benefits of aromatase inhibitors in breast cancer (683–685). Thus, the very low levels of estrogen that persist in the plasma of postmenopausal women result from their release by a variety of tissues, but they do not necessarily reflect the local concentrations of the steroid within these tissues. Moreover, the capacity of peripheral tissues to produce estrogens, such as adipose tissue, seems to increase with age (686). Consequently, after menopause, estrogens mainly become paracrine/autocrine signaling molecules within extragonadal tissues (687, 688).

The circulating substrates for the local formation of estrogens in postmenopausal women are androgens mainly derived from the adrenal glands, and DHEA, also of adrenal origin (688). As a consequence of the progressive decline in the circulating levels of DHEA and DHEA sulfate (DHEAS) with progressing age, substrate availability for the local synthesis of androgens and estrogens decreases with age (689–691). The measurement of DHEAS in a cohort of 3029 women from the SWAN study has revealed that its levels do not steadily decline during the transition to late perimenopause, and that they may even transiently increase in some women at this stage. Interestingly, changes in circulating testosterone and estradiol correlated with changes in DHEAS (692). These observations are consistent with results of a study in female macaques, showing a transient increase in DHEAS levels during the time of perimenopause (693). These observations point to an important role of the precursor steroid DHEA during the menopause transition period.

B. Synthesis of progesterone in the nervous system

The nervous system is an important site of steroid formation; neurons and glial cells not only have the capacity to convert circulating steroid precursors to neuroactive steroids, but they can also synthesize them *de novo* from cholesterol (669, 694). Steroids that are synthesized within the CNS or PNS have been named “neurosteroids” (695, 696). To qualify as a neurosteroid, there are two requirements: 1) persistence of the steroid in the nervous system in the absence of the steroidogenic endocrine glands (gonads and adrenal glands); and 2) expression and activity of the enzymes involved in their synthesis within the nervous system.

From an evolutionary point of view, progesterone can be synthesized within the nervous system of all species studied so far, and 3β -HSD enzymes are widely distributed throughout the brain of fishes, amphibians, birds, and mammals (189, 671, 697–704). Estradiol can also be formed within the brain in all these phyla, but particularly high amounts of estradiol are synthesized in the brains of fishes and birds. Thus, in goldfish, the activity of the aromatase enzyme is about 10-

fold higher in the hypothalamus than in the ovaries (705, 706). In songbird species, such as the zebrafinch, the brain synthesizes very large quantities of estradiol from androgens and is the exclusive source of circulating estradiol (707, 708). The presence, regulation, and functional significance of the aromatase has been studied in detail in the bird brain (709, 710).

In the rodent nervous system, the synthesis of neurosteroids has been extensively reviewed (696, 697, 703, 711) (for an overview of steroidogenic enzymes in the pathways from cholesterol to the different steroid hormones, see Ref. 712) (Fig. 5). The mitochondrial cytochrome P450_{scc}, the cholesterol side chain cleavage enzyme that catalyzes the *de novo* synthesis of PREG, is expressed at low levels throughout the

rodent brain and has been detected in most cell types (697, 713–717). Its presence and activity have recently been demonstrated within rat pain pathways: dorsal root ganglia, dorsal horns of the spinal cord, and somatosensory cortex. Results are consistent with an important role of neurosteroids in the control of pain mechanisms (718–720). The second enzyme necessary for the synthesis of progesterone, the 3 β -HSD, is also present in neurons and glial cells. The enzyme is present in the cytoplasm and can also form a catalytically active molecular complex with the cytochrome P450_{scc} at the inner mitochondrial membrane (721). Detailed *in situ* hybridization studies have shown its widespread expression in the rat CNS during development and adulthood (704, 722, 723). By measuring the cerebral accumulation of PREG in

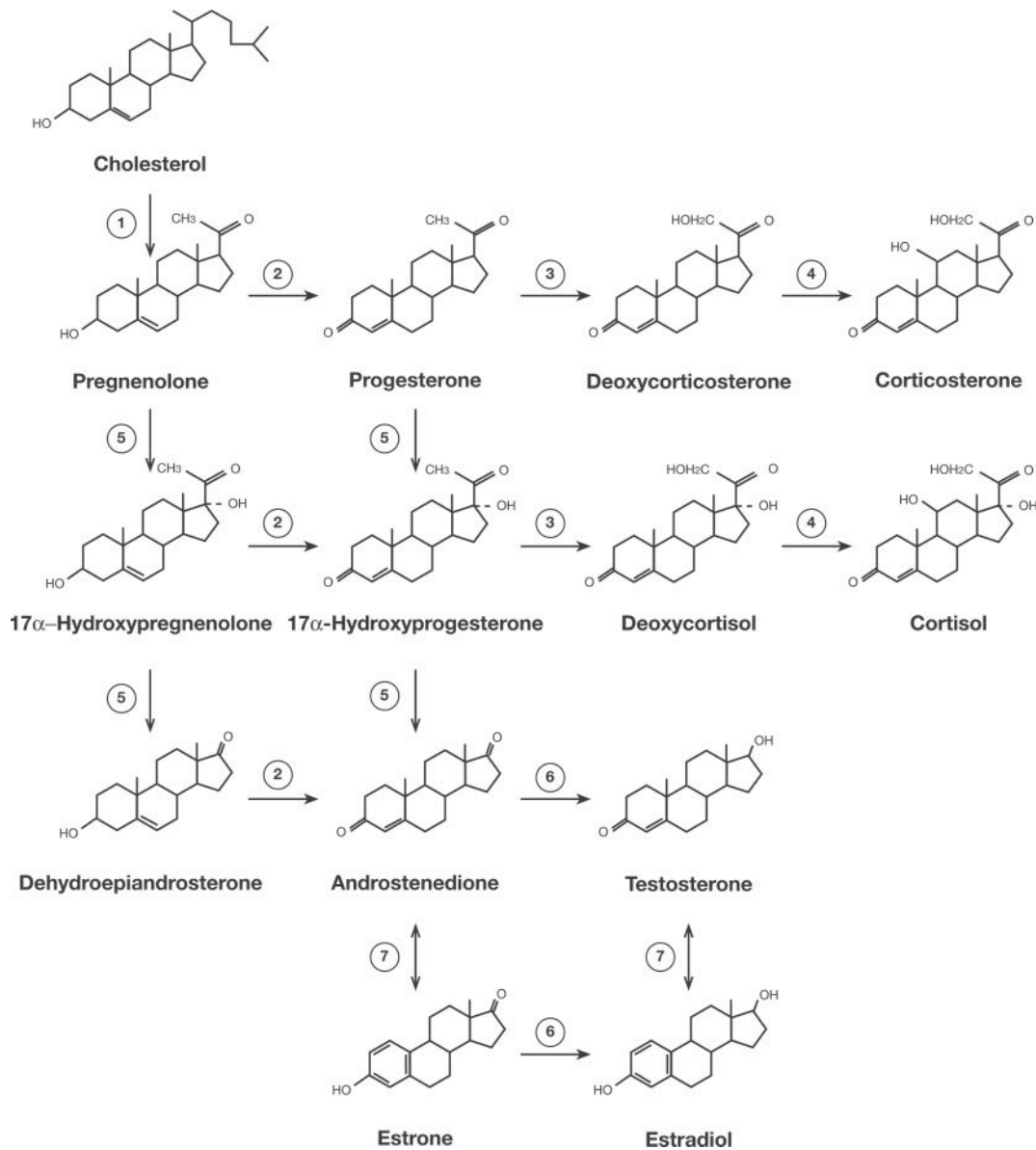


FIG. 5. Biosynthetic pathways of steroids. Conversion of cholesterol to pregnenolone is catalyzed within the mitochondria by the cytochrome P450_{scc} (scc = side chain cleavage) (1). The other enzymes involved in steroid metabolism are: 2) 3 β -HSD; 3) cytochrome P450_{c21} (21-hydroxylase); 4) cytochrome P450_{11 β} (11 β -hydroxylase); 5) cytochrome P450_{c17} (17 α -hydroxylase/17,20-lyase); 6) 17 β -hydroxysteroid oxidoreductases (17 β -hydroxysteroid oxidoreductase); and 7) aromatase. Note that most enzymes accept several substrates and can catalyze multiple reactions. It is thus necessary to know which steroid substrate is available in a cell before concluding which metabolite is formed.

castrated and adrenalectomized rats after administration of the 3β -HSD inhibitor trilostane, it has been demonstrated that the conversion of PREG to progesterone is a very active metabolic pathway in the rat brain (724). The synthesis and functions of progesterone have been extensively studied in Purkinje cells. These neurons also express the aromatase enzyme, suggesting the local formation of estradiol (725).

Which steroids can be locally synthesized within the mammalian nervous system? This question needs to be answered cautiously, and results of related studies should always be interpreted within their precise context. In fact, the expression and activity of steroidogenic enzymes are subject to complex regulations, and they may be expressed and functional in a defined compartment of the nervous system only under particular conditions, depending on environmental influences, cellular interactions, the presence of neurotransmitters or neuropeptides, the developmental stage, or the integrity of the nervous tissue. It is also important to be aware of some technical constraints. Thus, the relevance of studies on the formation of neurosteroids by cultured neural cells isolated from embryonic or newborn animals always needs to be examined *in vivo* because of the phenotypic plasticity these cells exhibit *in vitro*. Also, studies of the expression of steroidogenic enzymes by *in situ* hybridization, RT-PCR, or immunocytochemistry, although they provide very important information, do not demonstrate their functionality.

The synthesis of progesterone and its 5α -reduced metabolites by neurons and glial cells is now well established (726–730). Expression and activity of the cytochrome P450c17 in the brain and the possibility of a local synthesis of DHEA and androstenedione, the obligatory precursors of androgens and estrogens, has long remained controversial (in contrast to humans, DHEA is not secreted by the rat and mouse adrenal glands). Thus, P450c17 expression within specific sites of the rat PNS and CNS was found to be restricted to embryonic and early neonatal life (731). However, the mRNA of the enzyme has also been detected by RT-PCR in the adult rodent brain (715), and neurons and astrocytes in culture have been shown to convert PREG to DHEA if grown under specific conditions (732). By combining molecular, anatomical, and neurochemical approaches, expression and activity of the cytochrome P450c17 have recently been demonstrated in the rat spinal cord (733). Two laboratories have even provided evidence for a local synthesis of glucocorticoids (734) and estradiol (735) in the rat brain, at least within specific regions, and the term “synaptocrinology” has been suggested to refer to the effects of locally synthesized neurosteroids on synaptic plasticity (736).

A recent study has shown that an increase in PREG and progesterone synthesis is part of the responses of nervous tissues to injury, consistent with an important role of these neurosteroids in the protection and regeneration of nerve cells (730). Expression of the 3β -HSD and levels of progesterone are also strongly up-regulated in the brains of dysmyelinating jimpy and shiverer mouse mutants (737) and in the spinal cord of streptozotocin-treated diabetic rats (738). Neuropathic pain has also been shown to increase the neosynthesis of PREG and allopregnanolone in the rat spinal cord, suggesting a role for these neurosteroids in nociception

and neuroprotection (739, 740). All these observations provide strong evidence for the concept that an increase in the synthesis of progesterone and its metabolites may be part of the mechanisms by which nerve cells cope with neurodegeneration. A recent study has provided evidence that the synthesis of progesterone and other neuroprotective neurosteroids is affected by Alzheimer’s disease key proteins. Thus, the overexpression of amyloid precursor protein in human neuroblastoma cells inhibited progesterone synthesis. On the contrary, overexpression of human native tau protein enhanced progesterone formation, but tau protein with a pathogenic mutation was devoid of actions on neurosteroidogenesis (741).

Under normal circumstances, expression of the brain aromatase is restricted to specific neuronal populations. These aromatase-containing neurons are located in brain areas involved in neuroendocrine control. However, in response to different types of injury, aromatase expression and activity is induced in reactive astrocytes, strongly suggesting a role for local astroglial estrogen formation in brain repair (23, 742). Taken together, these results point to an important role for the local formation of progesterone and estradiol in the lesioned or diseased nervous system, which may complement an insufficient supply or override an inappropriate supply of steroid hormones by the endocrine glands. An important area for future investigation is changes in the endogenous capacity of the aged brain to synthesize and metabolize steroids (743–745).

C. Neurosteroids in the human nervous system

Evidence has accumulated over the past few years that neurosteroids are also synthesized and metabolized in the human nervous system (9, 746–748). The presence of the cytochrome P450scc was first detected in the human brain by immunocytochemistry (749), and subsequently, several studies have described the presence of P450scc mRNA in different brain regions (678, 750–752). A recent study has shown that P450scc and StAR are coexpressed in cells of the human brain, consistent with the active sites of neurosteroidogenesis (753). The type II isoform of the human 3β -HSD is largely expressed in different parts of the brain and spinal cord (678, 752), and the enzymes necessary for the metabolism of progesterone are also present in the human brain (754, 755). The activity of the aromatase has been characterized in microsomal preparations of temporal lobe biopsies as well as in cerebral cortex and subcortical white matter samples of adults and children with epilepsy (756, 757).

Because both the endocrine glands and a local production contribute to the pool of steroids present within nervous tissues, the age-dependent decrease in circulating levels of steroids may not necessarily reflect changes in their availability for neural cells. It is not known whether the capacity of neurosteroid synthesis changes with age in humans or whether neurosteroids can compensate for the age-dependent decrease in the activity of the steroidogenic endocrine glands. So far, only a few studies have investigated the distribution of steroid concentrations in aged human brain by RIA. The data of three studies show that elevated levels of PREG and DHEA remain present in the brains of the elderly

(758–760). In the study of Lacroix *et al.* (760), based on nine women and one man (ages, 76–93 yr), levels of PREG, progesterone, androstenedione, and DHEA varied little among different brain regions and were about seven to nine times higher in brain tissue when compared with plasma, consistent with their accumulation or local synthesis. Very high levels of PREG have also been measured in human peripheral nerves, where its mean concentration was about two orders of magnitude higher than in blood (761).

In a more recent study, brain levels of progesterone, 5 α -dihydroprogesterone, and allopregnanolone were found to be higher in fertile women in their luteal phase (ages 18–42 yr) compared with the postmenopausal women (ages 59–85 yr), and were thus obviously dependent on ovarian production (762). However, despite the correlation between blood and brain levels of progesterone, it is noteworthy that brain concentrations of progesterone and its metabolites remained elevated and that they were only decreased by about half in the postmenopausal women with very low serum concentrations. In another study, postmortem concentrations of estradiol were also found to be higher in the blood and in specific brain regions of fertile women when compared with postmenopausal women, but again, significant levels of estradiol were measured in the brains of the latter (763).

Levels of steroids have also been measured by RIA in brain and CSF of Alzheimer's patients and aged controls. Some of these studies have reported differences in brain steroid levels between controls and Alzheimer's patients, whereas others found no differences (764, 765). Brain progesterone and estradiol may have a direct influence on Alzheimer's disease. Thus, by crossing aromatase knockout mice with a transgenic mouse model of Alzheimer's disease, it has been demonstrated that β -amyloid peptide is more rapidly deposited in estrogen-deficient brains (765). Progesterone may also play a significant role in senile plaque formation because transcription of the gene encoding neprilysin, one of the major enzymes involved in β -amyloid degradation, is up-regulated by progesterone (766).

The development of very sensitive and precise analysis of steroids by gas chromatography/mass spectrometry corresponded to a major technological breakthrough (767). In fact, because of its great sensitivity, this method allows the analysis of small amounts of neurosteroids in nervous tissues with great precision and reproducibility. The different steps of the assay have been optimized to allow the simultaneous measure of a large range of free and conjugated neurosteroids within distinct brain regions (768, 769). A comparative analysis of the concentrations of several neurosteroids in various brain regions between aged Alzheimer's patients and aged nondemented controls has been recently reported (770). This study was also the first to use gas chromatography/mass spectrometry technology to quantify neurosteroids in human brain and thus provided reference values. In agreement with previous studies using RIA, PREG was the most abundant neurosteroid in the different brain regions analyzed. Steroids found at the highest concentrations were, in decreasing order, PREG > DHEA > progesterone > PREG sulfate > DHEAS > allopregnanolone. It is important to note that levels of all these steroids, except for allopregnanolone, were found to be elevated in the brains of the old patients,

and that they were much higher than previously reported blood levels (690, 771, 772). There was also a general trend toward lower levels of the steroids in different brain regions of Alzheimer's patients when compared with controls (770).

D. Regulation of the local synthesis of progesterone in the nervous system

Stimulating the local synthesis of neurosteroids with neuroprotective or neuroregenerative potentials may offer novel perspectives not only for treating lesions and diseases of the nervous system, but also for hormone therapies in the elderly. This line of thinking indeed receives support from observations that the formation of neuroactive steroids, and in particular of progesterone and its reduced metabolites as well as of estradiol, is strongly up-regulated in the lesioned and diseased nervous system (see *Section XI.B*). However, there are still serious gaps in our knowledge of the regulatory mechanisms involved in the biosynthetic pathways of neurosteroids, some of which may be distinct from those described for the steroidogenic endocrine glands. Thus, particular regulatory mechanisms of P450_{scc} transcription have been identified in the nervous system (773, 774). Interactions between neural cells obviously play an important role in the regulation of neurosteroid biosynthesis. Thus, progesterone synthesis is regulated in astrocytes by a still unidentified autocrine factor (775) and in Schwann cells by diffusible neuronal molecules (727).

Furthermore, neurotransmitters and neuropeptides play a particularly important role in modulating the synthesis of steroids within the nervous system. Thus, the GABA_A receptor agonist muscimol and the central-type benzodiazepine receptor agonist clonazepam both stimulated PREG synthesis in retinal ganglion cells (776). Several neurotransmitters and neuropeptides involved in neurosteroid regulation have been identified in the frog brain, where all the major steroidogenic enzymes are expressed and functional (702). In the frog hypothalamus, 3 β -HSD-immunoreactive neurons express GABA_A receptors involved in the inhibition of the conversion of PREG to progesterone or to DHEA (777), whereas activation of central-type benzodiazepine receptors by octadecaneuropeptide was found to increase the synthesis of these neurosteroids (778). Steroid-producing cells of the frog diencephalon are innervated by neuropeptide Y-immunoreactive fibers, and neuropeptide Y was shown to inhibit the sulfation of both PREG and DHEA (779). On the contrary, vasotocin and mesotocin, the respective amphibian orthologs of mammalian vasopressin and oxytocin, were found to stimulate the synthesis of progesterone and DHEA in diencephalic nuclei (780). These studies in an amphibian have demonstrated the important role of neuropeptides in the regulation of neurosteroid biosynthesis, but their relevance for mammalian species needs to be examined. A recent study has shown that in spinal sensory circuits of the rat, substance P, a major mediator of pain signals, inhibits the conversion of progesterone to allopregnanolone (719). In the rodent hippocampus, stimulation of neurons with NMDA induced a significant production of estradiol (735).

Circulating steroid hormones are also involved in the regulation of neurosteroid biosynthesis within the brain. A nice

example, which also illustrates well the functional significance of locally synthesized neurosteroids, is the induction of progesterone synthesis by estradiol within the hypothalamus. The Micevych group has indeed shown that systemic estrogen treatment of ovariectomized and adrenalectomized (ADX) rats increases levels of progesterone in the hypothalamus. Estradiol failed to induce an LH surge in the ovariectomized-ADX females if the increase in hypothalamic progesterone was blocked by an inhibitor of the 3β -HSD (676, 781). These observations strongly suggest that progesterone synthesis in the hypothalamus is critically involved in the positive feedback mechanisms of estradiol that trigger the LH surge. Other experiments have shown that estradiol induces the synthesis of progesterone in hypothalamic astrocytes by acting through a membrane-associated receptor and by releasing intracellular stores of Ca^{2+} (782). Noteworthy, estradiol increased the production of progesterone in astrocytes from postpubertal, but not from neonatal, female rats (782). Also, castrated/ADX male rats, which in contrast to females do not show an estrogen-induced LH surge, had no increase in hypothalamic progesterone after estrogen treatment (781).

Intramitochondrial cholesterol transporters offer very promising possibilities for stimulating the synthesis of neurosteroids and for promoting neuroprotection and neuroregeneration. One of them, the peripheral benzodiazepine receptor (PBR), is a mitochondrial protein particularly abundant in steroid-producing tissues and also in glial cells. Recently, the PBR has been renamed “translocator protein (18 kDa)” (TSPO) (783), and this nomenclature will be adopted here. Both *in vitro* and *in vivo* studies have demonstrated that the TSPO is necessary for the transport of cholesterol from the outer to the inner mitochondrial membrane, where the cytochrome P450_{sc} is located. This intramitochondrial transport of cholesterol is a rate-limiting step in the biosynthesis of steroids (784, 785).

Ligands of the TSPO not only increase the synthesis of steroids by the steroidogenic endocrine glands, but they also allow the stimulation of the synthesis of neurosteroids, as has been shown in cultured glial cells and in the brain of castrated and ADX rats (729, 786–789). The possibility of increasing the synthesis of neuroactive steroids has stimulated recent efforts to develop more selective and efficient TSPO ligands (790–792). Interestingly, in response to injury, TSPO expression is increased in the brain and in peripheral nerves (793–795). A strong up-regulation of TSPO expression is also observed during neurodegenerative diseases (Alzheimer’s disease) and demyelinating diseases (multiple sclerosis) (785, 796). In the hippocampus of Alzheimer’s patients, increased TSPO expression correlated with increased levels of PREG (785). These observations suggest that TSPO ligands may offer novel means for neuroprotection and for improving age-dependent dysfunctions of the nervous system. TSPO ligands have indeed been shown to protect neurons from excitotoxic injury, to promote their regeneration, to reduce inflammatory responses, to decrease reactive gliosis, and to reduce aging-associated myelin degeneration (792, 797–800). However, a role for neurosteroids in mediating these beneficial effects of TSPO ligands awaits demonstration. That is, TSPO ligands can influence many aspects of mitochondrial

activity and not only steroidogenesis because the TSPO is physically associated with the voltage-dependent anion channel and the adenosine nucleotide translocase, which form the backbone of the mitochondrial permeability pore (801, 802).

A second protein necessary for the intramitochondrial transport of cholesterol and steroidogenesis is the steroidogenic acute regulatory protein (StAR) (803, 804). Clinical studies of patients suffering from congenital lipoid adrenal hyperplasia, as well as studies of StAR null mice, have demonstrated the indispensable role of StAR in regulated steroidogenesis (805). Recent studies have shown that there is a functional interaction between StAR and TSPO required for cholesterol delivery into the mitochondria and steroid formation (806). In response to excitotoxic injury, StAR is strongly induced in hippocampal neurons (807, 808). Interestingly, whereas TSPO and StAR expression are decreased in the aging gonad (809), StAR expression has been reported to be up-regulated in the brains of aged animals (807). Whether the increased StAR expression reflects a compensatory increase in local neurosteroid synthesis within the aged brain would be an interesting working hypothesis. It is important to note that StAR expression may be regulated in a specific manner within distinct compartments of the nervous system, and as for the TSPO, in a different manner than in the endocrine glands (810). Thus, StAR expression is induced by cAMP within the gonads but down-regulated in Schwann cells (811).

A molecule that efficiently stimulates the synthesis of progesterone and its reduced metabolites in the rat brain is the anxiolytic drug etifoxin [2-ethylamino-6-chloro-4-methyl-4-phenyl-4H-3,1-benzoxazine hydrochloride (Stresam)], which is a ligand for both GABA_A receptors and TSPO (729, 812, 813). An increase in allopregnanolone formation indeed contributes to the anxiolytic effects of etifoxin, as has been demonstrated in the Vogel conflict test (729). As already mentioned, the anxiolytic properties of allopregnanolone have been studied in different stress models and also in aged animals (284, 814, 815).

Other drugs used in medicine directly affect the biosynthesis or metabolism of neurosteroids. For example, fluoxetine (Prozac) and other selective serotonin reuptake inhibitors (SSRIs), which are widely used for the treatment of depression, enhance allopregnanolone levels in the rat brain (816, 817). At the molecular level, SSRIs have been shown to stimulate the accumulation of allopregnanolone in the brain by increasing substrate affinity of the human ARK1C2 enzyme (type III 3α -HSD), which converts 5α -dihydroprogesterone to allopregnanolone. Sertraline also blocked the reverse oxidative reaction, the conversion of allopregnanolone to 5α -dihydroprogesterone catalyzed by microsomal RODH-like SDRs (818). Like the SSRI, the antidepressant mirtazapine enhanced the formation of allopregnanolone and inhibited the oxidation of allopregnanolone into 5α -dihydroprogesterone.

Preclinical observations strongly suggested a key role for neurosteroids in depression and an antidepressant potential of allopregnanolone: levels of allopregnanolone were shown to be decreased in human CSF and plasma during major depression, and their levels were restored by antidepressant

treatment (819, 820). However, more recent studies have shown that increased levels of allopregnanolone and other neuroactive steroids observed in response to antidepressant treatment merely reflect a specific pharmacological effect of the drugs, which may contribute but is not essential for the clinical responses: 1) changes in neurosteroid levels were found to be comparable between responders and nonresponders during antidepressant treatments; and 2) nonpharmacological antidepressive treatments, such as sleep deprivation and transcranial magnetic stimulation, can have beneficial effects without affecting steroid levels (815, 821, 822).

In conclusion, progesterone and estradiol continue to be locally produced within a variety of peripheral tissues, long after the arrest of ovarian functions. Age-dependent changes in their circulating levels thus do not necessarily reflect changes in their concentrations within hormone-sensitive tissues. This is particularly true for the brain, spinal cord, and peripheral nerves, where neurosteroids can be synthesized *de novo* from cholesterol. Only very limited information is available concerning changes in neurosteroid levels with age. The use of pharmaceutical agents that increase the synthesis of biologically active steroids within the nervous system offers novel therapeutic perspectives for promoting healthy aging and for treating age-related dysfunctions of the nervous system. Anxiolytic and antidepressant drugs also increase levels of neurosteroids, especially of 3α -reduced pregnane steroids, which in turn appear to play an important role in the pathophysiology of psychiatric disorders (815). Neurosteroids thus offer interesting perspectives for pharmacological intervention on age-associated psychiatric disorders, including mood and anxiety disorders.

XII. Conclusions

Whereas progesterone elicits increasing interest for its usefulness in treating lesions and degenerative diseases of the nervous system (13, 105, 270), its therapeutic potential for HRT is far from being completely appreciated. There are two major reasons for this situation: 1) little knowledge concerning the physiopathological effects of natural progesterone and of its metabolites in many tissues, which is rather surprising more than 70 yr after the discovery of the hormone; and 2) unjustified generalizations of the effects of specific compounds, often with an inappropriate nomenclature. In fact, different synthetic progestins have very different pharmacological and biological properties, sometimes very distinct from progesterone, and referring to them as a single class is not acceptable. Most serious are the risks associated with the use of some of the currently available progestins, such as increased risk of breast cancer and of cardiovascular complications. Within the nervous system, MPA has been shown to inhibit the beneficial effects of estradiol and even to exert damaging effects. On the contrary, these risks are not found with the use of progesterone, and until the development of more selective and safe progestins, micronized progesterone may be an option for efficient and safe HRT, although such an option may not be very attractive for the pharmaceutical industry.

At least in the nervous system, the beneficial pleiotropic actions of progesterone and its close reduced metabolites are now well recognized, including their neuroprotective, neurotrophic, and promyelinating effects, thus offering interesting perspectives for the protection and recovery of aged nerve cells. Particularly encouraging are recent experimental observations obtained in rodents, showing that the aging nervous tissue remains to some extent sensitive to trophic effects of progesterone, and that treatment with progesterone even allows reversal of age-related structural abnormalities and dysfunctions. These findings are coherent with recent neuroanatomical studies, which have demonstrated that the aging process is not associated with a massive loss of neurons, but rather with more subtle changes in neuronal circuits and alterations of the myelin sheaths. Some of these age-dependent changes appear to be reversible, making attempts to treat them meaningful.

However, one should be aware that information in the literature concerning the effects of ovarian steroids on the aging nervous system are still fragmentary and that they do not allow final conclusions concerning the preservation or modifications of steroid responses during aging to be drawn. This is obviously a fundamental question when considering HRT, and hormone therapies in general, for the elderly. Some reports indeed advise caution with respect to hormone actions in the aging nervous system. In particular at advanced ages or after long-term hormone deficiency, some brain functions may become less sensitive to steroids. Under such circumstances, the administration of steroids may even become dangerous by increasing the risk of cerebral stroke and by precipitating the development of cognitive dysfunctions and of neurodegenerative diseases. For some steroid effects, there may be windows of opportunity, during which hormone treatments may be most efficient. But this concept awaits further demonstration, and safety of prolonged hormone treatment remains an important issue.

It is also important to improve our knowledge concerning the molecular signaling mechanisms of progesterone. Although much progress has been achieved over the past few years, they are still poorly explored in the nervous system. For example, whereas the functions of the two PR isoforms have been extensively studied in reproductive tissues, their respective significance in the brain and peripheral nerves remains largely unknown. The recognition of gene- and cell-specific recruitment of nuclear coregulator proteins by steroid receptors and the recent identification and cloning of membrane receptors of progesterone open completely new perspectives for the development of more efficient, selective, and safe progestagen treatments. An important emerging concept is that steroids may use different signaling mechanisms in the normal, injured, and perhaps also aged nervous tissues. In the future, this increasing complexity will need to be taken into account when examining the usefulness of hormone replacement strategies.

When appreciating the consequences of steroidal aging, it is important to be aware that steroids are not only produced by the endocrine glands, but that they are also locally formed within hormone-sensitive tissues, where they act as autocrine/paracrine signaling molecules. As a consequence, the decline in circulating steroids does not provide information

concerning changes in steroid levels within specific tissues, and even after the menopause, women are not completely deprived of endogenous progesterone and estradiol. In the nervous system, progesterone and estradiol are synthesized by neurons and glial cells. The stimulation of their synthesis by pharmaceutical agents such as TSPO ligands offers another perspective for promoting healthy aging and for treating age-related dysfunctions of the nervous system.

Taken together, the rapid development of knowledge concerning the biosynthesis, mechanisms of action and effects of progesterone in the nervous system offer very promising possibilities for HRT. However, experimental and clinical data are still insufficient for making definitive therapeutic recommendations, and more basic research and carefully designed clinical trials are needed for better understanding hormone actions in the aging nervous system.

The quest for more selective and safe steroid compounds for HRT is a very important issue. Laboratories are searching for chemical modifications that eliminate the side effects associated with chronic hormone use, while enhancing their beneficial actions in the nervous system. Future HRT strategies will also need to take into account the marked individual differences in aging. Indeed, whereas some old individuals exhibit performances similar to those of young subjects, others are severely impaired (288). In addition, one should be aware that steroids such as progesterone are only part of the complex signaling systems, and that they exert their effects in concert with other hormones and growth factors, which all undergo major changes during the aging process (37, 823, 824). This may contribute to the influence of environmental factors and lifestyle choices, which also influence the outcome of hormone therapies (825).

Acknowledgments

Address all correspondence and requests for reprints to: Dr. Michael Schumacher, INSERM UMR 788, 80, rue du Général Leclerc, 94276 Kremlin-Bicêtre, France. E-mail: Michael.Schumacher@kb.inserm.fr
Disclosure Statement: The authors have nothing to declare.

References

1. Turgeon JL, McDonnell DP, Martin KA, Wise PM 2004 Hormone therapy: physiological complexity belies therapeutic simplicity. *Science* 304:1269–1273
2. Sohrabji F 2005 Estrogen: a neuroprotective or proinflammatory hormone? Emerging evidence from reproductive aging models. *Ann NY Acad Sci* 1052:75–90
3. Wise PM, Dubal DB, Rau SW, Brown CM, Suzuki S 2005 Are estrogens protective or risk factors in brain injury and neurodegeneration? Reevaluation after the Women's Health Initiative. *Endocr Rev* 26:308–312
4. Howell N, Dykens J, Moos WH 2006 Alzheimer's disease, estrogens, and clinical trials: a case study in drug development for complex disorders. *Drug Dev Res* 66:53–77
5. Morrison JH, Brinton RD, Schmidt PJ, Gore AC 2006 Estrogen, menopause, and the aging brain: how basic neuroscience can inform hormone therapy in women. *J Neurosci* 26:10332–10348
6. Wiegatz I, Kuhl H 2004 Progestogen therapies: differences in clinical effects? *Trends Endocrinol Metab* 15:277–285
7. Naftolin F, Silver D 2002 Is progestogen supplementation of ERT really necessary? *Menopause* 9:1–2
8. Schumacher M, Akwa Y, Guennoun R, Robert F, Labombarda F, Desarnaud F, Robel P, De Nicola AF, Baulieu EE 2000 Steroid synthesis and metabolism in the nervous system: trophic and protective effects. *J Neurocytol* 29:307–326
9. Schumacher M, Weill-Engerer S, Liere P, Robert F, Franklin RJ, Garcia-Segura LM, Lambert JJ, Mayo W, Melcangi RC, Parducz A, Suter U, Carelli C, Baulieu EE, Akwa Y 2003 Steroid hormones and neurosteroids in normal and pathological aging of the nervous system. *Prog Neurobiol* 71:3–29
10. Schumacher M, Guennoun R, Robert F, Carelli C, Gago N, Ghomari A, Gonzalez Deniselle MC, Gonzalez SL, Ibanez C, Labombarda F, Coirini H, Baulieu EE, De Nicola AF 2004 Local synthesis and dual actions of progesterone in the nervous system: neuroprotection and myelination. *Growth Horm IGF Res* 14 Suppl A:S18–S33
11. De Nicola AF, Gonzalez SL, Labombarda F, Deniselle MC, Garay L, Guennoun R, Schumacher M 2006 Progesterone treatment of spinal cord injury: effects on receptors, neurotrophins, and myelination. *J Mol Neurosci* 28:3–15
12. Wojtal K, Trojnar MK, Czuczwar SJ 2006 Endogenous neuroprotective factors: neurosteroids. *Pharmacol Rep* 58:335–340
13. Stein DG 2005 The case for progesterone. *Ann NY Acad Sci* 1052:152–169
14. Singh M 2005 Mechanisms of progesterone-induced neuroprotection. *Ann NY Acad Sci* 1052:145–151
15. Sudo S, Wen TC, Desaki J, Matsuda S, Tanaka J, Arai T, Maeda N, Sakanaka M 1997 β -Estradiol protects hippocampal CA1 neurons against transient forebrain ischemia in gerbil. *Neurosci Res* 29:345–354
16. McEwen BS 1999 Clinical review 108: the molecular and neuroanatomical basis for estrogen effects in the central nervous system. *J Clin Endocrinol Metab* 84:1790–1797
17. Garcia-Segura LM, Azcoitia I, DonCarlos LL 2001 Neuroprotection by estradiol. *Prog Neurobiol* 63:29–60
18. Wise PM, Dubal DB, Wilson ME, Rau SW 2000 Estradiol is a neuroprotective factor in vivo and in vitro models of brain injury. *J Neurocytol* 29:401–410
19. Wise PM, Dubal DB, Wilson ME, Rau SW, Bottner M, Rosewell KL 2001 Estradiol is a protective factor in the adult and aging brain: understanding of mechanisms derived from in vivo and in vitro studies. *Brain Res Rev* 37:313–319
20. Lee SJ, McEwen BS 2001 Neurotrophic and neuroprotective actions of estrogens and their therapeutic implications. *Annu Rev Pharmacol Toxicol* 41:569–591
21. Behl C 2002 Oestrogen as a neuroprotective hormone. *Nat Rev Neurosci* 3:433–442
22. Wise PM 2002 Estrogens and neuroprotection. *Trends Endocrinol Metab* 13:229–230
23. Garcia-Segura LM, Veiga S, Sierra A, Melcangi RC, Azcoitia I 2003 Aromatase: a neuroprotective enzyme. *Prog Neurobiol* 71:31–41
24. Smith SS, Woolley CS 2004 Cellular and molecular effects of steroid hormones on CNS excitability. *Cleve Clin J Med* 71(Suppl 2):S4–S10
25. Hulley S, Grady D, Bush T, Furberg C, Herrington D, Riggs B, Vittinghoff E 1998 Randomized trial of estrogen plus progestin for secondary prevention of coronary heart disease in postmenopausal women. Heart and Estrogen/Progestin Replacement Study (HERS) Research Group. *JAMA* 280:605–613
26. Viscoli CM, Brass LM, Kernan WN, Sarrel PM, Suissa S, Horwitz RI 2001 A clinical trial of estrogen-replacement therapy after ischemic stroke. *N Engl J Med* 345:1243–1249
27. Rossouw JE, Anderson GL, Prentice RL, Lacroix AZ, Kooperberg C, Stefanick ML, Jackson RD, Beresford SA, Howard BV, Johnson KC, Kotchen JM, Ockene J 2002 Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. *JAMA* 288:321–333
28. Pradhan AD, Manson JE, Rossouw JE, Siscovick DS, Mouton CP, Rifai N, Wallace RB, Jackson RD, Pettinger MB, Ridker PM 2002 Inflammatory biomarkers, hormone replacement therapy, and incident coronary heart disease: prospective analysis from the Women's Health Initiative observational study. *JAMA* 288:980–987
29. Hays J, Ockene JK, Brunner RL, Kotchen JM, Manson JE, Patter-

- son RE, Aragaki AK, Shumaker SA, Brzyski RG, LaCroix AZ, Granek IA, Valanis BG; Women's Health Initiative Investigators 2003 Effects of estrogen plus progestin on health-related quality of life. *N Engl J Med* 348:1839–1854
30. Wassertheil-Smoller S, Hendrix SL, Limacher M, Heiss G, Kooperberg C, Baird A, Kotchen T, Curb JD, Black H, Rossouw JE, Aragaki A, Safford M, Stein E, Laowattana S, Mysiw WJ; WHI Investigators 2003 Effect of estrogen plus progestin on stroke in postmenopausal women: the Women's Health Initiative: a randomized trial. *JAMA* 289:2673–2684
 31. Rapp SR, Espeland MA, Shumaker SA, Henderson VW, Brunner RL, Manson JE, Gass ML, Stefanick ML, Lane DS, Hays J, Johnson KC, Coker LH, Dailey M, Bowen D; WHIMS Investigators 2003 Effect of estrogen plus progestin on global cognitive function in postmenopausal women. *JAMA* 289:2663–2672
 32. Shumaker SA, Legault C, Rapp SR, Thal L, Wallace RB, Ockene JK, Hendrix SL, Jones 3rd BN, Assaf AR, Jackson RD, Kotchen JM, Wassertheil-Smoller S, Wactawski-Wende J; WHIMS Investigators 2003 Effect of estrogen plus progestin and the incidence of dementia and mild cognitive impairment in postmenopausal women. *JAMA* 289:2651–2662
 33. Craig MC, Maki PM, Murphy DG 2005 The Women's Health Initiative Memory Study: findings and implications for treatment. *Lancet Neurol* 4:190–194
 34. Anderson GL, Limacher M, Assaf AR, Bassford T, Beresford SA, Black H, Bonds D, Brunner R, Brzyski R, Caan B, Chlebowski R, Curb D, Gass M, Hays J, Heiss G, Hendrix S, Howard BV, Hsia J, Hubbell A, Jackson R, Johnson KC, Judd H, Kotchen JM, Kuller L, LaCroix AZ, Lane D, Langer RD, Lasser N, Lewis CE, Manson J, Margolis K, Ockene J, O'Sullivan MJ, Phillips L, Prentice RL, Ritenbaugh C, Robbins J, Rossouw JE, Sarto G, Stefanick ML, Van Horn L, Wactawski-Wende J, Wallace R, Wassertheil-Smoller S; Women's Health Initiative Steering Committee 2004 Effects of conjugated equine estrogen in postmenopausal women with hysterectomy. *JAMA* 291:1701–1712
 35. Bath PM, Gray LJ 2005 Association between hormone replacement therapy and subsequent stroke: a meta-analysis. *BMJ* 330:342
 36. Espeland MA, Rapp SR, Shumaker SA, Brunner R, Manson JE, Sherwin BB, Hsia J, Margolis KL, Hogan PE, Wallace R, Dailey M, Freeman R, Hays J 2004 Conjugated equine estrogens and global cognitive function in postmenopausal women: Women's Health Initiative Memory Study. *JAMA* 291:2959–2968
 37. Yaffe K 2003 Hormone therapy and the brain. *JAMA* 289:2717–2719
 38. Hulley SB, Grady D 2004 The WHI estrogen-alone trial - Do things look any better? *JAMA* 291:1769–1771
 39. Utian WH, Archer DF, Gallagher JC, Gass MLS, Gelfand MM, Henderson VW, Hodis HN, Lobo RA, McClung M, Reid RL, Schwartz PE, Stefanick ML, Woods NF 2004 Recommendations for estrogen and progestogen use in peri- and postmenopausal women: October 2004 position statement of The North American Menopause Society. *Menopause* 11:589–600
 40. Azoulay C 2004 Menopause in 2004: "hormone replacement therapy" is not what it used to be anymore. *Rev Med Interne* 25:806–815
 41. Ettinger B, Barrett-Connor E, Hoq LA, Vader JP, Dubois RW 2006 When is it appropriate to prescribe postmenopausal hormone therapy? *Menopause* 13:404–410
 42. Belelli D, Lambert JJ 2005 Neurosteroids: endogenous regulators of the GABA(A) receptor. *Nat Rev Neurosci* 6:565–575
 43. Melcangi RC, Poletti A, Cavarretta I, Celotti F, Colciago A, Magnaghi V, Motta M, Negri C, Martini L 1998 The 5 α -reductase in the central nervous system: expression and modes of control. *J Steroid Biochem Mol Biol* 65:295–299
 44. Patte-Mensah C, Penning TM, Mensah-Nyagan AG 2004 Anatomical and cellular localization of neuroactive 5 α /3 α -reduced steroid-synthesizing enzymes in the spinal cord. *J Comp Neurol* 477:286–299
 45. Penning TM, Jin Y, Steckelbroeck S, Lanisnik RT, Lewis M 2004 Structure-function of human 3 α -hydroxysteroid dehydrogenases: genes and proteins. *Mol Cell Endocrinol* 215:63–72
 46. Belyaeva OV, Kedishvili NY 2006 Comparative genomic and phylogenetic analysis of short-chain dehydrogenases/reductases with dual retinol/sterol substrate specificity. *Genomics* 88:820–830
 47. Bauman DR, Steckelbroeck S, Penning TM 2004 The roles of aldo-keto reductases in steroid hormone action. *Drug News Perspect* 17:563–578
 48. Huang XF, Luu-The V 2000 Molecular characterization of a first human 3($\alpha \rightarrow \beta$)-hydroxysteroid epimerase. *J Biol Chem* 275:29452–29457
 49. Belyaeva OV, Chetyrkin SV, Clark AL, Kostereva NV, Santacruz KS, Chronwall BM, Kedishvili NY 2007 Role of microsomal RoDH-Like short-chain dehydrogenases/reductases in the oxidation and epimerization of 3 α -hydroxysteroids in human tissues. *Endocrinology* 148:2148–2156
 50. Gee KW, Chang WC, Brinton RE, McEwen BS 1987 GABA-dependent modulation of the Cl-ionophore by steroids in rat brain. *Eur J Pharmacol* 136:419–423
 51. Lundgren P, Stromberg J, Bäckström T, Wang M 2003 Allopregnanolone-stimulated GABA-mediated chloride ion flux is inhibited by 3 β -hydroxy-5 α -pregnan-20-one (isoallopregnanolone). *Brain Res* 982:45–53
 52. Bäckström T, Wahlström G, Wahlström K, Zhu D, Wang MD 2005 Isoallopregnanolone; an antagonist to the anaesthetic effect of allopregnanolone in male rats. *Eur J Pharmacol* 512:15–21
 53. Schindler AE, Campagnoli C, Druckmann R, Huber J, Pasqualini JR, Schweppe KW, Thijssen JH 2003 Classification and pharmacology of progestins. *Maturitas* 46(Suppl 1):S7–S16
 54. Sitruk-Ware R 2004 New progestogens: a review of their effects in perimenopausal and postmenopausal women. *Drugs Aging* 21:865–883
 55. Sitruk-Ware R 2004 Pharmacological profile of progestins. *Maturitas* 47:277–283
 56. Bamberger CM, Else T, Bamberger AM, Beil FU, Schulte HM 1999 Dissociative glucocorticoid activity of medroxyprogesterone acetate in normal human lymphocytes. *J Clin Endocrinol Metab* 84:4055–4061
 57. Garcia-Becerra R, Cooney AJ, Borja-Cacho E, Lemus AE, Perez-Palacios G, Larrea F 2004 Comparative evaluation of androgen and progesterone receptor transcription selectivity indices of 19-nortestosterone-derived progestins. *J Steroid Biochem Mol Biol* 91:21–27
 58. Hapgood JP, Koubovec D, Louw A, Africander D 2004 Not all progestins are the same: implications for usage. *Trends Pharmacol Sci* 25:554–557
 59. Morali G, Lemus AE, Munguia R, Garcia GA, Grillasca I, Perez-Palacios G 2002 Hormone-like behavioral effects of levonorgestrel and its metabolites in the male rat. *Pharmacol Biochem Behav* 73:951–961
 60. Garcia-Becerra R, Borja-Cacho E, Cooney AJ, Jackson KJ, Lemus AE, Perez-Palacios G, Larrea F 2002 The intrinsic transcriptional estrogenic activity of a non-phenolic derivative of levonorgestrel is mediated via the estrogen receptor- α . *J Steroid Biochem Mol Biol* 82:333–341
 61. Lemus AE, Enriquez J, Garcia GA, Grillasca I, Perez-Palacios G 1997 5 α -Reduction of norethisterone enhances its binding affinity for androgen receptors but diminishes its androgenic potency. *J Steroid Biochem Mol Biol* 60:121–129
 62. Larrea F, Garcia-Becerra R, Lemus AE, Garcia GA, Perez-Palacios G, Jackson KJ, Coleman KM, Dace R, Smith CL, Cooney AJ 2001 A-ring reduced metabolites of 19-nor synthetic progestins as subtype selective agonists for ER α . *Endocrinology* 142:3791–3799
 63. Garcia-Becerra R, Borja-Cacho E, Cooney AJ, Smith CL, Lemus AE, Perez-Palacios G, Larrea F 2006 Synthetic 19-nortestosterone derivatives as estrogen receptor α subtype-selective ligands induce similar receptor conformational changes and steroid receptor co-activator recruitment than natural estrogens. *J Steroid Biochem Mol Biol* 99:108–114
 64. Winneker RC, Bitran D, Zhang Z 2003 The preclinical biology of a new potent and selective progestin: trimegestone. *Steroids* 68:915–920
 65. Bernardi F, Pieri M, Stomati M, Luisi S, Palumbo M, Pluchino N, Ceccarelli C, Genazzani AR 2003 Effect of different hormonal replacement therapies on circulating allopregnanolone and dehydroepiandrosterone levels in postmenopausal women. *Gynecol Endocrinol* 17:65–77

66. **Rapkin AJ, Biggio G, Concas A** 2006 Oral contraceptives and neuroactive steroids. *Pharmacol Biochem Behav* 84:628–634
67. **Belelli D, Herd MB** 2003 The contraceptive agent Provera enhances GABA(A) receptor-mediated inhibitory neurotransmission in the rat hippocampus: evidence for endogenous neurosteroids? *J Neurosci* 23:10013–10020
68. **Bernardi F, Pluchino N, Pieri M, Begliuomini S, Lenzi E, Puccetti S, Casarosa E, Luisi M, Genazzani AR** 2006 Progesterone and medroxyprogesterone acetate effects on central and peripheral allopregnanolone and β -endorphin levels. *Neuroendocrinology* 83: 348–359
69. **Newmark ME, Penry JK** 1980 Catamenial epilepsy: a review. *Epilepsia* 21:281–300
70. **Bjorn I, Bixo M, Nojd KS, Nyberg S, Bäckström T** 2000 Negative mood changes during hormone replacement therapy: a comparison between two progestogens. *Am J Obstet Gynecol* 183:1419–1426
71. **Cagnacci A, Arangino S, Baldassari F, Alessandrini C, Landi S, Volpe A** 2004 A comparison of the central effects of different progestins used in hormone replacement therapy. *Maturitas* 48: 456–462
72. **Nilsen J, Brinton RD** 2002 Impact of progestins on estrogen-induced neuroprotection: synergy by progesterone and 19-norprogesterone and antagonism by medroxyprogesterone acetate. *Endocrinology* 143:205–212
73. **Nilsen J, Brinton RD** 2003 Divergent impact of progesterone and medroxyprogesterone acetate (Provera) on nuclear mitogen-activated protein kinase signaling. *Proc Natl Acad Sci USA* 100:10506–10511
74. **Nilsen J, Morales A, Brinton RD** 2006 Medroxyprogesterone acetate exacerbates glutamate excitotoxicity. *Gynecol Endocrinol* 22: 355–361
75. **Littleton-Kearney MT, Klaus JA, Hurn PD** 2005 Effects of combined oral conjugated estrogens and medroxyprogesterone acetate on brain infarction size after experimental stroke in rat. *J Cereb Blood Flow Metab* 25:421–426
76. **Pazol K, Wilson ME, Wallen K** 2004 Medroxyprogesterone acetate antagonizes the effects of estrogen treatment on social and sexual behavior in female macaques. *J Clin Endocrinol Metab* 89:2998–3006
77. **Lee TC, Miller WL, Auchus RJ** 1999 Medroxyprogesterone acetate and dexamethasone are competitive inhibitors of different human steroidogenic enzymes. *J Clin Endocrinol Metab* 84:2104–2110
78. **Bhavnani BR** 2003 Estrogens and menopause: pharmacology of conjugated equine estrogens and their potential role in the prevention of neurodegenerative diseases such as Alzheimer's. *J Steroid Biochem Mol Biol* 85:473–482
79. **Berco M, Bhavnani BR** 2001 Differential neuroprotective effects of equine estrogens against oxidized low density lipoprotein-induced neuronal cell death. *J Soc Gynecol Investig* 8:245–254
80. **Zhang Y, Bhavnani BR** 2005 Glutamate-induced apoptosis in primary cortical neurons is inhibited by equine estrogens via down-regulation of caspase-3 and prevention of mitochondrial cytochrome c release. *BMC Neurosci* 6:1–23
81. **Zhao L, Brinton RD** 2006 Select estrogens within the complex formulation of conjugated equine estrogens (Premarin) are protective against neurodegenerative insults: implications for a composition of estrogen therapy to promote neuronal function and prevent Alzheimer's disease. *BMC Neurosci* 7:1–13
82. **de Lignieres B, de Vathaire F, Fournier S, Urbinelli R, Allaert F, Le MG, Kuttann F** 2002 Combined hormone replacement therapy and risk of breast cancer in a French cohort study of 3175 women. *Climacteric* 5:332–340
83. **Sitruk-Ware R** 2002 Progestogens in hormonal replacement therapy: new molecules, risks, and benefits. *Menopause* 9:6–15
84. **Campagnoli C, Abba C, Ambroggio S, Peris C** 2005 Pregnancy, progesterone and progestins in relation to breast cancer risk. *J Steroid Biochem Mol Biol* 97:441–450
85. **de Lignieres B** 1999 Oral micronized progesterone. *Clin Ther* 21: 41–60
86. 1995 Effects of estrogen or estrogen/progestin regimens on heart disease risk factors in postmenopausal women. The Postmenopausal Estrogen/Progestin Interventions (PEPI) Trial. The Writing Group for the PEPI Trial. *JAMA [Erratum (1995) 274:1676]* 273: 199–208
87. **Judd HL, Mebane-Sims I, Legault C, Wasilaukas C, Johnson S, Merino M, Barrett-Connor E, Trabala J** 1996 Effects of hormone replacement therapy on endometrial histology in postmenopausal women: the postmenopausal estrogen/progestin interventions (PEPI) trial. *JAMA* 275:370–375
88. **Dennerstein L, Morse C, Gotts G, Brown J, Smith M, Oats J, Burrows G** 1986 Treatment of premenstrual syndrome. A double-blind trial of dydrogesterone. *J Affect Disord* 11:199–205
89. **de Lignieres B, Vincens M** 1982 Differential effects of exogenous oestradiol and progesterone on mood in postmenopausal women: individual dose/effect relationship. *Maturitas* 4:67–72
90. **Fitzpatrick LA, Pace C, Wiita B** 2000 Comparison of regimens containing oral micronized progesterone or medroxyprogesterone acetate on quality of life in postmenopausal women: a cross-sectional survey. *J Womens Health Gend Based Med* 9:381–387
91. **Andreen L, Sundström-Poromaa I, Bixo M, Andersson A, Nyberg S, Bäckström T** 2005 Relationship between allopregnanolone and negative mood in postmenopausal women taking sequential hormone replacement therapy with vaginal progesterone. *Psychoneuroendocrinology* 30:212–224
92. **Andreen L, Sundström-Poromaa I, Bixo M, Nyberg S, Bäckström T** 2006 Allopregnanolone concentration and mood—a bimodal association in postmenopausal women treated with oral progesterone. *Psychopharmacology (Berl)* 187:209–221
93. **Gonzalez-Vidal MD, Cervera-Gaviria M, Ruelas R, Escobar A, Morali G, Cervantes M** 1998 Progesterone: protective effects on the rat hippocampal neuronal damage due to acute global cerebral ischemia. *Arch Med Res* 29:117–124
94. **Xu L, Sapolsky RM, Giffard RG** 2001 Differential sensitivity of murine astrocytes and neurons from different brain regions to injury. *Exp Neurol* 169:416–424
95. **Cervantes M, Gonzalez-Vidal MD, Ruelas R, Escobar A, Morali G** 2002 Neuroprotective effects of progesterone on damage elicited by acute global cerebral ischemia in neurons of the caudate nucleus. *Arch Med Res* 33:6–14
96. **Jiang N, Chopp M, Stein D, Feit H** 1996 Progesterone is neuroprotective after transient middle cerebral artery occlusion in male rats. *Brain Res* 735:101–107
97. **Kumon Y, Kim SC, Tompkins P, Stevens A, Sakaki S, Loftus CM** 2000 Neuroprotective effect of postischemic administration of progesterone in spontaneously hypertensive rats with focal cerebral ischemia. *J Neurosurg* 92:848–852
98. **Gibson CL, Murphy SP** 2004 Progesterone enhances functional recovery after middle cerebral artery occlusion in male mice. *J Cereb Blood Flow Metab* 24:805–813
99. **Hoffman SW, Fulop Z, Stein DG** 1994 Bilateral frontal cortical contusion in rats: behavioral and anatomic consequences. *J Neurotrauma* 11:417–431
100. **Stein DG** 2001 Brain damage, sex hormones and recovery: a new role for progesterone and estrogen? *Trends Neurosci* 24:386–391
101. **He J, Evans CO, Hoffman SW, Oyesiku NM, Stein DG** 2004 Progesterone and allopregnanolone reduce inflammatory cytokines after traumatic brain injury. *Exp Neurol* 189:404–412
102. **Roof RL, Duvdevani R, Stein DG** 1993 Gender influences outcome of brain injury—progesterone plays a protective role. *Brain Res* 607:333–336
103. **Roof RL, Duvdevani R, Braswell L, Stein DG** 1994 Progesterone facilitates cognitive recovery and reduces secondary neuronal loss caused by cortical contusion injury in male rats. *Exp Neurol* 129: 64–69
104. **Wright DW, Ritchie JC, Mullins RE, Kellermann AL, Denson DD** 2005 Steady-state serum concentrations of progesterone following continuous intravenous infusion in patients with acute moderate to severe traumatic brain injury. *J Clin Pharmacol* 45:640–648
105. **Wright DW, Kellermann AL, Hertzberg VS, Clark PL, Frankel M, Goldstein FC, Salomone JP, Dent LL, Harris OA, Ander DS, Lowery DW, Patel MM, Denson DD, Gordon AB, Wald MM, Gupta S, Hoffman SW, Stein DG** 2007 ProTECT: a randomized clinical trial of progesterone for acute traumatic brain injury. *Ann Emerg Med* 49:391–402
106. **Chen J, Chopp M, Li Y** 1999 Neuroprotective effects of proges-

- terone after transient middle cerebral artery occlusion in rat. *J Neurol Sci* 171:24–30
107. Roof RL, Duvdevani R, Heyburn JW, Stein DG 1996 Progesterone rapidly decreases brain edema: treatment delayed up to 24 hours is still effective. *Exp Neurol* 138:246–251
 108. Robertson CL, Puskar A, Hoffman GE, Murphy AZ, Saraswati M, Fiskum G 2006 Physiologic progesterone reduces mitochondrial dysfunction and hippocampal cell loss after traumatic brain injury in female rats. *Exp Neurol* 197:235–243
 109. Galani R, Hoffman SW, Stein DG 2001 Effects of the duration of progesterone treatment on the resolution of cerebral edema induced by cortical contusions in rats. *Restor Neurol Neurosci* 18:161–166
 110. Cutler SM, Vanlandingham JW, Murphy AZ, Stein DG 2006 Slow-release and injected progesterone treatments enhance acute recovery after traumatic brain injury. *Pharmacol Biochem Behav* 84:420–428
 111. Vanlandingham JW, Cutler SM, Virmani S, Hoffman SW, Covey DF, Krishnan K, Hammes SR, Jamnongjit M, Stein DG 2006 The enantiomer of progesterone acts as a molecular neuroprotectant after traumatic brain injury. *Neuropharmacology* 51:1078–1085
 112. Akwa Y, Ladurelle N, Covey DF, Baulieu EE 2001 The synthetic enantiomer of pregnenolone sulfate is very active on memory in rats and mice, even more so than its physiological neurosteroid counterpart: distinct mechanisms? *Proc Natl Acad Sci USA* 98:14033–14037
 113. Simpkins JW, Yang SH, Liu R, Perez E, Yun CZ, Covey DF, Green PS 2004 Estrogen-like compounds for ischemic neuroprotection. *Stroke* 35:2648–2651
 114. Covey DF, Nathan D, Kalkbrenner M, Nilsson KR, Hu Y, Zorumski CF, Evers AS 2000 Enantioselectivity of pregnanolone-induced γ -aminobutyric acid(A) receptor modulation and anesthesia. *J Pharmacol Exp Ther* 293:1009–1016
 115. Auchus RJ, Sampath KA, Andrew BC, Gupta MK, Bruce K, Rath NP, Covey DF 2003 The enantiomer of progesterone (ent-progesterone) is a competitive inhibitor of human cytochromes P450c17 and P450c21. *Arch Biochem Biophys* 409:134–144
 116. Callier S, Morissette M, Grandbois M, Pelaprat D, Di P 2001 Neuroprotective properties of 17 β -estradiol, progesterone, and raloxifene in MPTP C57Bl/6 mice. *Synapse* 41:131–138
 117. Ragonese P, D'Amelio M, Salemi G, Aridon P, Gammino M, Epifanio A, Morgante L, Savettieri G 2004 Risk of Parkinson disease in women: effect of reproductive characteristics. *Neurology* 62:2010–2014
 118. Thomas AJ, Nockels RP, Pan HQ, Shaffrey CI, Chopp M 1999 Progesterone is neuroprotective after acute experimental spinal cord trauma in rats. *Spine* 24:2134–2138
 119. Gonzalez Deniselle MC, Garay L, Gonzalez S, Guennoun R, Schumacher M, De Nicola AF 2005 Progesterone restores retrograde labeling of cervical motoneurons in Wobbler mouse motoneuron disease. *Exp Neurol* 195:518–523
 120. De Nicola AF, Labombarda F, Gonzalez SL, Gonzalez Deniselle MC, Guennoun R, Schumacher M 2003 Steroid effects on glial cells: detrimental or protective for spinal cord injury? *Ann NY Acad Sci* 1007:317–328
 121. Labombarda F, Gonzalez SL, Gonzalez DM, Guennoun R, Schumacher M, De Nicola AF 2002 Cellular basis for progesterone neuroprotection in the injured spinal cord. *J Neurotrauma* 19:343–355
 122. Gonzalez SL, Labombarda F, Gonzalez Deniselle MC, Guennoun R, Schumacher M, De Nicola AF 2004 Progesterone up-regulates neuronal brain-derived neurotrophic factor expression in the injured spinal cord. *Neuroscience* 125:605–614
 123. Schmitt-John T, Drepper C, Mussmann A, Hahn P, Kuhlmann M, Thiel C, Hafner M, Lengeling A, Heimann P, Jones JM, Meisler MH, Jockusch H 2005 Mutation of Vps54 causes motor neuron disease and defective spermiogenesis in the wobbler mouse. *Nat Genet* 37:1213–1215
 124. Duchon LW, Strich SJ 1968 An hereditary motor neurone disease with progressive denervation of muscle in the mouse: the mutant 'wobbler'. *J Neurol Neurosurg Psychiatry* 31:535–542
 125. Price DL, Cleveland DW, Koliatsos VE 1994 Motor neurone disease and animal models. *Neurobiol Dis* 1:3–11
 126. Gonzalez Deniselle MC, Gonzalez S, De Nicola AF 2001 Cellular basis of steroid neuroprotection in the Wobbler mouse, a model of motoneuron disease. *Cell Mol Neurobiol* 21:237–254
 127. Gonzalez Deniselle MC, Lopez Costa JJ, Gonzalez SL, Labombarda F, Garay L, Guennoun R, Schumacher M, De Nicola AF 2002 Basis of progesterone protection in spinal cord neurodegeneration. *J Steroid Biochem Mol Biol* 83:199–209
 128. Gonzalez Deniselle C, Lopez Costa JJ, Saavedra JP, Pietranera L, Gonzalez SL, Garay L, Guennoun R, Schumacher M, De Nicola AF 2002 Progesterone neuroprotection in the Wobbler mouse, a genetic model of spinal cord motor neuron disease. *Neurobiol Dis* 11:457–468
 129. Toung TJ, Chen TY, Littleton-Kearney MT, Hurn PD, Murphy SJ 2004 Effects of combined estrogen and progesterone on brain infarction in reproductively senescent female rats. *J Cereb Blood Flow Metab* 24:1160–1166
 130. Murphy SJ, Traystman RJ, Hurn PD, Duckles SP 2000 Progesterone exacerbates striatal stroke injury in progesterone-deficient female animals. *Stroke* 31:1173–1178
 131. Goss CW, Hoffman SW, Stein DG 2003 Behavioral effects and anatomic correlates after brain injury: a progesterone dose-response study. *Pharmacol Biochem Behav* 76:231–242
 132. Hall ED, Gibson TR, Pavel KM 2005 Lack of a gender difference in post-traumatic neurodegeneration in the mouse controlled cortical impact injury model. *J Neurotrauma* 22:669–679
 133. Hall ED, Sullivan PG, Gibson TR, Pavel KM, Thompson BM, Scheff SW 2005 Spatial and temporal characteristics of neurodegeneration after controlled cortical impact in mice: more than a focal brain injury. *J Neurotrauma* 22:252–265
 134. Rosario ER, Ramsden M, Pike CJ 2006 Progestins inhibit the neuroprotective effects of estrogen in rat hippocampus. *Brain Res* 1099:206–210
 135. Li W, Hoffman PN, Stirling W, Price DL, Lee MK 2004 Axonal transport of human α -synuclein slows with aging but is not affected by familial Parkinson's disease-linked mutations. *J Neurochem* 88:401–410
 136. Uchida A, Tashiro T, Komiya Y, Yorifuji H, Kishimoto T, Hisanaga S 2004 Morphological and biochemical changes of neurofilaments in aged rat sciatic nerve axons. *J Neurochem* 88:735–745
 137. Niewiadomska G, Baksalerska-Pazera M, Riedel G 2005 Altered cellular distribution of phospho-tau proteins coincides with impaired retrograde axonal transport in neurons of aged rats. *Ann NY Acad Sci* 1048:287–295
 138. Mukaetova-Ladinska EB, McKeith IG 2006 Pathophysiology of synuclein aggregation in Lewy body disease. *Mech Ageing Dev* 127:188–202
 139. Stokin GB, Lillo C, Falzone TL, Brusch RG, Rockenstein E, Mount SL, Raman R, Davies P, Masliah E, Williams DS, Goldstein LS 2005 Axonopathy and transport deficits early in the pathogenesis of Alzheimer's disease. *Science* 307:1282–1288
 140. Roof RL, Hoffman SW, Stein DG 1997 Progesterone protects against lipid peroxidation following traumatic brain injury in rats. *Mol Chem Neurobiol* 31:1–11
 141. Niki E, Yoshida Y, Saito Y, Noguchi N 2005 Lipid peroxidation: mechanisms, inhibition, and biological effects. *Biochem Biophys Res Commun* 338:668–676
 142. Pratico D 2002 Lipid peroxidation and the aging process. *Sci Aging Knowledge Environ* 2002:re5
 143. Pratico D, Sung S 2004 Lipid peroxidation and oxidative imbalance: early functional events in Alzheimer's disease. *J Alzheimers Dis* 6:171–175
 144. Moorthy K, Sharma D, Basir SF, Baquer NZ 2005 Administration of estradiol and progesterone modulate the activities of antioxidant enzyme and aminotransferases in naturally menopausal rats. *Exp Gerontol* 40:295–302
 145. Moorthy K, Yadav UC, Siddiqui MR, Mantha AK, Basir SF, Sharma D, Cowsik SM, Baquer NZ 2005 Effect of hormone replacement therapy in normalizing age-related neuronal markers in different age groups of naturally menopausal rats. *Biogerontology* 6:345–356
 146. Harman D 1972 A biological clock: the mitochondria? *J Am Geriatr Soc* 20:145–147
 147. Kokoszka JE, Coskun P, Esposito LA, Wallace DC 2001 Increased

- mitochondrial oxidative stress in the Sod2 (+/–) mouse results in the age-related decline of mitochondrial function culminating in increased apoptosis. *Proc Natl Acad Sci USA* 98:2278–2283
148. **Brand MD, Buckingham JA, Esteves TC, Green K, Lambert AJ, Miwa S, Murphy MP, Pakay JL, Talbot DA, Echtay KS** 2004 Mitochondrial superoxide and aging: uncoupling-protein activity and superoxide production. *Biochem Soc Symp* 71:203–213
 149. **Loeb LA, Wallace DC, Martin GM** 2005 The mitochondrial theory of aging and its relationship to reactive oxygen species damage and somatic mtDNA mutations. *Proc Natl Acad Sci USA* 102:18769–18770
 150. **Schriner SE, Linford NJ, Martin GM, Treuting P, Ogburn CE, Emond M, Coskun PE, Ladiges W, Wolf N, Van RH, Wallace DC, Rabinovitch PS** 2005 Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* 308:1909–1911
 151. **Melov S** 2004 Modeling mitochondrial function in aging neurons. *Trends Neurosci* 27:601–606
 152. **Beal MF** 2005 Mitochondria take center stage in aging and neurodegeneration. *Ann Neurol* 58:495–505
 153. **Toescu EC** 2005 Normal brain ageing: models and mechanisms. *Philos Trans R Soc Lond B Biol Sci* 360:2347–2354
 154. **Djebaili M, Guo Q, Pettus EH, Hoffman SW, Stein DG** 2005 The neurosteroids progesterone and allopregnanolone reduce cell death, gliosis, and functional deficits after traumatic brain injury in rats. *J Neurotrauma* 22:106–118
 155. **Yao XL, Liu J, Lee E, Ling GS, McCabe JT** 2005 Progesterone differentially regulates pro- and anti-apoptotic gene expression in cerebral cortex following traumatic brain injury in rats. *J Neurotrauma* 22:656–668
 156. **Schlesinger PH, Saito M** 2006 The Bax pore in liposomes, Biophysics. *Cell Death Differ* 13:1403–1408
 157. **Chen JQ, Yager JD, Russo J** 2005 Regulation of mitochondrial respiratory chain structure and function by estrogens/estrogen receptors and potential physiological/pathophysiological implications. *Biochim Biophys Acta* 1746:1–17
 158. **Stirone C, Duckles SP, Krause DN, Procaccio V** 2005 Estrogen increases mitochondrial efficiency and reduces oxidative stress in cerebral blood vessels. *Mol Pharmacol* 68:959–965
 159. **Duckles SP, Krause DN, Stirone C, Procaccio V** 2006 Estrogen and mitochondria: a new paradigm for vascular protection? *Mol Interv* 6:26–35
 160. **Psarra AM, Solakidi S, Sekeris CE** 2006 The mitochondrion as a primary site of action of steroid and thyroid hormones: presence and action of steroid and thyroid hormone receptors in mitochondria of animal cells. *Mol Cell Endocrinol* 246:21–33
 161. **Pedram A, Razandi M, Wallace DC, Levin ER** 2006 Functional estrogen receptors in the mitochondria of breast cancer cells. *Mol Biol Cell* 17:2125–2137
 162. **Guo Q, Sayeed I, Baronne LM, Hoffman SW, Guennoun R, Stein DG** 2006 Progesterone administration modulates AQP4 expression and edema after traumatic brain injury in male rats. *Exp Neurol* 198:469–478
 163. **Garcia-Segura LM, Cardona-Gomez GP, Naftolin F, Chowen JA** 1998 Estradiol upregulates BCL-2 expression in adult brain neurons. *Neuroreport* 9:593–597
 164. **Alkayed NJ, Goto S, Sugo N, Joh HD, Klaus J, Crain BJ, Bernard O, Traystman RJ, Hurn PD** 2001 Estrogen and Bcl-2: gene induction and effect of transgene in experimental stroke. *J Neurosci* 21:7543–7550
 165. **Wise PM** 2006 Estrogen therapy: does it help or hurt the adult and aging brain? Insights derived from animal models. *Neuroscience* 138:831–835
 166. **Miller L, Hunt JS** 1998 Regulation of TNF- α production in activated mouse macrophages by progesterone. *J Immunol* 160:5098–5104
 167. **Drew PD, Chavis JA** 2000 Female sex steroids: effects upon microglial cell activation. *J Neuroimmunol* 111:77–85
 168. **Pettus EH, Wright DW, Stein DG, Hoffman SW** 2005 Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. *Brain Res* 1049:112–119
 169. **Gonzalez SL, Labombarda F, Deniselle MC, Mouguel A, Guennoun R, Schumacher M, De Nicola AF** 2005 Progesterone neuroprotection in spinal cord trauma involves up-regulation of brain-derived neurotrophic factor in motoneurons. *J Steroid Biochem Mol Biol* 94:143–149
 170. **Scharfman HE, MacLusky NJ** 2005 Similarities between actions of estrogen and BDNF in the hippocampus: coincidence or clue? *Trends Neurosci* 28:79–85
 171. **Ogata T, Nakamura Y, Tsuji K, Shibata T, Kataoka K** 1993 Steroid hormones protect spinal cord neurons from glutamate toxicity. *Neuroscience* 55:445–449
 172. **Goodman Y, Bruce AJ, Cheng B, Mattson MP** 1996 Estrogens attenuate and corticosterone exacerbates excitotoxicity, oxidative injury, and amyloid β -peptide toxicity in hippocampal neurons. *J Neurochem* 66:1836–1844
 173. **Yin X, Crawford TO, Griffin JW, Tu P, Lee VM, Li C, Roder J, Trapp BD** 1998 Myelin-associated glycoprotein is a myelin signal that modulates the caliber of myelinated axons. *J Neurosci* 18:1953–1962
 174. **Faulkner J, Keirstead HS** 2005 Human embryonic stem cell-derived oligodendrocyte progenitors for the treatment of spinal cord injury. *Transpl Immunol* 15:131–142
 175. **Koenig HL, Schumacher M, Ferzaz B, Do Thi AN, Ressousches A, Guennoun R, Jung-Testas I, Robel P, Akwa Y, Baulieu EE** 1995 Progesterone synthesis and myelin formation by Schwann cells. *Science* 268:1500–1503
 176. **Chan JR, Phillips LJ, Glaser M** 1998 Glucocorticoids and progestins signal the initiation and enhance the rate of myelin formation. *Proc Natl Acad Sci USA* 95:10459–10464
 177. **Jung-Testas I, Schumacher M, Robel P, Baulieu EE** 1996 Demonstration of progesterone receptors in rat Schwann cells. *J Steroid Biochem Mol Biol* 58:77–82
 178. **Magnaghi V, Cavarretta I, Galbiati M, Martini L, Melcangi RC** 2001 Neuroactive steroids and peripheral myelin proteins. *Brain Res Rev* 37:360–371
 179. **Chan JR, Rodriguez-Waitkus PM, Ng BK, Liang P, Glaser M** 2000 Progesterone synthesized by Schwann cells during myelin formation regulates neuronal gene expression. *Mol Biol Cell* 11:2283–2295
 180. **Groyer G, Eychenne B, Girard C, Rajkowski K, Schumacher M, Cadepond F** 2006 Expression and functional state of the corticosteroid receptors and 11 β -hydroxysteroid dehydrogenase type 2 in Schwann cells. *Endocrinology* 147:4339–4350
 181. **Sereda MW, Meyer Z, Suter U, Uzma N, Nave KA** 2003 Therapeutic administration of progesterone antagonist in a model of Charcot-Marie-Tooth disease (CMT-1A). *Nat Med* 9:1533–1537
 182. **Leonelli E, Bianchi R, Cavaletti G, Caruso D, Crippa D, Garcia-Segura LM, Lauria G, Magnaghi V, Roglio I, Melcangi RC** 2007 Progesterone and its derivatives are neuroprotective agents in experimental diabetic neuropathy: a multimodal analysis. *Neuroscience* 144:1293–1304
 183. **Lubetzki C, Demerens C, Anglade P, Villarroya H, Frankfurter A, Lee VM, Zalc B** 1993 Even in culture, oligodendrocytes myelinate solely axons. *Proc Natl Acad Sci USA* 90:6820–6824
 184. **Baumann N, Pham-Dinh D** 2001 Biology of oligodendrocyte and myelin in the mammalian central nervous system. *Physiol Rev* 81:871–927
 185. **Ghoumari AM, Ibanez C, el-Etr M, Leclerc P, Eychenne B, O'Malley BW, Baulieu EE, Schumacher M** 2003 Progesterone and its metabolites increase myelin basic protein expression in organotypic slice cultures of rat cerebellum. *J Neurochem* 86:848–859
 186. **Dusart I, Airaksinen MS, Sotelo C** 1997 Purkinje cell survival and axonal regeneration are age dependent: an in vitro study. *J Neurosci* 17:3710–3726
 187. **Ghoumari AM, Wehrle R, De Zeeuw CI, Sotelo C, Dusart I** 2002 Inhibition of protein kinase C prevents Purkinje cell death but does not affect axonal regeneration. *J Neurosci* 22:3531–3542
 188. **Notterpek LM, Bullock PN, Malek H, Fisher R, Rome LH** 1993 Myelination in cerebellar slice cultures: development of a system amenable to biochemical analysis. *J Neurosci Res* 36:621–634
 189. **Ukena K, Honda Y, Inai Y, Kohchi C, Lea RW, Tsutsui K** 1999 Expression and activity of 3 β -hydroxysteroid dehydrogenase/ Δ 5- Δ 4-isomerase in different regions of the avian brain. *Brain Res* 818:536–542
 190. **Ghoumari AM, Baulieu EE, Schumacher M** 2005 Progesterone increases oligodendroglial cell proliferation in rat cerebellar slice cultures. *Neuroscience* 135:47–58

191. Jung-Testas I, Hu ZY, Baulieu EE, Robel P 1989 Neurosteroids: biosynthesis of pregnenolone and progesterone in primary cultures of rat glial cells. *Endocrinology* 125:2083–2091
192. Marin-Husstege M, Muggironi M, Raban D, Skoff RP, Casaccia-Bonnel P 2004 Oligodendrocyte progenitor proliferation and maturation is differentially regulated by male and female sex steroid hormones. *Dev Neurosci* 26:245–254
193. Ibanez C, Shields SA, el-Etr M, Baulieu EE, Schumacher M, Franklin RJ 2004 Systemic progesterone administration results in a partial reversal of the age-associated decline in CNS remyelination following toxin-induced demyelination in male rats. *Neuropathol Appl Neurobiol* 30:80–89
194. Labombarda F, Gonzalez S, Deniselle MC, Garay L, Guennoun R, Schumacher M, Nicola AF 2006 Progesterone increases the expression of myelin basic protein and the number of cells showing NG(2) immunostaining in the lesioned spinal cord. *J Neurotrauma* 23:181–192
195. Peters A 1996 Age-related changes in oligodendrocytes in monkey cerebral cortex. *J Comp Neurol* 371:153–163
196. Peters A 2002 Structural changes in the normally aging cerebral cortex of primates. *Prog Brain Res* 136:455–465
197. Peters A, Moss MB, Sethares C 2000 Effects of aging on myelinated nerve fibers in monkey primary visual cortex. *J Comp Neurol* 419:364–376
198. Peters A, Sethares C, Killiany RJ 2001 Effects of age on the thickness of myelin sheaths in monkey primary visual cortex. *J Comp Neurol* 435:241–248
199. Sandell JH, Peters A 2001 Effects of age on nerve fibers in the rhesus monkey optic nerve. *J Comp Neurol* 429:541–553
200. Tang Y, Nyengaard JR, Pakkenberg B, Gundersen HJ 1997 Age-induced white matter changes in the human brain: a stereological investigation. *Neurobiol Aging* 18:609–615
201. O'Sullivan M, Jones DK, Summers PE, Morris RG, Williams SC, Markus HS 2001 Evidence for cortical “disconnection” as a mechanism of age-related cognitive decline. *Neurology* 57:632–638
202. Pittella JEH 1997 Changes in white matter. In: Dani SU, Hori A, Walter GF, eds. *Principles of neural aging*. Amsterdam: Elsevier; 285–295
203. Uchida Y, Tomonaga M, Nomura K 1986 Age-related changes of myelin proteins in the rat peripheral nervous system. *J Neurochem* 46:1376–1381
204. Melcangi RC, Magnaghi V, Cavarretta I, Martini L, Piva F 1998 Age-induced decrease of glycoprotein Po and myelin basic protein gene expression in the rat sciatic nerve. Repair by steroid derivatives. *Neuroscience* 85:569–578
205. Melcangi RC, Magnaghi V, Cavarretta I, Riva MA, Piva F, Martini L 1998 Effects of steroid hormones on gene expression of glial markers in the central and peripheral nervous system: variations induced by aging. *Exp Gerontol* 33:827–836
206. Melcangi RC, Magnaghi V, Martini L 2000 Aging in peripheral nerves: regulation of myelin protein genes by steroid hormones. *Prog Neurobiol* 60:291–308
207. Peters A, Sethares C 2003 Is there remyelination during aging of the primate central nervous system? *J Comp Neurol* 460:238–254
208. Gilson J, Blakemore WF 1993 Failure of remyelination in areas of demyelination produced in the spinal cord of old rats. *Neuropathol Appl Neurobiol* 19:173–181
209. Shields SA, Gilson JM, Blakemore WF, Franklin RJ 1999 Remyelination occurs as extensively but more slowly in old rats compared to young rats following gliotoxin-induced CNS demyelination. *Glia* 28:77–83
210. Li WW, Penderis J, Zhao C, Schumacher M, Franklin RJM 2006 Females remyelinate more efficiently than males following demyelination in the aged but not young adult CNS. *Exp Neurol* 202: 250–254
211. Hinks GL, Franklin RJM 2000 Delayed changes in growth factor gene expression during slow remyelination in the CNS of aged rats. *Mol Cell Neurosci* 16:542–556
212. Sim FJ, Zhao C, Penderis J, Franklin RJM 2002 The age-related decrease in CNS remyelination efficiency is attributable to an impairment of both oligodendrocyte progenitor recruitment and differentiation. *J Neurosci* 22:2451–2459
213. Jones SJ 1993 Visual evoked potentials after optic neuritis. *J Neurol* 240:489–494
214. Confavreux C, Vukusic S 2006 Age at disability milestones in multiple sclerosis. *Brain* 129:595–605
215. Morrison JH, Hof PR 1997 Life and death of neurons in the aging brain. *Science* 278:412–419
216. West MJ 1999 Stereological methods for estimating the total number of neurons and synapses: issues of precision and bias. *Trends Neurosci* 22:51–61
217. Abel TW, Rance NE 2000 Stereologic study of the hypothalamic infundibular nucleus in young and older women. *J Comp Neurol* 424:679–688
218. Escobar CM, Krajewski SJ, Sandoval-Guzman T, Voytko ML, Rance NE 2004 Neuropeptide Y gene expression is increased in the hypothalamus of older women. *J Clin Endocrinol Metab* 89:2338–2343
219. Hof PR, Morrison JH 2004 The aging brain: morphomolecular senescence of cortical circuits. *Trends Neurosci* 27:607–613
220. Rasmussen T, Schliemann T, Sorensen JC, Zimmer J, West MJ 1996 Memory impaired aged rats: no loss of principal hippocampal and subicular neurons. *Neurobiol Aging* 17:143–147
221. Swaab DF, Dubelaar EJ, Scherder EJ, van Someren EJ, Verwer RW 2003 Therapeutic strategies for Alzheimer disease: focus on neuronal reactivation of metabolically impaired neurons. *Alzheimer Dis Assoc Disord* 17(Suppl 4):S114–S122
222. Mesulam MM 1999 Neuroplasticity failure in Alzheimer's disease: bridging the gap between plaques and tangles. *Neuron* 24:521–529
223. Selkoe DJ 2002 Alzheimer's disease is a synaptic failure. *Science* 298:789–791
224. West MJ, Kawas CH, Stewart WF, Rudow GL, Troncoso JC 2004 Hippocampal neurons in pre-clinical Alzheimer's disease. *Neurobiol Aging* 25:1205–1212
225. West MJ 1993 Regionally specific loss of neurons in the aging human hippocampus. *Neurobiol Aging* 14:287–293
226. Woodruff-Pak DS, Trojanowski JQ 1996 The older rabbit as an animal model: implications for Alzheimer's disease. *Neurobiol Aging* 17:283–290
227. Woodruff-Pak DS 2001 Eyeblink classical conditioning differentiates normal aging from Alzheimer's disease. *Integr Physiol Behav Sci* 36:87–108
228. Reynolds ML, Woolf CJ 1993 Reciprocal Schwann cell-axon interactions. *Curr Opin Neurobiol* 3:683–693
229. Snipes GJ, Suter U 1994 Signaling pathways mediating axon-Schwann cell interactions. *Trends Neurosci* 17:399–401
230. Demerens C, Stankoff B, Logak M, Anglade P, Allinquant B, Couraud F, Zalc B, Lubetzki C 1996 Induction of myelination in the central nervous system by electrical activity. *Proc Natl Acad Sci USA* 93:9887–9892
231. Fernandez PA, Tang DG, Cheng L, Prochiantz A, Mudge AW, Raff MC 2000 Evidence that axon-derived neuregulin promotes oligodendrocyte survival in the developing rat optic nerve. *Neuron* 28:81–90
232. Finch CE 2003 Neurons, glia, and plasticity in normal brain aging. *Neurobiol Aging* 24(Suppl 1):S123–S127
233. Rozovsky I, Wei M, Morgan TE, Finch CE 2005 Reversible age impairments in neurite outgrowth by manipulations of astrocytic GFAP. *Neurobiol Aging* 26:705–715
234. Menet V, Ribotta M, Chauvet N, Drian MJ, Lannoy J, Colucci-Guyon E, Privat A 2001 Inactivation of the glial fibrillary acidic protein gene, but not that of vimentin, improves neuronal survival and neurite growth by modifying adhesion molecule expression. *J Neurosci* 21:6147–6158
235. Ribotta MG, Menet V, Privat A 2004 Glial scar and axonal regeneration in the CNS: lessons from GFAP and vimentin transgenic mice. *Acta Neurochir Suppl* 89:87–92
236. Garcia-Segura LM, McCarthy MM 2004 Minireview: Role of glia in neuroendocrine function. *Endocrinology* 145:1082–1086
237. Dhandapani KM, Brann DW 2007 Role of astrocytes in estrogen-mediated neuroprotection. *Exp Gerontol* 42:70–75
238. Rozovsky I, Wei M, Stone DJ, Zanjani H, Anderson CP, Morgan TE, Finch CE 2002 Estradiol (E2) enhances neurite outgrowth by repressing glial fibrillary acidic protein expression and reorganizing laminin. *Endocrinology* 143:636–646

239. Garcia-Estrada J, Del Rio JA, Luquin S, Soriano E, Garcia-Segura LM 1993 Gonadal hormones down-regulate reactive gliosis and astrocyte proliferation after a penetrating brain injury. *Brain Res* 628:271–278
240. Garcia-Estrada J, Luquin S, Fernandez AM, Garcia-Segura LM 1999 Dehydroepiandrosterone, pregnenolone and sex steroids down-regulate reactive astroglia in the male rat brain after a penetrating brain injury. *Int J Dev Neurosci* 17:145–151
241. Labombarda F, Gonzalez S, Roig P, Lima A, Guennoun R, Schumacher M, De Nicola AF 2000 Modulation of NADPH-diaphorase and glial fibrillary acidic protein by progesterone in astrocytes from normal and injured rat spinal cord. *J Steroid Biochem Mol Biol* 73:159–169
242. Grossman KJ, Goss CW, Stein DG 2004 Effects of progesterone on the inflammatory response to brain injury in the rat. *Brain Res* 1008:29–39
243. Kimura D 2000 Sex and cognition. Cambridge, MA: MIT Press
244. Meador KJ, Loring DW, Ray PG, Helman SW, Vazquez BR, Neveu PJ 2004 Role of cerebral lateralization in control of immune processes in humans. *Ann Neurol* 55:840–844
245. Becker JB, Arnold AP, Berkley KJ, Blaustein JD, Eckel LA, Hampson E, Herman JP, Marts S, Sadee W, Steiner M, Taylor J, Young E 2005 Strategies and methods for research on sex differences in brain and behavior. *Endocrinology* 146:1650–1673
246. Raisman G, Field PM 1971 Sexual dimorphism in the preoptic area of the rat. *Science* 173:731–733
247. Arnold AP, Gorski RA 1984 Gonadal steroid induction of structural sex differences in the central nervous system. *Annu Rev Neurosci* 7:413–442
248. Resko JA, Roselli CE 1997 Prenatal hormones organize sex differences of the neuroendocrine reproductive system: observations on guinea pigs and nonhuman primates. *Cell Mol Neurobiol* 17:627–648
249. McEwen BS 1999 Permanence of brain sex differences and structural plasticity of the adult brain. *Proc Natl Acad Sci USA* 96:7128–7130
250. Toga AW, Thompson PM 2003 Mapping brain asymmetry. *Nat Rev Neurosci* 4:37–48
251. Swaab DF, Chung WC, Kruijver FP, Hofman MA, Hestiantoro A 2003 Sex differences in the hypothalamus in the different stages of human life. *Neurobiol Aging* 24(Suppl 1):S1–S16
252. Cohen-Bendahan CC, Buitelaar JK, Van Goozen SH, Cohen-Kettenis PT 2004 Prenatal exposure to testosterone and functional cerebral lateralization: a study in same-sex and opposite-sex twin girls. *Psychoneuroendocrinology* 29:911–916
253. Federman DD 2006 The biology of human sex differences. *N Engl J Med* 354:1507–1514
254. Coffey CE, Lucke JF, Saxton JA, Ratcliff G, Uritas LJ, Billig B, Bryan RN 1998 Sex differences in brain aging: a quantitative magnetic resonance imaging study. *Arch Neurol* 55:169–179
255. Pilgrim C, Hutchison JB 1994 Developmental regulation of sex differences in the brain: can the role of gonadal steroids be redefined? *Neuroscience* 60:843–855
256. Sibug R, Kuppers E, Beyer C, Maxson SC, Pilgrim C, Reisert I 1996 Genotype-dependent sex differentiation of dopaminergic neurons in primary cultures of embryonic mouse brain. *Dev Brain Res* 93:136–142
257. Arnold AP 2004 Sex chromosomes and brain gender. *Nat Rev Neurosci* 5:701–708
258. Arnold AP, Burgoyne PS 2004 Are XX and XY brain cells intrinsically different? *Trends Endocrinol Metab* 15:6–11
259. Magnaghi V, Veiga S, Ballabio M, Gonzalez LC, Garcia-Segura LM, Melcangi RC 2006 Sex-dimorphic effects of progesterone and its reduced metabolites on gene expression of myelin proteins by rat Schwann cells. *J Peripher Nerv Syst* 11:111–118
260. Stein DG, Hoffman SW 2003 Estrogen and progesterone as neuroprotective agents in the treatment of acute brain injuries. *Pediatr Rehabil* 6:13–22
261. Emerson CS, Headrick JP, Vink R 1993 Estrogen improves biochemical and neurologic outcome following traumatic brain injury in male rats, but not in females. *Brain Res* 608:95–100
262. Rusa R, Alkayed NJ, Crain BJ, Traystman RJ, Kimes AS, London ED, Klaus JA, Hurn PD 1999 17 β -Estradiol reduces stroke injury in estrogen-deficient female animals. *Stroke* 30:1665–1670
263. Miranda P, Williams CL, Einstein G 1999 Granule cells in aging rats are sexually dimorphic in their response to estradiol. *J Neurosci* 19:3316–3325
264. Groswasser Z, Cohen M, Keren O 1998 Female TBI patients recover better than males. *Brain Inj* 12:805–808
265. Bayir H, Marion DW, Puccio AM, Wisniewski SR, Janesko KL, Clark RS, Kochanek PM 2004 Marked gender effect on lipid peroxidation after severe traumatic brain injury in adult patients. *J Neurotrauma* 21:1–8
266. Wagner AK, Bayir H, Ren D, Puccio AM, Zafonte RD, Kochanek PM 2004 Relationship between cerebrospinal fluid markers of excitotoxicity, ischemia, and oxidative damage after severe TBI: the impact of gender, age and hypothermia. *J Neurotrauma* 21:125–136
267. Farace E, Alves WM 2000 Do women fare worse: a metaanalysis of gender differences in traumatic brain injury outcome. *J Neurosurg* 93:539–545
268. Coimbra R, Hoyt DB, Potenza BM, Fortlage D, Hollingsworth-Fridlund P 2003 Does sexual dimorphism influence outcome of traumatic brain injury patients? The answer is no! *J Trauma* 54:689–700
269. Voskuhl RR 2002 Gender issues and multiple sclerosis. *Curr Neurol Neurosci Rep* 2:277–286
270. el-Etr M, Vukusic S, Gignoux L, Durand-Dubief F, Achiti I, Baulieu EE, Confavreux C 2005 Steroid hormones in multiple sclerosis. *J Neurol Sci* 233:49–54
271. Tomassini V, Pozzilli C 2006 Sex hormones: a role in the control of multiple sclerosis? *Expert Opin Pharmacother* 7:857–868
272. Garidou L, Laffont S, Douin-Echinard V, Coureau C, Krust A, Chambon P, Guery JC 2004 Estrogen receptor α signaling in inflammatory leukocytes is dispensable for 17 β -estradiol-mediated inhibition of experimental autoimmune encephalomyelitis. *J Immunol* 173:2435–2442
273. Matejuk A, Hopke C, Vandenbark AA, Hurn PD, Offner H 2005 Middle-age male mice have increased severity of experimental autoimmune encephalomyelitis and are unresponsive to testosterone therapy. *J Immunol* 174:2387–2395
274. Cerghet M, Skoff RP, Bessert D, Zhang Z, Mullins C, Ghandour MS 2006 Proliferation and death of oligodendrocytes and myelin proteins are differentially regulated in male and female rodents. *J Neurosci* 26:1439–1447
275. Confavreux C, Vukusic S 2006 Natural history of multiple sclerosis: a unifying concept. *Brain* 129:606–616
276. Azoicita I, Leonelli E, Magnaghi V, Veiga S, Garcia S, Melcangi RC 2003 Progesterone and its derivatives dihydroprogesterone and tetrahydroprogesterone reduce myelin fiber morphological abnormalities and myelin fiber loss in the sciatic nerve of aged rats. *Neurobiol Aging* 24:853–860
277. Ibanez C, Shields SA, El Etr M, Leonelli E, Magnaghi V, Li WW, Sim FJ, Baulieu EE, Melcangi RC, Schumacher M, Franklin RJ 2003 Steroids and the reversal of age-associated changes in myelination and remyelination. *Prog Neurobiol* 71:49–56
278. Alkayed NJ, Murphy SJ, Traystman RJ, Hurn PD, Miller VM 2000 Neuroprotective effects of female gonadal steroids in reproductively senescent female rats. *Stroke* 31:161–168
279. Akwa Y, Purdy RH, Koob GF, Britton KT 1999 The amygdala mediates the anxiolytic-like effect of the neurosteroid allopregnanolone in rat. *Behav Brain Res* 106:119–125
280. Brot MD, Akwa Y, Purdy RH, Koob GF, Britton KT 1997 The anxiolytic-like effects of the neurosteroid allopregnanolone: interactions with GABA(A) receptors. *Eur J Pharmacol* 325:1–7
281. Walf AA, Sumida K, Frye CA 2006 Inhibiting 5 α -reductase in the amygdala attenuates antianxiety and antidepressive behavior of naturally receptive and hormone-primed ovariectomized rats. *Psychopharmacology (Berl)* 186:302–311
282. Reddy DS, O'Malley BW, Rogawski MA 2005 Anxiolytic activity of progesterone in progesterone receptor knockout mice. *Neuropharmacology* 48:14–24
283. Frye CA, Walf AA, Rhodes ME, Harney JP 2004 Progesterone enhances motor, anxiolytic, analgesic, and antidepressive behavior of wild-type mice, but not those deficient in type 1 5 α -reductase. *Brain Res* 1004:116–124

284. Frye CA, Sumida K, Dudek BC, Harney JP, Lydon JP, O'Malley BW, Pfaff DW, Rhodes ME 2006 Progesterone's effects to reduce anxiety behavior of aged mice do not require actions via intracellular progesterin receptors. *Psychopharmacology (Berl)* 186:312–322
285. Frye CA, Sumida K, Lydon JP, O'Malley BW, Pfaff DW 2006 Mid-aged and aged wild-type and progesterin receptor knockout (PRKO) mice demonstrate rapid progesterone and $3\alpha,5\alpha$ -THP-facilitated lordosis. *Psychopharmacology (Berl)* 185:423–432
286. Maurice T, Phan VL, Urani A, Kamei H, Noda Y, Nabeshima T 1999 Neuroactive neurosteroids as endogenous effectors for the sigma1 (σ_1) receptor: pharmacological evidence and therapeutic opportunities. *Jpn J Pharmacol* 81:125–155
287. Mayo W, Dellu F, Robel P, Cherkaoui J, Le Moal M, Baulieu EE, Simon H 1993 Infusion of neurosteroids into the nucleus basalis magnocellularis affects cognitive processes in the rat. *Brain Res* 607:324–328
288. Mayo W, George O, Darbra S, Bouyer JJ, Vallee M, Darnaudery M, Pallares M, Lemaire-Mayo V, Le MM, Piazza PV, Abrous N 2003 Individual differences in cognitive aging: implication of pregnenolone sulfate. *Prog Neurobiol* 71:43–48
289. Ladurelle N, Eychenne B, Denton D, Blair-West J, Schumacher M, Robel P, Baulieu E 2000 Prolonged intracerebroventricular infusion of neurosteroids affects cognitive performances in the mouse. *Brain Res* 858:371–379
290. Johansson IM, Birzniece V, Lindblad C, Olsson T, Bäckström T 2002 Allopregnanolone inhibits learning in the Morris water maze. *Brain Res* 934:125–131
291. Birzniece V, Bäckström T, Johansson IM, Lindblad C, Lundgren P, Lofgren M, Olsson T, Ragagnin G, Taube M, Turkmen S, Wahlström G, Wang MD, Wihlback AC, Zhu D 2006 Neuroactive steroid effects on cognitive functions with a focus on the serotonin and GABA systems. *Brain Res Rev* 51:212–239
292. Akwa Y, Purdy R, Koob GF, Britton KT 1999 The amygdala mediates the anxiolytic-like effect of the neurosteroid allopregnanolone in rat. *Behav Brain Res* 106:119–125
293. Barbaccia ML, Serra M, Purdy RH, Biggio G 2001 Stress and neuroactive steroids. *Int Rev Neurobiol* 46:243–272
294. Djebaili M, Hoffman SW, Stein DG 2004 Allopregnanolone and progesterone decrease cell death and cognitive deficits after contusion of the rat pre-frontal cortex. *Neuroscience* 123:349–359
295. Gibbs RB 2000 Long-term treatment with estrogen and progesterone enhances acquisition of a spatial memory task by ovariectomized aged rats. *Neurobiol Aging* 21:107–116
296. Bimonte-Nelson HA, Singleton RS, Williams BJ, Granholm AC 2004 Ovarian hormones and cognition in the aged female rat. II. Progesterone supplementation reverses the cognitive enhancing effects of ovariectomy. *Behav Neurosci* 118:707–714
297. Bimonte-Nelson HA, Nelson ME, Granholm AC 2004 Progesterone counteracts estrogen-induced increases in neurotrophins in the aged female rat brain. *Neuroreport* 15:2659–2663
298. Dubal DB, Wise PM 2001 Neuroprotective effects of estradiol in middle-aged female rats. *Endocrinology* 142:43–48
299. Markowska AL, Savonenko AV 2002 Effectiveness of estrogen replacement in restoration of cognitive function after long-term estrogen withdrawal in aging rats. *J Neurosci* 22:10985–10995
300. Singer CA, McMillan PJ, Dobie DJ, Dorsa DM 1998 Effects of estrogen replacement on choline acetyltransferase and trkA mRNA expression in the basal forebrain of aged rats. *Brain Res* 789:343–346
301. Tinkler GP, Voytko ML 2005 Estrogen modulates cognitive and cholinergic processes in surgically menopausal monkeys. *Prog Neuropsychopharmacol Biol Psychiatry* 29:423–431
302. Rapp PR, Morrison JH, Roberts JA 2003 Cyclic estrogen replacement improves cognitive function in aged ovariectomized rhesus monkeys. *J Neurosci* 23:5708–5714
303. Lacreuse A, Wilson ME, Herndon JG 2002 Estradiol, but not raloxifene, improves aspects of spatial working memory in aged ovariectomized rhesus monkeys. *Neurobiol Aging* 23:589–600
304. Lacreuse A 2006 Effects of ovarian hormones on cognitive function in nonhuman primates. *Neuroscience* 138:859–867
305. Murphy SJ, Littleton-Kearney MT, Hum PD 2002 Progesterone administration during reperfusion, but not preischemia alone, reduces injury in ovariectomized rats. *J Cereb Blood Flow Metab* 22:1181–1188
306. Marrone BL, Karavolas HJ 1982 Progesterone metabolism by the hypothalamus, pituitary, and uterus of the aged rat. *Endocrinology* 111:162–167
307. Wise PM, Smith MJ, Dubal DB, Wilson ME, Krajnak KM, Rosewell KL 1999 Neuroendocrine influences and repercussions of the menopause. *Endocr Rev* 20:243–248
308. Wise PM 2001 The 'menopause' and the aging brain: causes and repercussions of hypoestrogenicity. *Biogerontology* 2:113–115
309. Brown TJ, MacLusky NJ, Shanabrough M, Naftolin F 1990 Comparison of age- and sex-related changes in cell nuclear estrogen-binding capacity and progesterin receptor induction in the rat brain. *Endocrinology* 126:2965–2972
310. Funabashi T, Kleopoulos SP, Brooks PJ, Kimura F, Pfaff DW, Shinohara K, Mobbs CV 2000 Changes in estrogenic regulation of estrogen receptor α mRNA and progesterone receptor mRNA in the female rat hypothalamus during aging: an in situ hybridization study. *Neurosci Res* 38:85–92
311. Wilson ME, Rosewell KL, Kashon ML, Shughrue PJ, Merchenthaler I, Wise PM 2002 Age differentially influences estrogen receptor- α (ER α) and estrogen receptor- β (ER β) gene expression in specific regions of the rat brain. *Mech Ageing Dev* 123:593–601
312. Chakraborty TR, Ng L, Gore AC 2003 Age-related changes in estrogen receptor β in rat hypothalamus: a quantitative analysis. *Endocrinology* 144:4164–4171
313. Chakraborty TR, Gore AC 2004 Aging-related changes in ovarian hormones, their receptors, and neuroendocrine function. *Exp Biol Med (Maywood)* 229:977–987
314. Chakraborty TR, Hof PR, Ng L, Gore AC 2003 Stereologic analysis of estrogen receptor α (ER α) expression in rat hypothalamus and its regulation by aging and estrogen. *J Comp Neurol* 466:409–421
315. MacLusky NJ, McEwen BS 1978 Oestrogen modulates progesterin receptor concentrations in some rat brain regions but not in others. *Nature* 274:276–277
316. Kudwa AE, Rissman EF 2003 Double oestrogen receptor α and β knockout mice reveal differences in neural oestrogen-mediated progesterin receptor induction and female sexual behaviour. *J Neuroendocrinol* 15:978–983
317. Wise PM, McEwen BS, Parsons B, Rainbow TC 1984 Age-related changes in cytoplasmic estradiol receptor concentrations in microdissected brain nuclei: correlations with changes in steroid-induced sexual behavior. *Brain Res* 321:119–126
318. McKenna NJ, Lanz RB, O'Malley BW 1999 Nuclear receptor coregulators: cellular and molecular biology. *Endocr Rev* 20:321–344
319. Rowan BG, O'Malley BW 2000 Progesterone receptor coactivators. *Steroids* 65:545–549
320. Matsumoto A 2002 Age-related changes in nuclear receptor coactivator immunoreactivity in motoneurons of the spinal nucleus of the bulbocavernosus of male rats. *Brain Res* 943:202–205
321. Jezierski MK, Sohrabji F 2001 Neurotrophin expression in the reproductively senescent forebrain is refractory to estrogen stimulation. *Neurobiol Aging* 22:311–321
322. Gerhold LM, Rosewell KL, Wise PM 2005 Suppression of vasoactive intestinal polypeptide in the supraoptic nucleus leads to aging-like alterations in cAMP rhythms and activation of gonadotropin-releasing hormone neurons. *J Neurosci* 25:62–67
323. Cashion AB, Smith MJ, Wise PM 2003 The morphometry of astrocytes in the rostral preoptic area exhibits a diurnal rhythm on proestrus: relationship to the luteinizing hormone surge and effects of age. *Endocrinology* 144:274–280
324. Oliet SH, Piet R, Poulain DA, Theodosis DT 2004 Glial modulation of synaptic transmission: insights from the supraoptic nucleus of the hypothalamus. *Glia* 47:258–267
325. Gibbs RB 2003 Effects of ageing and long-term hormone replacement on cholinergic neurones in the medial septum and nucleus basalis magnocellularis of ovariectomized rats. *J Neuroendocrinol* 15:477–485
326. Savonenko AV, Markowska AL 2003 The cognitive effects of ovariectomy and estrogen replacement are modulated by aging. *Neuroscience* 119:821–830
327. Daniel JM, Hulst JL, Berbling JL 2006 Estradiol replacement enhances working memory in middle-aged rats when initiated immediately after ovariectomy but not after a long-term period of ovarian hormone deprivation. *Endocrinology* 147:607–614

328. Gould E, Woolley CS, Frankfurt M, McEwen BS 1990 Gonadal steroids regulate dendritic spine density in hippocampal pyramidal cells in adulthood. *J Neurosci* 10:1286–1291
329. Woolley CS, McEwen BS 1992 Estradiol mediates fluctuation in hippocampal synapse density during the estrous cycle in the adult rat. *J Neurosci* 12:2549–2554
330. Adams MM, Fink SE, Shah RA, Janssen WG, Hayashi S, Milner TA, McEwen BS, Morrison JH 2002 Estrogen and aging affect the subcellular distribution of estrogen receptor- α in the hippocampus of female rats. *J Neurosci* 22:3608–3614
331. Adams MM, Shah RA, Janssen WG, Morrison JH 2001 Different modes of hippocampal plasticity in response to estrogen in young and aged female rats. *Proc Natl Acad Sci USA* 98:8071–8076
332. Adams MM, Fink SE, Janssen WG, Shah RA, Morrison JH 2004 Estrogen modulates synaptic N-methyl-D-aspartate receptor subunit distribution in the aged hippocampus. *J Comp Neurol* 474:419–426
333. Adams MM, Morrison JH 2003 Estrogen and the aging hippocampal synapse. *Cereb Cortex* 13:1271–1275
334. Hao J, Janssen WG, Tang Y, Roberts JA, McKay H, Lasley B, Allen PB, Greengard P, Rapp PR, Kordower JH, Hof PR, Morrison JH 2003 Estrogen increases the number of spinophilin-immunoreactive spines in the hippocampus of young and aged female rhesus monkeys. *J Comp Neurol* 465:540–550
335. Hao J, Rapp PR, Leffler AE, Leffler SR, Janssen WG, Lou W, McKay H, Roberts JA, Wearne SL, Hof PR, Morrison JH 2006 Estrogen alters spine number and morphology in prefrontal cortex of aged female rhesus monkeys. *J Neurosci* 26:2571–2578
336. Finch CE, Morgan T, Rozovsky I 2005 Estrogens, aging and neurodegenerative diseases. In: Kordon C, Gaillard RC, Christen Y, eds. *Hormones and the brain*. Berlin: Springer; 213–225
337. Stone DJ, Rozovsky I, Morgan TE, Anderson CP, Lopez LM, Shick J, Finch CE 2000 Effects of age on gene expression during estrogen-induced synaptic sprouting in the female rat. *Exp Neurol* 165:46–57
338. Holzenberger M, Kappeler L, De Magalhaes FC 2004 IGF-1 signaling and aging. *Exp Gerontol* 39:1761–1764
339. Leroi AM, Bartke A, De Benedictis G, Franceschi C, Gartner A, Gonos E, Feder ME, Kivisild T, Lee S, Kartal-Ozer N, Schumacher M, Sikora E, Slagboom E, Tatar M, Yashin AI, Vijg J, Zwaan B 2005 What evidence is there for the existence of individual genes with antagonistic pleiotropic effects? *Mech Ageing Dev* 126:421–429
340. Nordell VL, Scarborough MM, Buchanan AK, Sohrabji F 2003 Differential effects of estrogen in the injured forebrain of young adult and reproductive senescent animals. *Neurobiol Aging* 24:733–743
341. Johnson AB, Sohrabji F 2005 Estrogens anti-inflammatory effects on central and circulating immune cells vary with reproductive age. *Neurobiol Aging* 26:1365–1374
342. Bake S, Sohrabji F 2004 17β -Estradiol differentially regulates blood-brain barrier permeability in young and aging female rats. *Endocrinology* 145:5471–5475
343. Arnal JF, Gourdy P, Elhage R, Garmy-Susini B, Delmas E, Brouchet L, Castano C, Barreira Y, Couloumiers JC, Prats H, Prats AC, Bayard F 2004 Estrogens and atherosclerosis. *Eur J Endocrinol* 150:113–117
344. Mikkola TS, Clarkson TB 2002 Estrogen replacement therapy, atherosclerosis, and vascular function. *Cardiovasc Res* 53:605–619
345. Siegfried T 2007 American Astronomical Society meeting. Snapshots from the meeting. *Science* 315:455
346. Weiss G, Skurnick JH, Goldsmith LT, Santoro NF, Park SJ 2004 Menopause and hypothalamic-pituitary sensitivity to estrogen. *JAMA* 292:2991–2996
347. Reame NE, Kelche RP, Beitins IZ, Yu MY, Zawacki CM, Padmanabhan V 1996 Age effects of follicle-stimulating hormone and pulsatile luteinizing hormone secretion across the menstrual cycle of premenopausal women. *J Clin Endocrinol Metab* 81:1512–1518
348. Santoro N, Brown JR, Adel T, Skurnick JH 1996 Characterization of reproductive hormonal dynamics in the perimenopause. *J Clin Endocrinol Metab* 81:1495–1501
349. Matt DW, Kauma SW, Pincus SM, Veldhuis JD, Evans WS 1998 Characteristics of luteinizing hormone secretion in younger versus older premenopausal women. *Am J Obstet Gynecol* 178:504–510
350. Sherwin BB 2005 Surgical menopause, estrogen, and cognitive function in women: what do the findings tell us? *Ann NY Acad Sci* 1052:3–10
351. Henderson VW 2006 Estrogen-containing hormone therapy and Alzheimer's disease risk: understanding discrepant inferences from observational and experimental research. *Neuroscience* 138:1031–1039
352. Sherwin BB 2003 Estrogen and cognitive functioning in women. *Endocr Rev* 24:133–151
353. Sherwin BB 2006 Estrogen and cognitive aging in women. *Neuroscience* 138:1021–1026
354. Zandi PP, Carlson MC, Plassman BL, Welsh-Bohmer KA, Mayer LS, Steffens DC, Breitner JC 2002 Hormone replacement therapy and incidence of Alzheimer disease in older women: the Cache County Study. *JAMA* 288:2123–2129
355. Brinton RD 2004 Impact of estrogen therapy on Alzheimer's disease: a fork in the road? *CNS Drugs* 18:405–422
356. Dunkin J, Rasgon N, Wagner-Steh K, David S, Altshuler L, Rapkin A 2005 Reproductive events modify the effects of estrogen replacement therapy on cognition in healthy postmenopausal women. *Psychoneuroendocrinology* 30:284–296
357. Bagger YZ, Tanko LB, Alexandersen P, Qin G, Christiansen C 2005 Early postmenopausal hormone therapy may prevent cognitive impairment later in life. *Menopause* 12:12–17
358. Maas AH, van der Graaf Y, van der Schouw YT, Grobbee DE 2004 HRT and heart disease: problems and prospects. *Maturitas* 47:255–258
359. Maas AH, van der Schouw YT, Grobbee DE, van der Graaf Y 2004 Rise and fall of hormone therapy in postmenopausal women with cardiovascular disease. *Menopause* 11:228–235
360. Koh KK, Sakuma I 2004 Should progestins be blamed for the failure of hormone replacement therapy to reduce cardiovascular events in randomized controlled trials? *Arterioscler Thromb Vasc Biol* 24:1171–1179
361. Barrett-Connor E, Laughlin GA 2005 Hormone therapy and coronary artery calcification in asymptomatic postmenopausal women: the Rancho Bernardo Study. *Menopause* 12:40–48
362. Grodstein F, Manson JE, Stampfer MJ 2006 Hormone therapy and coronary heart disease: the role of time since menopause and age at hormone initiation. *J Womens Health* 15:35–44
363. Alexandersen P, Tanko LB, Bagger YZ, Qin G, Christiansen C 2006 The long-term impact of 2–3 years of hormone replacement therapy on cardiovascular mortality and atherosclerosis in healthy women. *Climacteric* 9:108–118
364. Harman SM, Brinton EA, Clarkson T, Heward CB, Hecht HS, Karas RH, Judelson DR, Naftolin F 2004 Is the WHI relevant to HRT started in the perimenopause? *Endocrine* 24:195–202
365. Harman SM, Naftolin F, Brinton EA, Judelson DR 2005 Is the estrogen controversy over? Deconstructing the Women's Health Initiative study: a critical evaluation of the evidence. *Ann NY Acad Sci* 1052:43–56
366. Maki PM 2006 Hormone therapy and cognitive function: is there a critical period for benefit? *Neuroscience* 138:1027–1030
367. Erickson KI, Colcombe SJ, Raz N, Korol DL, Scalf P, Webb A, Cohen NJ, McAuley E, Kramer AF 2005 Selective sparing of brain tissue in postmenopausal women receiving hormone replacement therapy. *Neurobiol Aging* 26:1205–1213
368. Eberling JL, Wu C, Haan MN, Mungas D, Buonocore M, Jagust WJ 2003 Preliminary evidence that estrogen protects against age-related hippocampal atrophy. *Neurobiol Aging* 24:725–732
369. Ghidoni R, Boccardi M, Benussi L, Testa C, Villa A, Pievani M, Gigola L, Sabatoli F, Barbiero L, Frisoni GB, Binetti G 2006 Effects of estrogens on cognition and brain morphology: involvement of the cerebellum. *Maturitas* 54:222–228
370. Low LF, Anstey KJ, Maller J, Kumar R, Wen W, Lux O, Salonikas C, Naidoo D, Sachdev P 2006 Hormone replacement therapy, brain volumes and white matter in postmenopausal women aged 60–64 years. *Neuroreport* 17:101–104
371. Maki PM, Resnick SM 2000 Longitudinal effects of estrogen replacement therapy on PET cerebral blood flow and cognition. *Neurobiol Aging* 21:373–383

372. **Rasgon NL, Silverman D, Siddarth P, Miller K, Ercoli LM, Elman S, Lavretsky H, Huang SC, Phelps ME, Small GW** 2005 Estrogen use and brain metabolic change in postmenopausal women. *Neurobiol Aging* 26:229–235
373. **Harman SM, Brinton EA, Cedars M, Lobo R, Manson JE, Merriam GR, Miller VM, Naftolin F, Santoro N** 2005 KEEPS: the Kronos Early Estrogen Prevention Study. *Climacteric* 8:3–12
374. **Menon DV, Vongpatanasin W** 2006 Effects of transdermal estrogen replacement therapy on cardiovascular risk factors. *Treat Endocrinol* 5:37–51
375. **Butenandt A, Westphal U** 1934 Zur Isolierung and Charakterisierung des corpus-luteum-hormons. *Ber Dtsch Chem Ges* 67:1440–1442
376. **Slotta K, Rusching H, Fels E** 1934 Reindarstellung der hormone aus dem corpus luteum. *Ber Dtsch Chem Ges* 67:1270–1273
377. **Hartman M, Wettstein A** 1934 Ein kristallisiertes hormon aus dem corpus luteum. *Helv Chim Acta* 17:878–882
378. **Wintersteiner O, Allen WM** 1934 Crystalline progesterin. *J Biol Chem* 107:321–336
379. **Allen WM** 2005 My life with progesterone. 1970. *Am J Obstet Gynecol* 193:1575–1577
380. **Thomas T, Rhodin J, Clark L, Garces A** 2003 Progestins initiate adverse events of menopausal estrogen therapy. *Climacteric* 6:293–301
381. **Adams MR, Kaplan JR, Manuck SB, Koritnik DR, Parks JS, Wolfe MS, Clarkson TB** 1990 Inhibition of coronary artery atherosclerosis by 17- β estradiol in ovariectomized monkeys. Lack of an effect of added progesterone. *Arteriosclerosis* 10:1051–1057
382. **Wagner JD, Clarkson TB, St Clair RW, Schwenke DC, Shively CA, Adams MR** 1991 Estrogen and progesterone replacement therapy reduces low density lipoprotein accumulation in the coronary arteries of surgically postmenopausal cynomolgus monkeys. *J Clin Invest* 88:1995–2002
383. **Williams JK, Cline JM, Honore EK, Delansorne R, Paris J** 1998 Coadministration of norethisterone acetate does not diminish the beneficial effects of estradiol on coronary artery dilator responses in nonhuman primates (*Macaca fascicularis*). *Am J Obstet Gynecol* 179:1288–1294
384. **Perusquia M, Villalon CM, Navarrete E, Garcia GA, Perez-Palacios G, Lemus AE** 2003 Vasodilating effect of norethisterone and its 5 α metabolites: a novel nongenomic action. *Eur J Pharmacol* 475:161–169
385. **Hanke H, Hanke S, Finking G, Muhic-Lohrer A, Muck AO, Schmahl FW, Haasis R, Hombach V** 1996 Different effects of estrogen and progesterone on experimental atherosclerosis in female versus male rabbits. Quantification of cellular proliferation by bromodeoxyuridine. *Circulation* 94:175–181
386. **Hanke H, Hanke S, Bruck B, Brehme U, Gugel N, Finking G, Muck AO, Schmahl FW, Hombach V, Haasis R** 1996 Inhibition of the protective effect of estrogen by progesterone in experimental atherosclerosis. *Atherosclerosis* 121:129–138
387. **Sunday L, Tran MM, Krause DN, Duckles SP** 2006 Estrogen and progestagens differentially modulate vascular proinflammatory factors. *Am J Physiol Endocrinol Metab* 291:E261–E267
388. **Medina RA, Aranda E, Verdugo C, Kato S, Owen GI** 2003 The action of ovarian hormones in cardiovascular disease. *Biol Res* 36:325–341
389. **Adams MR, Williams JK, Kaplan JR** 2004 Estrogens, progestins, and atherosclerosis. *Arterioscler Thromb Vasc Biol* 24:e190–e191
390. **Williams JK, Honore EK, Washburn SA, Clarkson TB** 1994 Effects of hormone replacement therapy on reactivity of atherosclerotic coronary arteries in cynomolgus monkeys. *J Am Coll Cardiol* 24:1757–1761
391. **Williams JK, Hall J, Anthony MS, Register TC, Reis SE, Clarkson TB** 2002 A comparison of tibolone and hormone replacement therapy on coronary artery and myocardial function in ovariectomized atherosclerotic monkeys. *Menopause* 9:41–51
392. **Adams MR, Register TC, Golden DL, Wagner JD, Williams JK** 1997 Medroxyprogesterone acetate antagonizes inhibitory effects of conjugated equine estrogens on coronary artery atherosclerosis. *Arterioscler Thromb Vasc Biol* 17:217–221
393. **Miyagawa K, Rosch J, Stanczyk F, Hermsmeyer K** 1997 Medroxyprogesterone interferes with ovarian steroid protection against coronary vasospasm. *Nat Med* 3:324–327
394. **Miller AP, Chen YF, Xing D, Feng W, Oparil S** 2003 Hormone replacement therapy and inflammation: interactions in cardiovascular disease. *Hypertension* 42:657–663
395. **Simoncini T, Caruso A, Garibaldi S, Fu XD, Giretti MS, Baldacci C, Scorticati C, Fornari L, Mannella P, Genazzani AR** 2006 Activation of nitric oxide synthesis in human endothelial cells using norethisterone acetate. *Obstet Gynecol* 108:969–978
396. **Pedersen SH, Pedersen NG, Dalsgaard T, Lund CO, Nilas L, Ottesen B** 2004 Different cerebrovascular effects of medroxyprogesterone acetate and norethisterone acetate in the New Zealand White rabbit. *Climacteric* 7:12–22
397. **Bain CA, Walters MR, Lees KR, Lumsden MA** 2004 The effect of HRT on cerebral haemodynamics and cerebral vasomotor reactivity in post-menopausal women. *Hum Reprod* 19:2411–2414
398. **Graham JD, Clarke CL** 1997 Physiological action of progesterone in target tissues. *Endocr Rev* 18:502–519
399. **Laidlaw IJ, Clarke RB, Howell A, Owen AW, Potten CS, Anderson E** 1995 The proliferation of normal human breast tissue implanted into athymic nude mice is stimulated by estrogen but not progesterone. *Endocrinology* 136:164–171
400. **Clarke RB** 2003 Steroid receptors and proliferation in the human breast. *Steroids* 68:789–794
401. **Clarke RB** 2006 Ovarian steroids and the human breast: regulation of stem cells and cell proliferation. *Maturitas* 54:327–334
402. **Lydon JP, Ge G, Kittrell FS, Medina D, O'Malley BW** 1999 Murine mammary gland carcinogenesis is critically dependent on progesterone receptor function. *Cancer Res* 59:4276–4284
403. **Conneely OM, Mulac J, DeMayo F, Lydon JP, O'Malley BW** 2002 Reproductive functions of progesterone receptors. *Recent Prog Horm Res* 57:339–355
404. **Sivaraman L, Hilsenbeck SG, Zhong L, Gay J, Conneely OM, Medina D, O'Malley BW** 2001 Early exposure of the rat mammary gland to estrogen and progesterone blocks co-localization of estrogen receptor expression and proliferation. *J Endocrinol* 171:75–83
405. **Graham JD, Clarke CL** 2002 Expression and transcriptional activity of progesterone receptor A and progesterone receptor B in mammalian cells. *Breast Cancer Res* 4:187–190
406. **Mote PA, Bartow S, Tran N, Clarke CL** 2002 Loss of co-ordinate expression of progesterone receptors A and B is an early event in breast carcinogenesis. *Breast Cancer Res Treat* 72:163–172
407. **Mulac-Jericevic B, Conneely OM** 2005 Reproductive tissue-selective actions of progesterone receptors. *Ernst Schering Res Found Workshop* 52:19–37
408. **Wiebe JP** 2006 Progesterone metabolites in breast cancer. *Endocr Relat Cancer* 13:717–738
409. **Ma Y, Katiyar P, Jones LP, Fan S, Zhang Y, Furth PA, Rosen EM** 2006 The breast cancer susceptibility gene BRCA1 regulates progesterone receptor signaling in mammary epithelial cells. *Mol Endocrinol* 20:14–34
410. **Poole AJ, Li Y, Kim Y, Lin SC, Lee WH, Lee EY** 2006 Prevention of Brca1-mediated mammary tumorigenesis in mice by a progesterone antagonist. *Science* 314:1467–1470
411. **Micheli A, Muti P, Secretò G, Krogh V, Meneghini E, Venturelli E, Sieri S, Pala V, Berrino F** 2004 Endogenous sex hormones and subsequent breast cancer in premenopausal women. *Int J Cancer* 112:312–318
412. **Kaaks R, Berrino F, Key T, Rinaldi S, Dossus L, Biessy C, Secretò G, Amiano P, Bingham S, Boeing H, Bueno de Mesquita HB, Chang-Claude J, Clavel-Chapelon F, Fournier A, van Gils CH, Gonzalez CA, Gurrea AB, Critselis E, Khaw KT, Krogh V, Lahmann PH, Nagel G, Olsen A, Onland-Moret NC, Overvad K, Palli D, Panico S, Peeters P, Quiros JR, Roddam A, Thiebaut A, Tjonneland A, Chirlaque MD, Trichopoulou A, Trichopoulos D, Tumino R, Vineis P, Norat T, Ferrari P, Slimani N, Riboli E** 2005 Serum sex steroids in premenopausal women and breast cancer risk within the European Prospective Investigation into Cancer and Nutrition (EPIC). *J Natl Cancer Inst* 97:755–765
413. **de Lignieres B** 2002 Effects of progestogens on the postmenopausal breast. *Climacteric* 5:229–235
414. **Fournier A, Berrino F, Riboli E, Avenel V, Clavel-Chapelon F** 2005

- Breast cancer risk in relation to different types of hormone replacement therapy in the E3N-EPIC cohort. *Int J Cancer* 114:448–454
415. Fournier A, Berrino F, Riboli E, Avenel V, Clavel-Chapelon F 2005 Breast cancer risk in relation to different types of hormone replacement therapy: update of the E3N cohort study results. *Climacteric* 8(Suppl 2):235
 416. Wood CE, Register TC, Lees CJ, Chen H, Kimrey S, Mark CJ 2007 Effects of estradiol with micronized progesterone or medroxyprogesterone acetate on risk markers for breast cancer in postmenopausal monkeys. *Breast Cancer Res Treat* 101:125–134
 417. Beral V, Bull D, Doll R, Key T, Peto R, Reeve G 1997 Breast cancer and hormone replacement therapy: collaborative reanalysis of data from 51 epidemiological studies of 52,705 women with breast cancer and 108,411 women without breast cancer. *Lancet* 350:1047–1059
 418. Beral V 2003 Breast cancer and hormone-replacement therapy in the Million Women Study. *Lancet* 362:419–427
 419. Chlebowski RT, Hendrix SL, Langer RD, Stefanick ML, Gass M, Lane D, Rodabough RJ, Gilligan MA, Cyr MG, Thomson CA, Khandekar J, Petrovitch H, McTiernan A 2003 Influence of estrogen plus progestin on breast cancer and mammography in healthy postmenopausal women: the Women's Health Initiative Randomized Trial. *JAMA* 289:3243–3253
 420. Chlebowski RT, Anderson GL 2005 Progestins and recurrence in breast cancer survivors. *J Natl Cancer Inst* 97:471–472
 421. von Schoultz E, Rutqvist LE 2005 Menopausal hormone therapy after breast cancer: the Stockholm randomized trial. *J Natl Cancer Inst* 97:533–535
 422. Collins JA, Blake JM, Crosignani PG 2005 Breast cancer risk with postmenopausal hormonal treatment. *Hum Reprod Update* 11:545–560
 423. Greiser CM, Greiser EM, Doren M 2005 Menopausal hormone therapy and risk of breast cancer: a meta-analysis of epidemiological studies and randomized controlled trials. *Hum Reprod Update* 11:561–573
 424. Buchanan G, Birrell SN, Peters AA, Bianco-Miotto T, Ramsay K, Cops EJ, Yang M, Harris JM, Simila HA, Moore NL, Bentel JM, Ricciardelli C, Horsfall DJ, Butler LM, Tilley WD 2005 Decreased androgen receptor levels and receptor function in breast cancer contribute to the failure of response to medroxyprogesterone acetate. *Cancer Res* 65:8487–8496
 425. Ouatas T, Halverson D, Steeg PS 2003 Dexamethasone and medroxyprogesterone acetate elevate Nm23-H1 metastasis suppressor gene expression in metastatic human breast carcinoma cells: new uses for old compounds. *Clin Cancer Res* 9:3763–3772
 426. Melton III LJ, Chrischilles EA, Cooper C, Lane AW, Riggs BL 1992 Perspective. How many women have osteoporosis? *J Bone Miner Res* 7:1005–1010
 427. Riggs BL, Khosla S, Melton LJ 2002 Sex steroids and the construction and conservation of the adult skeleton. *Endocr Rev* 23:279–302
 428. Epstein S 2006 Update of current therapeutic options for the treatment of postmenopausal osteoporosis. *Clin Ther* 28:151–173
 429. Balasch J 2003 Sex steroids and bone: current perspectives. *Hum Reprod Update* 9:207–222
 430. Prior JC 1990 Progesterone as a bone-trophic hormone. *Endocr Rev* 11:386–398
 431. O'Brien CA, Jia D, Plotkin LI, Bellido T, Powers CC, Stewart SA, Manolagas SC, Weinstein RS 2004 Glucocorticoids act directly on osteoblasts and osteocytes to induce their apoptosis and reduce bone formation and strength. *Endocrinology* 145:1835–1841
 432. Shea B, Wells G, Cranney A, Zytaruk N, Robinson V, Griffith L, Ortiz Z, Peterson J, Adachi J, Tugwell P, Guyatt G 2002 Meta-analysis of the therapies for postmenopausal osteoporosis. VII. Meta-analysis of calcium supplementation for the prevention of postmenopausal osteoporosis. *Endocr Rev* 23:552–559
 433. Papadimitropoulos E, Wells G, Shea B, Gillespie W, Weaver B, Zytaruk N, Cranney A, Adachi J, Tugwell P, Josse R, Greenwood C, Guyatt G 2002 Meta-analysis of the efficacy of vitamin D treatment in preventing osteoporosis in postmenopausal women. *Endocr Rev* 23:560–569
 434. Speroff L, Rowan J, Symons J, Genant H, Wilborn W 1996 The comparative effect on bone density, endometrium, and lipids of continuous hormones as replacement therapy (CHART study). A randomized controlled trial. *JAMA* 276:1397–1403
 435. Slootweg MC, Ederveen AG, Schot LP, Schoonen WG, Kloosterboer HJ 1992 Oestrogen and progestogen synergistically stimulate human and rat osteoblast proliferation. *J Endocrinol* 133:R5–R8
 436. Wei LL, Leach MW, Miner RS, Demers LM 1993 Evidence for progesterone receptors in human osteoblast-like cells. *Biochem Biophys Res Commun* 195:525–532
 437. Liang M, Liao EY, Xu X, Luo XH, Xiao XH 2003 Effects of progesterone and 18-methyl levonorgestrel on osteoblastic cells. *Endocr Res* 29:483–501
 438. Grosse B, Kachkache M, Le M, V, Lieberherr M 2000 Membrane signalling and progesterone in female and male osteoblasts. I. Involvement of intracellular Ca(2+), inositol trisphosphate, and diacylglycerol, but not cAMP. *J Cell Biochem* 79:334–345
 439. Le Mellay V, Lieberherr M 2000 Membrane signaling and progesterone in female and male osteoblasts. II. Direct involvement of G α q/11 coupled to PLC- β 1 and PLC- β 3. *J Cell Biochem* 79:173–181
 440. Luo XH, Liao EY, Su X 2002 Progesterone upregulates TGF- β isoforms (b1, b2, and b3) expression in normal human osteoblast-like cells. *Calcif Tissue Int* 71:329–334
 441. Bellows CG, Pei W, Jia Y, Heersche JN 2003 Proliferation, differentiation and self-renewal of osteoprogenitors in vertebral cell populations from aged and young female rats. *Mech Ageing Dev* 124:747–757
 442. Pei W, Bellows CG, Jia Y, Heersche JN 2006 Effect of age on progesterone receptor expression, and osteoprogenitor proliferation and differentiation in female rat vertebral cell populations. *J Endocrinol* 190:261–270
 443. Eriksen EF, Colvard DS, Berg NJ, Graham ML, Mann KG, Spelsberg TC, Riggs BL 1988 Evidence of estrogen receptors in normal human osteoblast-like cells. *Science* 241:84–86
 444. Viereck V, Grundker C, Blaschke S, Niederkleine B, Siggelkow H, Frosch KH, Raddatz D, Emons G, Hofbauer LC 2003 Raloxifene concurrently stimulates osteoprotegerin and inhibits interleukin-6 production by human trabecular osteoblasts. *J Clin Endocrinol Metab* 88:4206–4213
 445. Rickard DJ, Monroe DG, Ruesink TJ, Khosla S, Riggs BL, Spelsberg TC 2003 Phytoestrogen genistein acts as an estrogen agonist on human osteoblastic cells through estrogen receptors α and β . *J Cell Biochem* 89:633–646
 446. Rickard DJ, Waters KM, Ruesink TJ, Khosla S, Katzenellenbogen JA, Katzenellenbogen BS, Riggs BL, Spelsberg TC 2002 Estrogen receptor isoform-specific induction of progesterone receptors in human osteoblasts. *J Bone Miner Res* 17:580–592
 447. Ishida Y, Heersche JN 1999 Progesterone- and dexamethasone-dependent osteoprogenitors in bone cell populations derived from rat vertebrae are different and distinct. *Endocrinology* 140:3210–3218
 448. Huang QX, Zhou HD, Liao EY, Hu PA 2005 Effects of estradiol and progesterone on the expression of insulin receptor substrate in human osteoblasts. *Zhonghua Yi Xue Za Zhi* 85:743–746
 449. Akune T, Ogata N, Hoshi K, Kubota N, Terauchi Y, Tobe K, Takagi H, Azuma Y, Kadowaki T, Nakamura K, Kawaguchi H 2002 Insulin receptor substrate-2 maintains predominance of anabolic function over catabolic function of osteoblasts. *J Cell Biol* 159:147–156
 450. The Writing Group of the PEPI Trial 1996 Effects of hormone therapy on bone mineral density. Results from the postmenopausal estrogen/progesterone intervention (PEPI) trial. *JAMA* 276:1389–1396
 451. Liu JH, Muse KN 2005 The effects of progestins on bone density and bone metabolism in postmenopausal women: a randomized controlled trial. *Am J Obstet Gynecol* 192:1316–1323
 452. Lindsay R, Gallagher JC, Kleerekoper M, Pickar JH 2002 Effect of lower doses of conjugated equine estrogens with and without medroxyprogesterone acetate on bone in early postmenopausal women. *JAMA* 287:2668–2676
 453. Banks E, Beral V, Reeves G, Balkwill A, Barnes I 2004 Fracture incidence in relation to the pattern of use of hormone therapy in postmenopausal women. *JAMA* 291:2212–2220

454. Wells G, Tugwell P, Shea B, Guyatt G, Peterson J, Zytaruk N, Robinson V, Henry D, O'Connell D, Cranney A 2002 Meta-analysis of therapies for postmenopausal osteoporosis. V. Meta-analysis of the efficacy of hormone replacement therapy in treating and preventing osteoporosis in postmenopausal women. *Endocr Rev* 23:529–539
455. Horowitz M, Wishart JM, Need AG, Morris HA, Nordin BE 1993 Effects of norethisterone on bone related biochemical variables and forearm bone mineral in post-menopausal osteoporosis. *Clin Endocrinol (Oxf)* 39:649–655
456. Banks E, Berrington A, Casabonne D 2001 Overview of the relationship between use of progestogen-only contraceptives and bone mineral density. *BJOG* 108:1214–1221
457. Ishida Y, Ishida Y, Heersche JN 2002 Pharmacologic doses of medroxyprogesterone may cause bone loss through glucocorticoid activity: an hypothesis. *Osteoporos Int* 13:601–605
458. Brown JP, Josse RG 2002 Clinical practice guidelines for the diagnosis and management of osteoporosis in Canada. *CMAJ* 167: S1–S34
459. Ciana P, Vegeto E, Beato M, Chambon P, Gustafsson JA, Parker M, Wahli W, Maggi A 2002 Looking at nuclear receptors from the heights of Erice. Workshop on nuclear receptor structure and function. *EMBO Rep* 3:125–129
460. Li X, O'Malley BW 2003 Unfolding the action of progesterone receptors. *J Biol Chem* 278:39261–39264
461. Lösel R, Falkenstein E, Feuring M, Schultz A, Tillmann HC, Rossol H, Wehling M 2003 Nongenomic steroid action: controversies, questions, and answers. *Physiol Rev* 83:965–1016
462. Schumacher M, Ghomari AM, Guennoun R, Labombarda F, Gonzalez SL, Gonzalez Deniselle MC, Massaad C, Grenier J, Rajkowski KM, Robert F, Baulieu EE, De Nicola AF 2005 Progestins and antiprogestins: mechanisms of action, neuroprotection and myelination. In: Kordon C, Gaillard RC, Christen Y, eds. *Hormones and the brain*. Berlin: Springer; 111–154
463. Kastner P, Krust A, Turcotte B, Stropp U, Tora L, Gronemeyer H, Chambon P 1990 Two distinct estrogen-regulated promoters generate transcripts encoding the two functionally different human progesterone receptor forms A and B. *EMBO J* 9:1603–1614
464. Conneely OM, Lydon JP 2000 Progesterone receptors in reproduction: functional impact of the A and B isoforms. *Steroids* 65: 571–577
465. Takimoto GS, Tung L, Abdel H, Abel MG, Sartorius CA, Richer JK, Jacobsen BM, Bain DL, Horwitz KB 2003 Functional properties of the N-terminal region of progesterone receptors and their mechanistic relationship to structure. *J Steroid Biochem Mol Biol* 85: 209–219
466. Guiochon-Mantel A, Milgrom E 1993 Cytoplasmic-nuclear trafficking of steroid hormone receptors. *Trends Endocrinol Metab* 4:322–328
467. Beato M, Klug J 2000 Steroid hormone receptors: an update. *Hum Reprod Update* 6:225–236
468. Meyer ME, Quirin-Stricker C, Lerouge T, Bocquel MT, Gronemeyer H 1992 A limiting factor mediates the differential activation of promoters by the human progesterone receptor isoforms. *J Biol Chem* 267:10882–10887
469. Vegeto E, Shahbaz MM, Wen DX, Goldman ME, O'Malley BW, McDonnell DP 1993 Human progesterone receptor A form is a cell- and promoter-specific repressor of human progesterone receptor B function. *Mol Endocrinol* 7:1244–1255
470. Horwitz KB, Sartorius CA, Hovland AR, Jackson TA, Groshong SD, Tung L, Takimoto GS 1995 Surprises with antiprogestins: novel mechanisms of progesterone receptor action. In: Bock GR, Goode JA, eds. *Nonreproductive actions of sex steroids* (CIBA Foundation Symposium 191). Chichester: John Wiley; 235–253
471. Dijkema R, Schoonen WG, Teuwen R, van der Struik E, de Ries RJ, van der Kar BA, Olijve W 1998 Human progesterone receptor A and B isoforms in CHO cells. I. Stable transfection of receptor and receptor-responsive reporter genes: transcription modulation by (anti)progestagens. *J Steroid Biochem Mol Biol* 64:147–156
472. Hovland AR, Powell RL, Takimoto GS, Tung L, Horwitz KB 1998 An N-terminal inhibitory function, IF, suppresses transcription by the A-isoform but not the B-isoform of human progesterone receptors. *J Biol Chem* 273:5455–5460
473. Wen DX, Xu YF, Mais DE, Goldman ME, McDonnell DP 1994 The A and B isoforms of the human progesterone receptor operate through distinct signaling pathways within target cells. *Mol Cell Biol* 14:8356–8364
474. Kraus WL, Weis KE, Katzenellenbogen BS 1995 Inhibitory cross-talk between steroid hormone receptors: differential targeting of estrogen receptor in the repression of its transcriptional activity by agonist- and antagonist-occupied progesterone receptors. *Mol Cell Biol* 15:1847–1857
475. Abdel-Hafiz H, Takimoto GS, Tung L, Horwitz KB 2002 The inhibitory function in human progesterone receptor N terminus binds SUMO-1 protein to regulate autoinhibition and transrepression. *J Biol Chem* 277:33950–33956
476. Jacobsen BM, Richer JK, Schittone SA, Horwitz KB 2002 New human breast cancer cells to study progesterone receptor isoform ratio effects and ligand-independent gene regulation. *J Biol Chem* 277:27793–27800
477. Richer JK, Jacobsen BM, Manning NG, Abel MG, Wolf DM, Horwitz KB 2002 Differential gene regulation by the two progesterone receptor isoforms in human breast cancer cells. *J Biol Chem* 277:5209–5218
478. Shyamala G, Yang X, Silberstein G, Barcellos H, Dale E 1998 Transgenic mice carrying an imbalance in the native ratio of A to B forms of progesterone receptor exhibit developmental abnormalities in mammary glands. *Proc Natl Acad Sci USA* 95:696–701
479. Shyamala G, Yang X, Cardiff RD, Dale E 2000 Impact of progesterone receptor on cell-fate decisions during mammary gland development. *Proc Natl Acad Sci USA* 97:3044–3049
480. Mulac-Jericevic B, Mullinax RA, DeMayo FJ, Lydon JP, Conneely OM 2000 Subgroup of reproductive functions of progesterone mediated by progesterone receptor-B isoform. *Science* 289:1751–1754
481. Mote PA, Balleine RL, McGowan EM, Clarke CL 1999 Colocalization of progesterone receptors A and B by dual immunofluorescent histochemistry in human endometrium during the menstrual cycle. *J Clin Endocrinol Metab* 84:2963–2971
482. Fang X, Wong S, Mitchell BF 2002 Messenger RNA for progesterone receptor isoforms in the late-gestation rat uterus. *Am J Physiol Endocrinol Metab* 283:E1167–E1172
483. Kato J, Hirata S, Nozawa A, Mouri N 1993 The ontogeny of gene expression of progesterone receptors in the female rat brain. *J Steroid Biochem Mol Biol* 47:173–182
484. Camacho-Arroyo I, Perez-Palacios G, Pasapera AM, Cerbon MA 1994 Intracellular progesterone receptors are differentially regulated by sex steroid hormones in the hypothalamus and the cerebral cortex of the rabbit. *J Steroid Biochem Mol Biol* 50:299–303
485. Szabo M, Kilen SM, Nho SJ, Schwartz NB 2000 Progesterone receptor A and B messenger ribonucleic acid levels in the anterior pituitary of rats are regulated by estrogen. *Biol Reprod* 62:95–102
486. Guerra-Araiza C, Coyoy-Salgado A, Camacho-Arroyo I 2002 Sex differences in the regulation of progesterone receptor isoforms expression in the rat brain. *Brain Res Bull* 59:105–109
487. Beyer C, Damm N, Brito V, Küppers E 2002 Developmental expression of progesterone receptor isoforms in the mouse midbrain. *Neuroreport* 13:877–880
488. Inoue T, Akahira JI, Takeyama J, Suzuki T, Darnel AD, Kaneko C, Kurokawa Y, Satomi S, Sasano H 2001 Spatial and topological distribution of progesterone receptor A and B isoforms during human development. *Mol Cell Endocrinol* 182:83–89
489. Wei LL, Hawkins P, Baker C, Norris B, Sheridan PL, Quinn PG 1996 An amino-terminal truncated progesterone receptor isoform, PRc, enhances progesterone-induced transcriptional activity. *Mol Endocrinol* 10:1379–1387
490. Richer JK, Lange CA, Wierman AM, Brooks KM, Tung L, Takimoto GS, Horwitz KB 1998 Progesterone receptor variants found in breast cells repress transcription by wild-type receptors. *Breast Cancer Res Treat* 48:231–241
491. Misao R, Sun WS, Iwagaki S, Fujimoto J, Tamaya T 1998 Identification of various exon-deleted progesterone receptor mRNAs in human endometrium and ovarian endometriosis. *Biochem Biophys Res Commun* 252:302–306
492. Misao R, Nakanishi Y, Sun WS, Iwagaki S, Fujimoto J, Tamaya T 2000 Identification of exon-deleted progesterone receptor mRNAs in human uterine endometrial cancers. *Oncology* 58:60–65

493. Hodges YK, Richer JK, Horwitz KB, Horwitz LD 1999 Variant estrogen and progesterone receptor messages in human vascular smooth muscle. *Circulation* 99:2688–2693
494. Balleine RL, Hunt SM, Clarke CL 1999 Coexpression of alternatively spliced estrogen and progesterone receptor transcripts in human breast cancer. *J Clin Endocrinol Metab* 84:1370–1377
495. Hirata S, Shoda T, Kato J, Hoshi K 2000 The novel isoform of the progesterone receptor cDNA in the human testis and detection of its mRNA in the human uterine endometrium. *Oncology* 59:39–44
496. Hirata S, Shoda T, Kato J, Hoshi K 2002 The novel exon, exon T, of the human progesterone receptor gene and the genomic organization of the gene. *J Steroid Biochem Mol Biol* 80:365–367
497. Hirata S, Shoda T, Kato J, Hoshi K 2003 Isoform/variant mRNAs for sex steroid hormone receptors in humans. *Trends Endocrinol Metab* 14:124–129
498. Ogle TF, Dai D, George P, Mahesh VB 1998 Regulation of the progesterone receptor and estrogen receptor in decidua basalis by progesterone and estradiol during pregnancy. *Biol Reprod* 58:1188–1198
499. Goldman S, Weiss A, Almalah I, Shalev E 2005 Progesterone receptor expression in human decidua and fetal membranes before and after contractions: possible mechanism for functional progesterone withdrawal. *Mol Hum Reprod* 11:269–277
500. Taylor AH, McParland PC, Taylor DJ, Bell SC 2005 The progesterone receptor in human term amniochorion and placenta is isoform C. *Endocrinology* 147:687–693
501. McKenna NJ, O'Malley BW 2002 Combinatorial control of gene expression by nuclear receptors and coregulators. *Cell* 108:465–474
502. Smith CL, O'Malley BW 2004 Coregulator function: a key to understanding tissue specificity of selective receptor modulators. *Endocr Rev* 25:45–71
503. Pearce KH, Iannone MA, Simmons CA, Gray JG 2004 Discovery of novel nuclear receptor modulating ligands: an integral role for peptide interaction profiling. *Drug Discov Today* 9:741–751
504. Privalsky ML 2004 The role of corepressors in transcriptional regulation by nuclear hormone receptors. *Annu Rev Physiol* 66:315–360
505. Kalkhoven E 2004 CBP and p300: HATs for different occasions. *Biochem Pharmacol* 68:1145–1155
506. Lonard DM, O'Malley BW 2005 Expanding functional diversity of the coactivators. *Trends Biochem Sci* 30:126–132
507. Wu RC, Smith CL, O'Malley BW 2005 Transcriptional regulation by steroid receptor coactivator phosphorylation. *Endocr Rev* 26:393–399
508. Mahajan MA, Samuels HH 2005 Nuclear hormone receptor coregulator: role in hormone action, metabolism, growth, and development. *Endocr Rev* 26:583–597
509. Onate SA, Tsai SY, Tsai MJ, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354–1357
510. Hong H, Kohli K, Garabedian MJ, Stallcup MR 1997 GRIP1, a transcriptional coactivator for the AF-2 transactivation domain of steroid, thyroid, retinoid, and vitamin D receptors. *Mol Cell Biol* 17:2735–2744
511. Gehin M, Mark M, Dennefeld C, Dierich A, Gronemeyer H, Chambon P 2002 The function of TIF2/GRIP1 in mouse reproduction is distinct from those of SRC-1 and p/CIP. *Mol Cell Biol* 22:5923–5937
512. Nishihara E, O'Malley BW, Xu J 2004 Nuclear receptor coregulators are new players in nervous system development and function. *Mol Neurobiol* 30:307–325
513. Xu J, Qiu Y, DeMayo FJ, Tsai SY, Tsai MJ, O'Malley BW 1998 Partial hormone resistance in mice with disruption of the steroid receptor coactivator-1 (SRC-1) gene. *Science* 279:1922–1925
514. Xu J, Liao L, Ning G, Yoshida-Komiya H, Deng C, O'Malley BW 2000 The steroid receptor coactivator SRC-3 (p/CIP/RAC3/AIB1/ACTR/TRAM-1) is required for normal growth, puberty, female reproductive function, and mammary gland development. *Proc Natl Acad Sci USA* 97:6379–6384
515. Han SJ, DeMayo FJ, Xu J, Tsai SY, Tsai MJ, O'Malley BW 2006 Steroid receptor coactivator (SRC)-1 and SRC-3 differentially modulate tissue-specific activation functions of the progesterone receptor. *Mol Endocrinol* 20:45–55
516. Mukherjee A, Soyal SM, Fernandez-Valdivia R, Gehin M, Chambon P, DeMayo FJ, Lydon JP, O'Malley BW 2006 Steroid receptor coactivator 2 is critical for progesterone-dependent uterine function and mammary morphogenesis in the mouse. *Mol Cell Biol* 26:6571–6583
517. Mitev YA, Wolf SS, Almeida OF, Patchev VK 2003 Developmental expression profiles and distinct regional estrogen responsiveness suggest a novel role for the steroid receptor coactivator SRC-1 as a discriminative amplifier of estrogen signalling in the rat brain. *FASEB J* 17:518–519
518. Charlier TD, Ball GF, Balthazart J 2005 Inhibition of steroid receptor coactivator-1 blocks estrogen and androgen action on male sex behavior and associated brain plasticity. *J Neurosci* 25:906–913
519. Xu J, Qiu Y, DeMayo FJ, Tsai SY, Tsai MJ, O'Malley BW 1998 Partial hormone resistance in mice with disruption of the steroid receptor coactivator-1 (SRC-1) gene. *Science* 279:1922–1925
520. Nishihara E, Yoshida-Komiya H, Chan CS, Liao L, Davis RL, O'Malley BW, Xu J 2003 SRC-1 null mice exhibit moderate motor dysfunction and delayed development of cerebellar Purkinje cells. *J Neurosci* 23:213–222
521. Auger AP, Tetel MJ, McCarthy MM 2000 Steroid receptor coactivator-1 (SRC-1) mediates the development of sex-specific brain morphology and behavior. *Proc Natl Acad Sci USA* 97:7551–7555
522. Molenda HA, Griffin AL, Auger AP, McCarthy MM, Tetel MJ 2002 Nuclear receptor coactivators modulate hormone-dependent gene expression in brain and female reproductive behavior in rats. *Endocrinology* 143:436–444
523. Auger AP, Perrot S, Auger CJ, Ekas LA, Tetel MJ, McCarthy MM 2002 Expression of the nuclear receptor coactivator, cAMP response element-binding protein, is sexually dimorphic and modulates sexual differentiation of neonatal rat brain. *Endocrinology* 143:3009–3016
524. Apostolakis EM, Ramamurphy M, Zhou D, Onate S, O'Malley BW 2002 Acute disruption of select steroid receptor coactivators prevents reproductive behavior in rats and unmasks genetic adaptation in knockout mice. *Mol Endocrinol* 16:1511–1523
525. Grenier J, Trousson A, Chauchereau A, Amazit L, Lamirand A, Leclerc P, Guiochon-Mantel A, Schumacher M, Massaad C 2004 Selective recruitment of p160 coactivators on glucocorticoid-regulated promoters in Schwann cells. *Mol Endocrinol* 18:2866–2879
526. Grenier J, Trousson A, Chauchereau A, Cartaud J, Schumacher M, Massaad C 2006 Differential recruitment of p160 coactivators by glucocorticoid receptor between Schwann cells and astrocytes. *Mol Endocrinol* 20:254–267
527. Fonte C, Grenier J, Trousson A, Chauchereau A, Lahuna O, Baulieu EE, Schumacher M, Massaad C 2005 Involvement of β -catenin and unusual behavior of CBP and p300 in glucocorticosteroid signaling in Schwann cells. *Proc Natl Acad Sci USA* 102:14260–14265
528. Gronemeyer H, Gustafsson JA, Laudet V 2004 Principles for modulation of the nuclear receptor superfamily. *Nat Rev Drug Discov* 3:950–964
529. Spitz IM 2005 Progesterone receptor antagonists and selective progesterone receptor modulators (SPRMs). *Semin Reprod Med* 23:3–7
530. Chabbert-Buffet N, Meduri G, Bouchard P, Spitz IM 2005 Selective progesterone receptor modulators and progesterone antagonists: mechanisms of action and clinical applications. *Hum Reprod Update* 11:293–307
531. Wardell SE, Edwards DP 2005 Mechanisms controlling agonist and antagonist potential of selective progesterone receptor modulators (SPRMs). *Semin Reprod Med* 23:9–21
532. Zhao L, O'Neill K, Diaz BR 2005 Selective estrogen receptor modulators (SERMs) for the brain: current status and remaining challenges for developing NeuroSERMs. *Brain Res Rev* 49:472–493
533. Edwards DP 2005 Regulation of signal transduction pathways by estrogen and progesterone. *Annu Rev Physiol* 67:335–376
534. Boonyaratanakornkit V, Scott MP, Ribon V, Sherman L, Anderson SM, Maller JL, Miller WT, Edwards DP 2001 Progesterone receptor contains a proline-rich motif that directly interacts with SH3 domains and activates c-Src family tyrosine kinases. *Mol Cell* 8:269–280
535. Edwards DP, Wardell SE, Boonyaratanakornkit V 2003 Progesterone receptor interacting coregulatory proteins and cross talk

- with cell signaling pathways. *J Steroid Biochem Mol Biol* 83:173–186
536. **Migliaccio A, Piccolo D, Castoria G, Di Domenico M, Bilancio A, Lombardi M, Gong W, Beato M, Auricchio F** 1998 Activation of the Src/p21ras/Erk pathway by progesterone receptor via cross-talk with estrogen receptor. *EMBO J* 17:2008–2018
 537. **Leonhardt SA, Boonyaratankornkit V, Edwards DP** 2003 Progesterone receptor transcription and non-transcription signaling mechanisms. *Steroids* 68:761–770
 538. **Lambert JJ, Belelli D, Peden DR, Vardy AW, Peters JA** 2003 Neurosteroid modulation of GABA_A receptors. *Prog Neurobiol* 71:67–80
 539. **Majewska MD, Harrison NLS, Barker JL, Paul SM** 1986 Steroid hormone metabolites are barbiturate-like modulators of the GABA receptor. *Science* 232:1004–1007
 540. **Belelli D, Casula A, Ling A, Lambert JJ** 2002 The influence of subunit composition on the interaction of neurosteroids with GABA_A receptors. *Neuropharmacology* 43:651–661
 541. **Rupprecht R, Holsboer F** 1999 Neuroactive steroids: mechanisms of action and neuropsychopharmacological perspectives. *Trends Neurosci* 22:410–416
 542. **Morrow AL, VanDoren MJ, Penland SN, Matthews DB** 2001 The role of GABAergic neuroactive steroids in ethanol action, tolerance and dependence. *Brain Res Rev* 37:98–109
 543. **Covey DF, Evers AS, Mennerick S, Zorumski CF, Purdy RH** 2001 Recent developments in structure-activity relationships for steroid modulators of GABA(A) receptors. *Brain Res Rev* 37:91–97
 544. **Mennerick S, He Y, Jiang X, Manion BD, Wang M, Shute A, Benz A, Evers AS, Covey DF, Zorumski CF** 2004 Selective antagonism of 5 α -reduced neurosteroid effects at GABA(A) receptors. *Mol Pharmacol* 65:1191–1197
 545. **Hosie AM, Wilkins ME, da Silva HM, Smart TG** 2006 Endogenous neurosteroids regulate GABA_A receptors through two discrete transmembrane sites. *Nature* 444:486–489
 546. **Darbandi-Tonkabon R, Hastings WR, Zeng CM, Akk G, Manion BD, Bracamontes JR, Steinbach JH, Mennerick SJ, Covey DF, Evers AS** 2003 Photoaffinity labeling with a neuroactive steroid analogue. 6-Azi-pregnanolone labels voltage-dependent anion channel-1 in rat brain. *J Biol Chem* 278:13196–13206
 547. **Darbandi-Tonkabon R, Manion BD, Hastings WR, Craigen WJ, Akk G, Bracamontes JR, He Y, Sheiko TV, Steinbach JH, Mennerick SJ, Covey DF, Evers AS** 2004 Neuroactive steroid interactions with voltage-dependent anion channels: lack of relationship to GABA(A) receptor modulation and anesthesia. *J Pharmacol Exp Ther* 308:502–511
 548. **Sayeed I, Guo Q, Hoffman SW, Stein DG** 2006 Allopregnanolone, a progesterone metabolite, is more effective than progesterone in reducing cortical infarct volume after transient middle cerebral artery occlusion. *Ann Emerg Med* 47:381–389
 549. **Ciriza I, Carrero P, Azcoitia I, Lundeen SG, Garcia-Segura LM** 2004 Selective estrogen receptor modulators protect hippocampal neurons from kainic acid excitotoxicity: differences with the effect of estradiol. *J Neurobiol* 61:209–221
 550. **Ciriza I, Carrero P, Frye CA, Garcia-Segura LM** 2006 Reduced metabolites mediate neuroprotective effects of progesterone in the adult rat hippocampus. The synthetic progestin medroxyprogesterone acetate (Provera) is not neuroprotective. *J Neurobiol* 66:916–928
 551. **Xilouri M, Papazafiri P** 2006 Anti-apoptotic effects of allopregnanolone on P19 neurons. *Eur J Neurosci* 23:43–54
 552. **Ardeshiri A, Kelley MH, Korner IP, Hurn PD, Herson PS** 2006 Mechanism of progesterone neuroprotection of rat cerebellar Purkinje cells following oxygen-glucose deprivation. *Eur J Neurosci* 24:2567–2574
 553. **Brinton RD** 1994 The neurosteroid 3 α -hydroxy-5 α -pregnan-20-one induces cytoarchitectural regression in cultured fetal hippocampal neurons. *J Neurosci* 14:2763–2774
 554. **Wang JM, Johnston PB, Ball BG, Brinton RD** 2005 The neurosteroid allopregnanolone promotes proliferation of rodent and human neural progenitor cells and regulates cell-cycle gene and protein expression. *J Neurosci* 25:4706–4718
 555. **Gago N, Akwa Y, Sananes N, Guennoun R, Baulieu EE, El-Etr M, Schumacher M** 2001 Progesterone and the oligodendroglial lineage: stage-dependent biosynthesis and metabolism. *Glia* 36:295–308
 556. **Gago N, Avellana A, Evercooren AB, Schumacher M** 2003 Control of cell survival and proliferation of postnatal PSA-NCAM(+) progenitors. *Mol Cell Neurosci* 22:162–178
 557. **Gago N, el-Etr M, Sananes N, Cadepond F, Samuel D, Avellana-Adalid V, Baron-Van Evercooren A, Schumacher M** 2004 3 α ,5 α -tetrahydroprogesterone (allopregnanolone) and GABA: autocrine/paracrine interactions in the control of neonatal PSA-NCAM+ progenitor proliferation. *J Neurosci Res* 78:770–783
 558. **Brinton RD, Wang JM** 2006 Preclinical analyses of the therapeutic potential of allopregnanolone to promote neurogenesis in vitro and in vivo in transgenic mouse model of Alzheimer's disease. *Curr Alzheimer Res* 3:11–17
 559. **Schwartz-Bloom RD, Sah R** 2001 γ -Aminobutyric acid(A) neurotransmission and cerebral ischemia. *J Neurochem* 77:353–371
 560. **Galeffi F, Sah R, Pond BB, George A, Schwartz-Bloom RD** 2004 Changes in intracellular chloride after oxygen-glucose deprivation of the adult hippocampal slice: effect of diazepam. *J Neurosci* 24:4478–4488
 561. **Green AR, Hainsworth AH, Jackson DM** 2000 GABA potentiation: a logical pharmacological approach for the treatment of acute ischaemic stroke. *Neuropharmacology* 39:1483–1494
 562. **Han SK, Abraham IM, Herbison AE** 2002 Effect of GABA on GnRH neurons switches from depolarization to hyperpolarization at puberty in the female mouse. *Endocrinology* 143:1459–1466
 563. **Ben-Ari Y** 2002 Excitatory actions of GABA during development: the nature of the nurture. *Nat Rev Neurosci* 3:728–739
 564. **McCarthy MM, Auger AP, Perrot-Sinal TS** 2002 Getting excited about GABA and sex differences in the brain. *Trends Neurosci* 25:307–312
 565. **Ben-Ari Y, Khalilov I, Represa A, Gozlan H** 2004 Interneurons set the tune of developing networks. *Trends Neurosci* 27:422–427
 566. **Wolff JR, Joo F, Dames W** 1978 Plasticity in dendrites shown by continuous GABA administration in superior cervical ganglion of adult rat. *Nature* 274:72–74
 567. **Kater SB, Mattson MP, Cohan C, Connor J** 1988 Calcium regulation of the neuronal growth cone. *Trends Neurosci* 11:315–321
 568. **Meier E, Hertz L, Schousboe A** 1991 Neurotransmitter as developmental signals. *Neurochem Int* 19:1–15
 569. **Waagepetersen HS, Sonnewald U, Schousboe A** 1999 The GABA paradox: multiple roles as metabolite, neurotransmitter, and neurodifferentiative agent. *J Neurochem* 73:1335–1342
 570. **LoTurco JJ, Owens DF, Heath MJ, Davis MB, Kriegstein AR** 1995 GABA and glutamate depolarize cortical progenitor cells and inhibit DNA synthesis. *Neuron* 15:1287–1298
 571. **Marty S, Berzaghi M, Berninger B** 1997 Neurotrophins and activity-dependent plasticity of cortical interneurons. *Trends Neurosci* 20:198–202
 572. **Behar TN, Schaffner AE, Scott CA, O'Connell C, Barker JL** 1998 Differential response of cortical plate and ventricular zone cells to GABA as a migration stimulus. *J Neurosci* 18:6378–6387
 573. **Grobin AC, Gizerian S, Lieberman JA, Morrow AL** 2006 Perinatal allopregnanolone influences prefrontal cortex structure, connectivity and behavior in adult rats. *Neuroscience* 138:809–819
 574. **Lukasiuk K, Pitkanen A** 2000 GABA(A)-mediated toxicity of hippocampal neurons in vitro. *J Neurochem* 74:2445–2454
 575. **Nunez JL, McCarthy MM** 2003 Estradiol exacerbates hippocampal damage in a model of preterm infant brain injury. *Endocrinology* 144:2350–2359
 576. **Nunez JL, Alt JJ, McCarthy MM** 2003 A new model for prenatal brain damage. I. GABA_A receptor activation induces cell death in developing rat hippocampus. *Exp Neurol* 181:258–269
 577. **Stein V, Nicoll RA** 2003 GABA generates excitation. *Neuron* 37:375–378
 578. **Gulledge AT, Stuart GJ** 2003 Excitatory actions of GABA in the cortex. *Neuron* 37:299–309
 579. **Wagner S, Castel M, Gainer H, Yarom Y** 1997 GABA in the mammalian suprachiasmatic nucleus and its role in diurnal rhythmicity. *Nature* 387:598–603
 580. **Cohen I, Navarro V, Clemenceau S, Baulac M, Miles R** 2002 On the origin of interictal activity in human temporal lobe epilepsy in vitro. *Science* 298:1418–1421

581. Fujiwara-Tsukamoto Y, Isomura Y, Nambu A, Takada M 2003 Excitatory GABA input directly drives seizure-like rhythmic synchronization in mature hippocampal CA1 pyramidal cells. *Neuroscience* 119:265–275
582. Nabekura J, Ueno T, Okabe A, Furuta A, Iwaki T, Shimizu-Okabe C, Fukuda A, Akaike N 2002 Reduction of KCC2 expression and GABA_A receptor-mediated excitation after in vivo axonal injury. *J Neurosci* 22:4412–4417
583. Griffin LD, Gong W, Verot L, Mellon SH 2004 Niemann-Pick type C disease involves disrupted neurosteroidogenesis and responds to allopregnanolone. *Nat Med* 10:704–711
584. Langmade SJ, Gale SE, Frolov A, Mohri I, Suzuki K, Mellon SH, Walkley SU, Covey DF, Schaffer JE, Ory DS 2006 Pregnane X receptor (PXR) activation: a mechanism for neuroprotection in a mouse model of Niemann-Pick C disease. *Proc Natl Acad Sci USA* 103:13807–13812
585. Léna C, Changeux JP 1993 Allosteric modulations of the nicotinic acetylcholine receptor. *Trends Neurosci* 16:181–186
586. Valera S, Ballivet M, Bertrand D 1992 Progesterone modulates a neuronal nicotinic acetylcholine receptor. *Proc Natl Acad Sci USA* 89:9949–9953
587. Buisson B, Bertrand D 1999 Steroid modulation of the nicotinic acetylcholine receptor. In: Baulieu EE, Robel P, Schumacher M, eds. *Neurosteroids. A new regulatory function in the nervous system*. Totowa, NJ: Humana Press; 207–223
588. Blanton MP, Xie Y, Dangott LJ, Cohen JB 1999 The steroid promegestone is a noncompetitive antagonist of the Torpedo nicotinic acetylcholine receptor that interacts with the lipid-protein interface. *Mol Pharmacol* 55:269–278
589. Godeau JF, Schorderet-Slatkine S, Hubert P, Baulieu EE 1978 Induction of maturation in *Xenopus laevis* oocytes by a steroid linked to a polymer. *Proc Natl Acad Sci USA* 75:2353–2357
590. Baulieu EE, Godeau JF, Schorderet M, Schorderet-Slatkine S 1978 Steroid induced meiotic division in *Xenopus laevis* oocytes: surface and calcium. *Nature* 275:593–598
591. Finidori-Lepicard J, Schorderet-Slatkine S, Hanoune J, Baulieu EE 1981 Progesterone inhibits membrane-bound adenylate cyclase in *Xenopus laevis* oocytes. *Nature* 292:255
592. Ferrell JEJ 1999 *Xenopus* oocyte maturation: new lessons from a good egg. *BioEssays* 21:833–842
593. Bayaa M, Booth RA, Sheng Y, Liu XJ 2000 The classical progesterone receptor mediates *Xenopus* oocyte maturation through a nongenomic mechanism. *Proc Natl Acad Sci USA* 97:12607–12612
594. Maller JL 2001 The elusive progesterone receptor in *Xenopus* oocytes. *Proc Natl Acad Sci USA* 98:8–10
595. Bagowski CP, Myers JW, Ferrell JE 2001 The classical progesterone receptor associates with p42 MAPK and is involved in phosphatidylinositol 3-kinase signaling in *Xenopus* oocytes. *J Biol Chem* 276:37708–37714
596. Tischkau SA, Ramirez VD 1993 A specific membrane binding protein for progesterone in rat brain: sex differences and induction by estrogen. *Proc Natl Acad Sci USA* 90:1285–1289
597. Falkenstein E, Meyer C, Eisen C, Scriba PC, Wehling M 1996 Full-length cDNA sequence of a progesterone membrane-binding protein from porcine vascular smooth muscle cells. *Biochem Biophys Res Commun* 229:86–89
598. Meyer C, Schmid R, Schmieding K, Falkenstein E, Wehling M 1998 Characterization of high affinity progesterone-binding membrane proteins by anti-peptide antiserum. *Steroids* 63:111–116
599. Lösel R, Wehling M 2003 Nongenomic actions of steroid hormones. *Nat Rev Mol Cell Biol* 4:46–56
600. Krebs CJ, Jarvis ED, Chan J, Lydon JP, Ogawa S, Pfaff DW 2000 A membrane-associated progesterone-binding protein, 25-Dx, is regulated by progesterone in brain regions involved in female reproductive behavior. *Proc Natl Acad Sci USA* 97:12816–12821
601. Labombarda F, Gonzalez SL, Deniselle MC, Vinson GP, Schumacher M, De Nicola AF, Guennoun R 2003 Effects of injury and progesterone treatment on progesterone receptor and progesterone binding protein 25-Dx expression in the rat spinal cord. *J Neurochem* 87:902–913
602. Sakamoto H, Ukena K, Takemori H, Okamoto M, Kawata M, Tsutsui K 2004 Expression and localization of 25-Dx, a membrane-associated putative progesterone-binding protein, in the developing Purkinje cell. *Neuroscience* 126:325–334
603. Meffre D, Delespierre B, Guezou M, Leclerc P, Vinson GP, Schumacher M, Stein DG, Guennoun R 2005 The membrane-associated progesterone-binding protein 25-Dx is expressed in brain regions involved in water homeostasis and is up-regulated after traumatic brain injury. *J Neurochem* 93:1314–1326
604. Lösel R, Dorn-Beineke A, Falkenstein E, Wehling M, Feuring M 2004 Porcine spermatozoa contain more than one membrane progesterone receptor. *Int J Biochem Cell Biol* 36:1532–1541
605. Falkenstein E, Heck M, Gerdes D, Grube D, Christ M, Weigel M, Buddhikot M, Meizel S, Wehling M 1999 Specific progesterone binding to a membrane protein and related nongenomic effects on Ca²⁺-fluxes in sperm. *Endocrinology* 140:5999–6002
606. Peluso JJ 2006 Multiplicity of progesterone's actions and receptors in the mammalian ovary. *Biol Reprod* 75:2–8
607. Peluso JJ, Pappalardo A, Lösel R, Wehling M 2006 Progesterone membrane receptor component 1 expression in the immature rat ovary and its role in mediating progesterone's antiapoptotic action. *Endocrinology* 147:3133–3140
608. Engmann L, Lösel R, Wehling M, Peluso JJ 2006 Progesterone regulation of human granulosa/luteal cell viability by an RU486-independent mechanism. *J Clin Endocrinol Metab* 91:4962–4968
609. Quirion R, Bowen WD, Itzhak Y, Junien JL, Musacchio JM, Rothman RB, Su TP, Tam SW, Taylor DP 1992 A proposal for the classification of σ binding sites. *Trends Pharmacol Sci* 13:85–86
610. Bastianetto S, Monnet F, Junien JL, Quirion R 1999 Steroidal modulation of σ receptor function. In: Baulieu EE, Robel P, Schumacher M, eds. *Neurosteroids. A new regulatory function in the nervous system*. Totowa, NJ: Humana Press; 191–205
611. Hanner M, Moebius FF, Flandorfer A, Knaus HG, Striessnig J, Kempner E, Glossmann H 1996 Purification, molecular cloning, and expression of the mammalian σ 1-binding site. *Proc Natl Acad Sci USA* 93:8072–8077
612. Kekuda R, Prasad PD, Fei YJ, Leibach FH, Ganapathy V 1996 Cloning and functional expression of the human type 1 σ receptor (σ R1). *Biochem Biophys Res Commun* 229:553–558
613. Prasad PD, Li HW, Fei YJ, Ganapathy ME, Fujita T, Plumley LH, Yang-Feng TL, Leibach FH, Ganapathy V 1998 Exon-intron structure, analysis of promoter region, and chromosomal localization of the human type 1 σ receptor gene. *J Neurochem* 70:443–451
614. Seth P, Fei YJ, Li HW, Huang W, Leibach FH, Ganapathy V 1998 Cloning and functional characterization of a σ receptor from rat brain. *J Neurochem* 70:922–931
615. Morin-Surun MP, Collin T, Denavit-Saubie M, Baulieu EE, Monnet FP 1999 Intracellular σ 1 receptor modulates phospholipase C and protein kinase C activities in the brainstem. *Proc Natl Acad Sci USA* 96:8196–8199
616. Hayashi T, Maurice T, Su TP 2000 Ca(2+) signaling via σ (1)-receptors: novel regulatory mechanism affecting intracellular Ca(2+) concentration. *J Pharmacol Exp Ther* 293:788–798
617. Phan VL, Miyamoto Y, Nabeshima T, Maurice T 2005 Age-related expression of σ 1 receptors and antidepressant efficacy of a selective agonist in the senescence-accelerated (SAM) mouse. *J Neurosci Res* 79:561–572
618. Lecanu L, Wenguo Y, Xu J, Greeson J, Papadopoulos V 2005 Local anesthetic procaine protects rat pheochromocytoma PC12 cells against β -amyloid-induced neurotoxicity. *Pharmacology* 74:65–78
619. Monnet FP, Maurice T 2006 The σ 1 protein as a target for the non-genomic effects of neuro(active)steroids: molecular, physiological, and behavioral aspects. *J Pharmacol Sci* 100:93–118
620. Su TP, London ED, Jaffe JH 1988 Steroid binding at σ receptors suggests a link between endocrine, nervous, and immune systems. *Science* 240:219–221
621. McCann DJ, Weissman AD, Su TP 1994 σ -1 and σ -2 sites in rat brain: comparison of regional, ontogenetic, and subcellular patterns. *Synapse* 17:182–189
622. Monnet FP, Mahé V, Robel P, Baulieu EE 1995 Neurosteroids, via σ receptors, modulate the [3H]norepinephrine release evoked by N-methyl-D-aspartate in the rat hippocampus. *Proc Natl Acad Sci USA* 92:3774–3778
623. Debonnel G, Bergeron R, Monnet FP, de Montigny C 1996 Differential effects of σ ligands on the N-methyl-D-aspartate response

- in the CA1 and CA3 regions of the dorsal hippocampus: effect of mossy fiber lesioning. *Neuroscience* 71:977–987
624. **Bergeron R, de Montigny C, Debonnel G** 1999 Pregnancy reduces brain σ receptor function. *Br J Pharmacol* 127:1769–1776
625. **Phan VL, Urani A, Romieu P, Maurice T** 2002 Strain differences in σ (1) receptor-mediated behaviours are related to neurosteroid levels. *Eur J Neurosci* 15:1523–1534
626. **Maurice T** 2004 Neurosteroids and σ 1 receptors, biochemical and behavioral relevance. *Pharmacopsychiatry* 37(Suppl 3):S171–S182
627. **Zhu Y, Rice CD, Pang Y, Pace M, Thomas P** 2003 Cloning, expression, and characterization of a membrane progesterin receptor and evidence it is an intermediary in meiotic maturation of fish oocytes. *Proc Natl Acad Sci USA* 100:2231–2236
628. **Zhu Y, Thomas P** 2003 Identification, classification, and partial characterization of genes in humans and other vertebrates homologous to a fish membrane progesterin receptor. *Proc Natl Acad Sci USA* 100:2237–2242
629. **Thomas P, Pang Y, Zhu Y, Detweiler C, Doughty K** 2004 Multiple rapid progesterin actions and progesterin membrane receptor subtypes in fish. *Steroids* 69:567–573
630. **Lyons TJ, Villa NY, Regalla LM, Kupchak BR, Vagstad A, Eide DJ** 2004 Metalloregulation of yeast membrane steroid receptor homologs. *Proc Natl Acad Sci USA* 101:5506–5511
631. **Fernandes MS, Pierron V, Michalovich D, Astle S, Thornton S, Peltoketo H, Lam EW, Gellersen B, Huhtaniemi I, Allen J, Brosens JJ** 2005 Regulated expression of putative membrane progesterin receptor homologs in human endometrium and gestational tissues. *J Endocrinol* 187:89–101
632. **Josefsberg Ben-Yehoshua L, Lewellyn AL, Thomas P, Maller JL** 2007 The role of *Xenopus* membrane progesterone receptor β in mediating the effect of progesterone on oocyte maturation. *Mol Endocrinol* 21:664–673
633. **Thomas P, Pang Y, Dong J, Groenen P, Kelder J, de Vlieg J, Zhu Y, Tubbs C** 2007 Steroid and G protein binding characteristics of the seatrout and human progesterin membrane receptor α subtypes and their evolutionary origins. *Endocrinology* 148:705–718
634. **Cai Z, Stocco C** 2005 Expression and regulation of progesterin membrane receptors in the rat corpus luteum. *Endocrinology* 146:5522–5532
635. **Karteris E, Zervou S, Pang Y, Dong J, Hillhouse EW, Randeva HS, Thomas P** 2006 Progesterone signaling in human myometrium through two novel membrane G protein-coupled receptors: potential role in functional progesterone withdrawal at term. *Mol Endocrinol* 20:1519–1534
636. **Dressing GE, Thomas P** 2007 Identification of membrane progesterin receptors in human breast cancer cell lines and biopsies and their potential involvement in breast cancer. *Steroids* 72:111–116
637. **Krietsch T, Fernandes MS, Kero J, Lösel R, Heyens M, Lam EW, Huhtaniemi I, Brosens JJ, Gellersen B** 2006 Human homologs of the putative G protein-coupled membrane progesterin receptors (mPRA α , β , γ) localize to the endoplasmic reticulum and are not activated by progesterone. *Mol Endocrinol* 20:3146–3164
638. **Meyer C, Schmid R, Scriba PC, Wehling M** 1996 Purification and partial sequencing of high-affinity progesterone-binding site(s) from porcine liver membranes. *Eur J Biochem* 239:726–731
639. **Wehling M, Schultz A, Lösel R** 2006 Nongenomic actions of estrogens: exciting opportunities for pharmacology. *Maturitas* 54:321–326
640. **Thomas P, Pang Y, Filardo EJ, Dong J** 2005 Identity of an estrogen membrane receptor coupled to a G protein in human breast cancer cells. *Endocrinology* 146:624–632
641. **Pedram A, Razandi M, Levin ER** 2006 Nature of functional estrogen receptors at the plasma membrane. *Mol Endocrinol* 20:1996–2009
642. **Toran-Allerand CD, Guan X, MacLusky NJ, Horvath TL, Diano S, Singh M, Connolly Jr ES, Nethrapalli IS, Tinnikov AA** 2002 ER-X: a novel, plasma membrane-associated, putative estrogen receptor that is regulated during development and after ischemic brain injury. *J Neurosci* 22:8391–8401
643. **Toran-Allerand CD** 2004 Minireview: a plethora of estrogen receptors in the brain: where will it end? *Endocrinology* 145:1069–1074
644. **Leonard MO, Godson C, Brady HR, Taylor CT** 2005 Potentiation of glucocorticoid activity in hypoxia through induction of the glucocorticoid receptor. *J Immunol* 174:2250–2257
645. **Burger HG, Dudley EC, Robertson DM, Dennerstein L** 2002 Hormonal changes in the menopause transition. *Recent Prog Horm Res* 57:257–275
646. **Resko JA** 1969 Endocrine control of adrenal progesterone secretion in the ovariectomized rat. *Science* 164:70–71
647. **Feder HH, Resko JA, Goy RW** 1968 Progesterone levels in the arterial plasma of pre-ovulatory and ovariectomized rats. *J Endocrinol* 41:563–569
648. **Mann DR, Barraclough CA** 1973 Changes in peripheral plasma progesterone during the rat 4-day estrous cycle: an adrenal diurnal rhythm. *Proc Soc Exp Biol Med* 142:1226–1229
649. **Feder HH, Ruf KB** 1969 Stimulation of progesterone release and estrous behavior by ACTH in ovariectomized rodents. *Endocrinology* 84:171–174
650. **Fajer AB, Holzbauer M, Newport HM** 1971 The contribution of the adrenal gland to the total amount of progesterone produced in the female rat. *J Physiol (Lond)* 214:115–126
651. **Schaeffer C, Chabli A, Aron C** 1986 Endogenous progesterone and lordosis behavior in male rats given estrogen alone. *J Steroid Biochem* 25:99–102
652. **Bartosik D, Szarowski DH, Watson DJ** 1971 Influence of functioning ovarian tissue on the secretion of progesterone by the adrenal glands of female rats. *Endocrinology* 88:1425–1427
653. **Meites J, Huang HH, Riegler GD** 1976 Relation of the hypothalamo-pituitary-gonadal system to decline of reproductive functions in aging female rats. *Curr Top Mol Endocrinol* 3:3–20
654. **Lu KH, Hopper BR, Vargo TM, Yen SS** 1979 Chronological changes in sex steroid, gonadotropin and prolactin secretions in aging female rats displaying different reproductive states. *Biol Reprod* 21:193–203
655. **Tsai HW, Lapolt PS, Olcott AP, Lu JK** 2004 Temporal changes occur in the neuroendocrine control of gonadotropin secretion in aging female rats: role of progesterone. *Biol Reprod* 71:845–852
656. **Huang HH, Marshall S, Meites J** 1976 Induction of estrous cycles in old non-cyclic rats by progesterone, ACTH, ether stress or L-dopa. *Neuroendocrinology* 20:21–34
657. **Mora OA, Sanchez-Criado JE** 2004 Involvement of the corticoadrenal hormones in the pheromonal restoration of reproductive activity in aging rats. *Life Sci* 74:3285–3290
658. **Eldar-Geva T, Margalioth EJ, Brooks B, Algur N, Zylber-Haran E, Diamant YZ** 1998 The origin of serum progesterone during the follicular phase of menotropin-stimulated cycles. *Hum Reprod* 13:9–14
659. **Gutai JP, Meyer WJ, Kowarski AA, Migeon CJ** 1977 Twenty-four hour integrated concentrations of progesterone, 17-hydroxyprogesterone and cortisol in normal male subjects. *J Clin Endocrinol Metab* 44:116–120
660. **Puder JJ, Freda PU, Golland RS, Ferin M, Wardlaw SL** 2000 Stimulatory effects of stress on gonadotropin secretion in estrogen-treated women. *J Clin Endocrinol Metab* 85:2184–2188
661. **Genazzani AR, Petraglia F, Bernardi F, Casarosa E, Salvestroni C, Tonetti A, Nappi RE, Luisi S, Palumbo M, Purdy RH, Luisi M** 1998 Circulating levels of allopregnanolone in humans: gender, age, and endocrine influences. *J Clin Endocrinol Metab* 83:2099–2103
662. **Vermeulen A** 1976 The hormonal activity of the postmenopausal ovary. *J Clin Endocrinol Metab* 42:247–253
663. **Adashi EY** 1994 The climacteric ovary as a functional gonadotropin-driven androgen-producing gland. *Fertil Steril* 62:20–27
664. **Shifren JL, Schiff I** 2000 The aging ovary. *J Womens Health Gen Based Med* 9(Suppl 1):S3–S7
665. **Laughlin GA, Barrett-Connor E** 2000 Sexual dimorphism in the influence of advanced aging on adrenal hormone levels: the Rancho Bernardo Study. *J Clin Endocrinol Metab* 85:3561–3568
666. **Couzinet B, Meduri G, Lecce MG, Young J, Brailly S, Loosfelt H, Milgrom E, Schaison G** 2001 The postmenopausal ovary is not a major androgen-producing gland. *J Clin Endocrinol Metab* 86:5060–5066
667. **Havelock JC, Rainey WE, Bradshaw KD, Carr BR** 2006 The postmenopausal ovary displays a unique pattern of steroidogenic enzyme expression. *Hum Reprod* 21:309–317

668. Zhao HF, Labrie C, Simard J, de Launoit Y, Trudel C, Martel C, Rheume E, Dupont E, Luu-The V, Pelletier G 1991 Characterization of rat 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ isomerase cDNAs and differential tissue-specific expression of the corresponding mRNAs in steroidogenic and peripheral tissues. *J Biol Chem* 266:583–593
669. Baulieu EE, Robel P, Schumacher M 1999 Neurosteroids. A new regulatory function in the nervous system. Totowa, NJ: Humana Press; 1–378
670. Gingras S, Cote S, Simard J 2000 Multiple signaling pathways mediate interleukin-4-induced 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ isomerase type 1 gene expression in human breast cancer cells. *Mol Endocrinol* 14:229–240
671. Tsutsui K, Ukena K, Usui M, Sakamoto H, Takase M 2000 Novel brain function: biosynthesis and actions of neurosteroids in neurons. *Neurosci Res* 36:261–273
672. Sakamoto H, Ukena K, Tsutsui K 2001 Effects of progesterone synthesized de novo in the developing Purkinje cell on its dendritic growth and synaptogenesis. *J Neurosci* 21:6221–6232
673. Belanger C, Luu T, Dupont P, Tchernof A 2002 Adipose tissue intracrinology: potential importance of local androgen/estrogen metabolism in the regulation of adiposity. *Horm Metab Res* 34:737–745
674. Rhee HS, Oh SH, Ko BJ, Han DM, Jeon BH, Park H, Moon HB, Kim WS 2003 Expression of 3β -hydroxysteroid dehydrogenase and P450 side chain cleavage enzyme in the human uterine endometrium. *Exp Mol Med* 35:160–166
675. Nakamura Y, Suzuki T, Inoue T, Tazawa C, Ono K, Moriya T, Saito H, Ishibashi T, Takahashi S, Yamada S, Sasano H 2005 Progesterone receptor subtypes in vascular smooth muscle cells of human aorta. *Endocr J* 52:245–252
676. Soma KK, Sinchak K, Lakhter A, Schlinger BA, Micevych PE 2005 Neurosteroids and female reproduction: estrogen increases 3β -HSD mRNA and activity in rat hypothalamus. *Endocrinology* 146:4386–4390
677. Simard J, Ricketts ML, Gingras S, Soucy P, Feltus FA, Melner MH 2005 Molecular biology of the 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ isomerase gene family. *Endocr Rev* 26:525–582
678. Inoue T, Akahira J, Suzuki T, Darnel AD, Kaneko C, Takahashi K, Hatori M, Shirane R, Kumabe T, Kurokawa Y, Satomi S, Sasano H 2002 Progesterone production and actions in the human central nervous system and neurogenic tumors. *J Clin Endocrinol Metab* 87:5325–5331
679. Simard J, Durocher F, Mebarki F, Turgeon C, Sanchez R, Labrie Y, Couet J, Trudel C, Rheume E, Morel Y, Luu-The V, Labrie F 1996 Molecular biology and genetics of the 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ isomerase gene family. *J Endocrinol* 150 Suppl:S189–S207
680. Payne AH, Abbaszade IG, Clarke TR, Bain PA, Park CH 1997 The multiple murine 3β -hydroxysteroid dehydrogenase isoforms: structure, function, and tissue- and developmentally specific expression. *Steroids* 62:169–175
681. Peng L, Arensburg J, Orly J, Payne AH 2002 The murine 3β -hydroxysteroid dehydrogenase (3β -HSD) gene family: a postulated role for 3β -HSD VI during early pregnancy. *Mol Cell Endocrinol* 187:213–221
682. Simpson ER, Misso M, Hewitt KN, Hill RA, Boon WC, Jones ME, Kovacic A, Zhou J, Clyne CD 2005 Estrogen—the good, the bad, and the unexpected. *Endocr Rev* 26:322–330
683. Bertelli G 2005 Sequencing of aromatase inhibitors. *Br J Cancer* 93(Suppl 1):S6–S9
684. Howell A 2005 New developments in the treatment of postmenopausal breast cancer. *Trends Endocrinol Metab* 16:420–428
685. Riggins RB, Bouton AH, Liu MC, Clarke R 2005 Antiestrogens, aromatase inhibitors, and apoptosis in breast cancer. *Vitam Horm* 71:201–237
686. Misso ML, Jang C, Adams J, Tran J, Murata Y, Bell R, Boon WC, Simpson ER, Davis SR 2005 Adipose aromatase gene expression is greater in older women and is unaffected by postmenopausal estrogen therapy. *Menopause* 12:210–215
687. Labrie F, Luu T, Labrie C, Simard J 2001 DHEA and its transformation into androgens and estrogens in peripheral target tissues: intracrinology. *Front Neuroendocrinol* 22:185–212
688. Labrie F, Luu-The V, Labrie C, Belanger A, Simard J, Lin SX, Pelletier G 2003 Endocrine and intracrine sources of androgens in women: inhibition of breast cancer and other roles of androgens and their precursor dehydroepiandrosterone. *Endocr Rev* 24:152–182
689. Orentreich N, Brind JL, Rizer RL, Vogelmann JH 1984 Age changes and sex differences in serum dehydroepiandrosterone sulfate concentrations throughout adulthood. *J Clin Endocrinol Metab* 59:551–555
690. Labrie F, Belanger A, Cusan L, Gomez JL, Candas B 1997 Marked decline in serum concentrations of adrenal C19 sex steroid precursors and conjugated androgen metabolites during aging. *J Clin Endocrinol Metab* 82:2396–2402
691. Mazat L, Lafont S, Debuire B, Tessier JF, Dartigues JF, Baulieu EE 2001 Prospective measurements of dehydroepiandrosterone sulfate in a cohort of elderly subjects: relationship to gender, subjective health, smoking habits, and 10-year mortality. *Proc Natl Acad Sci USA* 98:8145–8150
692. Lasley BL, Santoro N, Randolph JF, Gold EB, Crawford S, Weiss G, McConnell DS, Sowers MF 2002 The relationship of circulating dehydroepiandrosterone, testosterone, and estradiol to stages of the menopausal transition and ethnicity. *J Clin Endocrinol Metab* 87:3760–3767
693. Shideler SE, Gee NA, Chen J, Lasley BL 2001 Estrogen and progesterone metabolites and follicle-stimulating hormone in the aged macaque female. *Biol Reprod* 65:1718–1725
694. Baulieu EE 1997 Neurosteroids: of the nervous system, by the nervous system, for the nervous system. *Recent Prog Horm Res* 52:1–32
695. Baulieu EE 1981 Steroid hormones in the brain: several mechanisms? In: Fuxe K, Gustafsson JA, Wetterberg L, eds. Steroid hormone regulation of the brain. Oxford, UK: Pergamon Press; 3–14
696. Baulieu EE, Robel P, Schumacher M 2001 Neurosteroids: beginning of the story. *Int Rev Neurobiol* 46:1–32
697. Robel P, Schumacher M, Baulieu EE 1999 Neurosteroids: from definition and biochemistry to physiological function. In: Baulieu EE, Robel P, Schumacher M, eds. Neurosteroids. A new regulatory function in the nervous system. Totowa, NJ: Humana Press; 1–25
698. Mensah-Nyagan AG, Feuilloley M, Dupont E, Do-Rego JL, Lebloulenger F, Pelletier G, Vaudry H 1994 Immunocytochemical localization and biological activity of 3β -hydroxysteroid dehydrogenase in the central nervous system of the frog. *J Neurosci* 14:7306–7318
699. Sakamoto H, Ukena K, Tsutsui K 2001 Activity and localization of 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ -isomerase in the zebrafish central nervous system. *J Comp Neurol* 439:291–305
700. Usui M, Yamazaki T, Kominami S, Tsutsui K 1995 Avian neurosteroids. II. Localization of a cytochrome P450_{scc}-like substance in the quail brain. *Brain Res* 678:10–20
701. Hu MC, Chiang EF, Tong SK, Lai W, Hsu NC, Wang LC, Chung BC 2001 Regulation of steroidogenesis in transgenic mice and zebrafish. *Mol Cell Endocrinol* 171:9–14
702. Mensah-Nyagan AG, Beaujean D, Luu-The V, Pelletier G, Vaudry H 2001 Anatomical and biochemical evidence for the synthesis of unconjugated and sulfated neurosteroids in amphibians. *Brain Res Rev* 37:13–24
703. Mellon SH, Vaudry H 2001 Biosynthesis of neurosteroids and regulation of their synthesis. *Int Rev Neurobiol* 46:33–78
704. Guennoun R, Fiddes RJ, Gouézou M, Lombès M, Baulieu EE 1995 A key enzyme in the biosynthesis of neurosteroids, 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ - $\Delta 4$ -isomerase (3β -HSD), is expressed in rat brain. *Mol Brain Res* 30:287–300
705. Pasmanik M, Callard GV 1988 Changes in brain aromatase and 5α -reductase activities correlate significantly with seasonal reproductive cycles in goldfish (*Carassius auratus*). *Endocrinology* 122:1349–1356
706. Callard GV, Tchoudakova AV, Kishida M, Wood E 2001 Differential tissue distribution, developmental programming, estrogen regulation and promoter characteristics of CYP19 genes in teleost fish. *J Steroid Biochem Mol Biol* 79:305–314
707. Schlinger BA, Arnold AP 1991 Brain is the major site of estrogen synthesis in a male songbird. *Proc Natl Acad Sci USA* 88:4191–4194
708. Schlinger BA, Arnold AP 1992 Circulating estrogens in a male

- songbird originate in the brain. *Proc Natl Acad Sci USA* 89:7650–7653
709. **Balthazart J, Ball GF** 1998 New insights into the regulation and function of brain estrogen synthase (aromatase). *Trends Neurosci* 21:243–249
710. **Balthazart J, Baillien M, Ball GF** 2006 Rapid control of brain aromatase activity by glutamatergic inputs. *Endocrinology* 147:359–366
711. **Tsutsui K** 2006 Biosynthesis and organizing action of neurosteroids in the developing Purkinje cell. *Cerebellum* 5:89–96
712. **Payne AH, Hales DB** 2004 Overview of steroidogenic enzymes in the pathway from cholesterol to active steroid hormones. *Endocr Rev* 25:947–970
713. **Mellon SH, Deschepper CF** 1993 Neurosteroid biosynthesis: genes for adrenal steroidogenic enzymes are expressed in the brain. *Brain Res* 629:283–292
714. **Sanne JL, Krueger KE** 1995 Expression of cytochrome P450 side-chain cleavage enzyme and 3 β -hydroxysteroid dehydrogenase in the rat central nervous system: a study by polymerase chain reaction and *in situ* hybridization. *J Neurochem* 65:528–536
715. **Strömstedt M, Waterman MR** 1995 Messenger RNAs encoding steroidogenic enzymes are expressed in rodent brain. *Mol Brain Res* 34:75–88
716. **Kohchi C, Ukena K, Tsutsui K** 1998 Age- and region-specific expressions of the messenger RNAs encoding for steroidogenic enzymes p450scc, P450c17 and 3 β -HSD in the postnatal rat brain. *Brain Res* 801:233–238
717. **Mellon SH, Griffin LD, Compagnone NA** 2001 Biosynthesis and action of neurosteroids. *Brain Res Rev* 37:3–12
718. **Patte-Mensah C, Kappes V, Freund-Mercier MJ, Tsutsui K, Mensah-Nyagan AG** 2003 Cellular distribution and bioactivity of the key steroidogenic enzyme, cytochrome P450side chain cleavage, in sensory neural pathways. *J Neurochem* 86:1233–1246
719. **Patte-Mensah C, Kibaly C, Mensah-Nyagan AG** 2005 Substance P inhibits progesterone conversion to neuroactive metabolites in spinal sensory circuit: a potential component of nociception. *Proc Natl Acad Sci USA* 102:9044–9049
720. **Poisbeau P, Patte-Mensah C, Keller AF, Barrot M, Breton JD, Luis-Delgado OE, Freund-Mercier MJ, Mensah-Nyagan AG, Schlichter R** 2005 Inflammatory pain upregulates spinal inhibition via endogenous neurosteroid production. *J Neurosci* 25:11768–11776
721. **Cherradi N, Chambaz EM, Defaye G** 1995 Organization of 3 β -hydroxysteroid dehydrogenase/isomerase and cytochrome P450scc into a catalytically active molecular complex in bovine adrenocortical mitochondria. *J Steroid Biochem Mol Biol* 55:507–514
722. **Coirini H, Gouezou M, Liere P, Delespierre B, Pianos A, Eychenne B, Schumacher M, Guennoun R** 2002 3 β -Hydroxysteroid dehydrogenase expression in rat spinal cord. *Neuroscience* 113:883–891
723. **Ibanez C, Guennoun R, Liere P, Eychenne B, Pianos A, El Etr M, Baulieu EE, Schumacher M** 2003 Developmental expression of genes involved in neurosteroidogenesis: 3 β -hydroxysteroid dehydrogenase/ Δ 5- Δ 4 isomerase in the rat brain. *Endocrinology* 144:2902–2911
724. **Young J, Corpéchet C, Perche M, Haug M, Baulieu EE, Robel P** 1994 Neurosteroids: pharmacological effects of a 3 β -hydroxysteroid dehydrogenase inhibitor. *Endocrine J* 2:505–509
725. **Tsutsui K, Sakamoto H, Ukena K** 2003 A novel aspect of the cerebellum: biosynthesis of neurosteroids in the Purkinje cell. *Cerebellum* 2:215–222
726. **Guennoun R, Schumacher M, Robert F, Delespierre B, Gouézou M, Eychenne B, Akwa Y, Robel P, Baulieu EE** 1997 Neurosteroids: expression of functional 3 β -hydroxysteroid dehydrogenase by rat sensory neurons and Schwann cells. *Eur J Neurosci* 9:2236–2247
727. **Robert F, Guennoun R, Désarnaud F, Do-Thi A, Benmessahel Y, Baulieu EE, Schumacher M** 2001 Synthesis of progesterone in Schwann cells: regulation by sensory neurons. *Eur J Neurosci* 13:916–924
728. **Coirini H, Gouezou M, Delespierre B, Schumacher M, Guennoun R** 2003 3 β -Hydroxysteroid dehydrogenase isomerase (3 β -HSD) activity in the rat sciatic nerve: kinetic analysis and regulation by steroids. *J Steroid Biochem Mol Biol* 85:89–94
729. **Verleye M, Akwa Y, Liere P, Ladurelle N, Pianos A, Eychenne B, Schumacher M, Gillardin JM** 2005 The anxiolytic etiofoxine activates the peripheral benzodiazepine receptor and increases the neurosteroid levels in rat brain. *Pharmacol Biochem Behav* 82:712–720
730. **Labombarda F, Pianos A, Liere P, Eychenne B, Gonzalez S, Cambourg A, De Nicola AF, Schumacher M, Guennoun R** 2006 Injury elicited increase in spinal cord neurosteroid content analysed by gas chromatography mass spectrometry. *Endocrinology* 147:1847–1859
731. **Compagnone NA, Bulfone A, Rubenstein JL, Mellon SH** 1995 Steroidogenic enzyme P450c17 is expressed in the embryonic central nervous system. *Endocrinology* 136:5212–5223
732. **Zwain IH, Yen SS** 1999 Dehydroepiandrosterone: biosynthesis and metabolism in the brain. *Endocrinology* 140:880–887
733. **Kibaly C, Patte-Mensah C, Mensah-Nyagan AG** 2005 Molecular and neurochemical evidence for the biosynthesis of dehydroepiandrosterone in the adult rat spinal cord. *J Neurochem* 93:1220–1230
734. **Davies E, MacKenzie SM** 2003 Extra-adrenal production of corticosteroids. *Clin Exp Pharmacol Physiol* 30:437–445
735. **Hojo Y, Hattori TA, Enami T, Furukawa A, Suzuki K, Ishii HT, Mukai H, Morrison JH, Janssen WG, Kominami S, Harada N, Kimoto T, Kawato S** 2004 Adult male rat hippocampus synthesizes estradiol from pregnenolone by cytochromes P45017 α and P450 aromatase localized in neurons. *Proc Natl Acad Sci USA* 101:865–870
736. **Mukai H, Takata N, Ishii HT, Tanabe N, Hojo Y, Furukawa A, Kimoto T, Kawato S** 2006 Hippocampal synthesis of estrogens and androgens which are paracrine modulators of synaptic plasticity: synaptocrinology. *Neuroscience* 138:757–764
737. **Le Goascogne C, Eychenne B, Tonon MC, Lachapelle F, Baumann N, Robel P** 2000 Neurosteroid progesterone is up-regulated in the brain of jimpy and shiverer mice. *Glia* 29:14–24
738. **Saredi S, Patte-Mensah C, Melcangi RC, Mensah-Nyagan AG** 2005 Effect of streptozotocin-induced diabetes on the gene expression and biological activity of 3 β -hydroxysteroid dehydrogenase in the rat spinal cord. *Neuroscience* 135:869–877
739. **Patte-Mensah C, Kibaly C, Boudard D, Schaeffer V, Begle A, Saredi S, Meyer L, Mensah-Nyagan AG** 2006 Neurogenic pain and steroid synthesis in the spinal cord. *J Mol Neurosci* 28:17–31
740. **Schlichter R, Keller AF, De Roo M, Breton JD, Inquimbert P, Poisbeau P** 2006 Fast nongenomic effects of steroids on synaptic transmission and role of endogenous neurosteroids in spinal pain pathways. *J Mol Neurosci* 28:33–51
741. **Schaeffer V, Patte-Mensah C, Eckert A, Mensah-Nyagan AG** 2006 Modulation of neurosteroid production in human neuroblastoma cells by Alzheimer's disease key proteins. *J Neurobiol* 66:868–881
742. **Garcia-Segura LM, Wozniak A, Azcoitia I, Rodriguez JR, Hutchison RE, Hutchison JB** 1999 Aromatase expression by astrocytes after brain injury: implications for local estrogen formation in brain repair. *Neuroscience* 89:567–578
743. **Azcoitia I, DonCarlos LL, Garcia-Segura LM** 2003 Are gonadal steroid hormones involved in disorders of brain aging? *Aging Cell* 2:31–37
744. **Azcoitia I, Sierra A, Veiga S, Garcia-Segura LM** 2005 Brain steroidogenesis: emerging therapeutic strategies to prevent neurodegeneration. *J Neural Transm* 112:171–176
745. **Veiga S, Melcangi RC, DonCarlos LL, Garcia-Segura LM, Azcoitia I** 2004 Sex hormones and brain aging. *Exp Gerontol* 39:1623–1631
746. **Beyenburg S, Stoffel-Wagner B, Bauer J, Watzka M, Blumcke J, Bidlingmaier F, Elger CE** 2001 Neuroactive steroids and seizure susceptibility. *Epilepsy Res* 44:141–153
747. **Stoffel-Wagner B** 2001 Neurosteroid metabolism in the human brain. *Eur J Endocrinol* 145:669–679
748. **Weill-Engerer S, David JP, Sazdovitch V, Liere P, Schumacher M, Delacourte A, Baulieu EE, Akwa Y** 2003 *In vitro* metabolism of dehydroepiandrosterone (DHEA) to 7 α -hydroxy-DHEA and Δ 5-androstene-3 β ,17 β -diol in specific regions of the aging brain from Alzheimer's and non-demented patients. *Brain Res* 969:117–125
749. **Le Goascogne C, Gouézou M, Robel P, Defaye G, Chambaz E, Waterman MR, Baulieu EE** 1989 The cholesterol side-chain cleav-

- age complex in human brain white matter. *J Neuroendocrinol* 1:153–156
750. **Beyenburg S, Stoffel-Wagner B, Watzka M, Blumcke I, Bauer J, Schramm J, Bidlingmaier F, Elger CE** 1999 Expression of cytochrome P450scc mRNA in the hippocampus of patients with temporal lobe epilepsy. *Neuroreport* 10:3067–3070
751. **Watzka M, Bidlingmaier F, Schramm J, Klingmuller D, Stoffel-Wagner B** 1999 Sex- and age-specific differences in human brain CYP11A1 mRNA expression. *J Neuroendocrinol* 11:901–905
752. **Yu L, Romero DG, Gomez-Sanchez CE, Gomez-Sanchez EP** 2002 Steroidogenic enzyme gene expression in the human brain. *Mol Cell Endocrinol* 190:9–17
753. **King SR, Manna PR, Ishii T, Syapin PJ, Ginsberg SD, Wilson K, Walsh LP, Parker KL, Stocco DM, Smith RG, Lamb DJ** 2002 An essential component in steroid synthesis, the steroidogenic acute regulatory protein, is expressed in discrete regions of the brain. *J Neurosci* 22:10613–10620
754. **Steckelbroeck S, Watzka M, Reichelt R, Hans VH, Stoffel-Wagner B, Heidrich DD, Schramm J, Bidlingmaier F, Klingmuller D** 2001 Characterization of the 5 α -reductase-3 α -hydroxysteroid dehydrogenase complex in the human brain. *J Clin Endocrinol Metab* 86:1324–1331
755. **Stoffel-Wagner B, Watzka M, Steckelbroeck S, Ludwig M, Clusmann H, Bidlingmaier F, Casarosa E, Luisi S, Elger CE, Beyenburg S** 2003 Allopregnanolone serum levels and expression of 5 α -reductase and 3 α -hydroxysteroid dehydrogenase isoforms in hippocampal and temporal cortex of patients with epilepsy. *Epilepsy Res* 54:11–19
756. **Wozniak A, Hutchison RE, Morris CM, Hutchison JB** 1998 Neuroblastoma and Alzheimer's disease brain cells contain aromatase activity. *Steroids* 63:263–267
757. **Steckelbroeck S, Heidrich DD, Stoffel-Wagner B, Hans VH, Schramm J, Bidlingmaier F, Klingmuller D** 1999 Characterization of aromatase cytochrome P450 activity in the human temporal lobe. *J Clin Endocrinol Metab* 84:2795–2801
758. **Hammond GL, Hirvonen J, Vihko R** 1983 Progesterone, androstenedione, testosterone, 5 α -dihydrotestosterone and androstereone concentrations in specific regions of the human brain. *J Steroid Biochem* 18:185–189
759. **Lanthier A, Patwardhan VV** 1986 Sex steroids and 5-en-3 β -hydroxysteroids in specific regions of the human brain and cranial nerves. *J Steroid Biochem* 25:445–449
760. **Lacroix C, Fiet J, Benais JP, Gueux B, Bonete R, Villette JM, Gourmel B, Dreux C** 1987 Simultaneous radioimmunoassay of progesterone, androst-4-enedione, pregnenolone, dehydroepiandrosterone and 17-hydroxyprogesterone in specific regions of human brain. *J Steroid Biochem* 28:317–325
761. **Morfín R, Young J, Corpéchet C, Egestad B, Sjövall J, Baulieu EE** 1992 Neurosteroids: pregnenolone in human sciatic nerves. *Proc Natl Acad Sci USA* 89:6790–6793
762. **Bixo M, Andersson A, Winblad B, Purdy RH, Bäckström T** 1997 Progesterone, 5 α -pregnane-3,20-dione and 3 α -hydroxy-5 α -pregnane-20-one in specific regions of the human female brain in different endocrine states. *Brain Res* 764:173–178
763. **Bixo M, Bäckström T, Winblad B, Andersson A** 1995 Estradiol and testosterone in specific regions of the human female brain in different endocrine states. *J Steroid Biochem Mol Biol* 55:297–303
764. **Twist SJ, Taylor GA, Weddell A, Weightman DR, Edwardson JA, Morris JA** 2000 Brain oestradiol and testosterone levels in Alzheimer's disease. *Neurosci Lett* 286:1–4
765. **Yue X, Lu M, Lancaster T, Cao P, Honda S, Staufenbiel M, Harada N, Zhong Z, Shen Y, Li R** 2005 Brain estrogen deficiency accelerates A β plaque formation in an Alzheimer's disease animal model. *Proc Natl Acad Sci USA* 102:19198–19203
766. **Shen R, Sumitomo M, Dai J, Hardy DO, Navarro D, Usmani B, Papandreou CN, Hersh LB, Shipp MA, Freedman LP, Nanus DM** 2000 Identification and characterization of two androgen response regions in the human neutral endopeptidase gene. *Mol Cell Endocrinol* 170:131–142
767. **Cheney DL, Uzunov D, Costa E, Guidotti A** 1995 Gas chromatographic-mass fragmentographic quantitation of 3 α -hydroxy-5 α -pregnan-20-one (allopregnanolone) and its precursors in blood and brain of adrenalectomized and castrated rats. *J Neurosci* 15:4641–4650
768. **Liere P, Akwa Y, Weill-Engerer S, Eychenne B, Pianos A, Robel P, Sjövall J, Schumacher M, Baulieu EE** 2000 Validation of an analytical procedure to measure trace amounts of neurosteroids in brain tissue by gas chromatography-mass spectrometry. *J Chromatogr B* 739:301–312
769. **Liere P, Pianos A, Eychenne B, Cambourg A, Liu S, Griffiths W, Schumacher M, Sjövall J, Baulieu EE** 2004 Novel lipoidal derivatives of pregnenolone and dehydroepiandrosterone and absence of their sulfated counterparts in rodent brain. *J Lipid Res* 45:2287–2302
770. **Weill-Engerer S, David JP, Sazdovitch V, Liere P, Eychenne B, Pianos A, Schumacher M, Delacourte A, Baulieu EE, Akwa Y** 2002 Neurosteroid quantification in human brain regions: comparison between Alzheimer's and non-demented patients. *J Clin Endocrinol Metab* 87:5138–5143
771. **Morley JE, Kaiser F, Raum WJ, Perry HM, Flood JF, Jensen J, Silver AJ, Roberts E** 1997 Potentially predictive and manipulable blood serum correlates of aging in the healthy human male: progressive decreases in bioavailable testosterone, dehydroepiandrosterone sulfate, and the ratio of insulin-like growth factor 1 to growth hormone. *Proc Natl Acad Sci USA* 94:7537–7542
772. **Hill M, Lukac D, Lapcik O, Sulcova J, Hampl R, Pouzar V, Starka L** 1999 Age relationships and sex differences in serum levels of pregnenolone and 17-hydroxypregnenolone in healthy subjects. *Clin Chem Lab Med* 37:439–447
773. **Zhang P, Rodriguez H, Mellon SH** 1995 Transcriptional regulation of P450scc gene expression in neural and steroidogenic cells: implications for regulation of neurosteroidogenesis. *Mol Endocrinol* 9:1571–1582
774. **Hammer F, Compagnone NA, Vigne JL, Bair SR, Mellon SH** 2004 Transcriptional regulation of P450scc gene expression in the embryonic rodent nervous system. *Endocrinology* 145:901–912
775. **Akwa Y, Sananes N, Gouezou M, Robel P, Baulieu EE, Le Goascogne C** 1993 Astrocytes and neurosteroids: metabolism of pregnenolone and dehydroepiandrosterone. Regulation by cell density. *J Cell Biol* 121:135–143
776. **Guarneri P, Guarneri R, Cascio C, Piccoli F, Papadopoulos V** 1995 γ -Aminobutyric acid type A/benzodiazepine receptors regulate rat retina neurosteroidogenesis. *Brain Res* 683:65–72
777. **Do-Rego JL, Mensah-Nyagan GA, Beaujean D, Vaudry D, Sieghart W, Luu-The V, Pelletier G, Vaudry H** 2000 γ -Aminobutyric acid, acting through γ -aminobutyric acid type A receptors, inhibits the biosynthesis of neurosteroids in the frog hypothalamus. *Proc Natl Acad Sci USA* 97:13925–13930
778. **Do-Rego JL, Mensah-Nyagan AG, Beaujean D, Leprince J, Tonon MC, Luu-The V, Pelletier G, Vaudry H** 2001 The octadecanuropeptide ODN stimulates neurosteroid biosynthesis through activation of central-type benzodiazepine receptors. *J Neurochem* 76:128–138
779. **Beaujean D, Do R, Galas L, Mensah N, Fredriksson R, Larhammar D, Fournier A, Luu T, Pelletier G, Vaudry H** 2002 Neuropeptide Y inhibits the biosynthesis of sulfated neurosteroids in the hypothalamus through activation of Y(1) receptors. *Endocrinology* 143:1950–1963
780. **Do-Rego JL, Acharjee S, Seong JY, Galas L, Alexandre D, Bizet P, Burlet A, Kwon HB, Luu-The V, Pelletier G, Vaudry H** 2006 Vasotocin and mesotocin stimulate the biosynthesis of neurosteroids in the frog brain. *J Neurosci* 26:6749–6760
781. **Micevych P, Sinchak K, Mills RH, Tao L, LaPolta P, Lu JK** 2003 The luteinizing hormone surge is preceded by an estrogen-induced increase of hypothalamic progesterone in ovariectomized and adrenalectomized rats. *Neuroendocrinology* 78:29–35
782. **Micevych PE, Chaban V, Ogi J, Dewing P, Lu JK, Sinchak K** 2007 Estradiol stimulates progesterone synthesis in hypothalamic astrocyte cultures. *Endocrinology* 148:782–789
783. **Papadopoulos V, Baraldi M, Guilarte TR, Knudsen TB, Lacapere JJ, Lindemann P, Norenberg MD, Nutt D, Weizman A, Zhang MR, Gavish M** 2006 Translocator protein (18 kDa): new nomenclature for the peripheral-type benzodiazepine receptor based on its structure and molecular function. *Trends Pharmacol Sci* 27:402–409

784. **Papadopoulos V** 1993 Peripheral-type benzodiazepine/diazepam binding inhibitor receptor: biological role in steroidogenic cell function. *Endocr Rev* 14:222–240
785. **Papadopoulos V, Lecanu L, Brown RC, Han Z, Yao ZX** 2006 Peripheral-type benzodiazepine receptor in neurosteroid biosynthesis, neuropathology and neurological disorders. *Neuroscience* 138:749–756
786. **Papadopoulos V, Guarneri P, Kreuger KE, Guidotti A, Costa E** 1992 Pregnenolone biosynthesis in C6–2B glioma cell mitochondria: regulation by a mitochondrial diazepam binding inhibitor receptor. *Proc Natl Acad Sci USA* 89:5113–5117
787. **Korneyev A, Pan BS, Polo A, Romeo E, Guidotti A, Costa E** 1993 Stimulation of brain pregnenolone synthesis by mitochondrial diazepam binding inhibitor receptor ligands in vivo. *J Neurochem* 61:1515–1524
788. **Romeo E, Cavallaro S, Korneyev A, Kozikowski AP, Ma D, Polo A, Costa E, Guidotti A** 1993 Stimulation of brain steroidogenesis by 2-aryl-indole-3-acetamide derivatives acting at the mitochondrial diazepam-binding inhibitor receptor complex. *J Pharmacol Exp Ther* 267:462–471
789. **Bitran D, Foley M, Audette D, Leslie N, Frye CA** 2000 Activation of peripheral mitochondrial benzodiazepine receptors in the hippocampus stimulates allopregnanolone synthesis and produces anxiolytic-like effects in the rat. *Psychopharmacology* 151:64–71
790. **Serra M, Madau P, Chessa MF, Caddeo M, Sanna E, Trapani G, Franco M, Liso G, Purdy RH, Barbaccia ML, Biggio G** 1999 2-Phenyl-imidazo[1,2-a]pyridine derivatives as ligands for peripheral benzodiazepine receptors: stimulation of neurosteroid synthesis and anticonflict action in rats. *Br J Pharmacol* 127:177–187
791. **Trapani G, Franco M, Latrofa A, Ricciardi L, Carotti A, Serra M, Sanna E, Biggio G, Liso G** 1999 Novel 2-phenylimidazo[1,2-a]pyridine derivatives as potent and selective ligands for peripheral benzodiazepine receptors: synthesis, binding affinity, and in vivo studies. *J Med Chem* 42:3934–3941
792. **Ferzaz B, Brault E, Bourliand G, Robert JP, Poughon G, Claustre Y, Marguet F, Liere P, Schumacher M, Nowicki JP, Fournier J, Marabout B, Sevrin M, George P, Soubrie P, Benavides J, Scatton B** 2002 SSR180575 (7-chloro-N,N,5-trimethyl-4-oxo-3-phenyl-3,5-dihydro-4H-pyridazino[4,5-b]indole-1-acetamide), a peripheral benzodiazepine receptor ligand, promotes neuronal survival and repair. *J Pharmacol Exp Ther* 301:1067–1078
793. **Lacor P, Gandolfo P, Tonon MC, Brault E, Dalibert I, Schumacher M, Benavides J, Ferzaz B** 1999 Regulation of the expression of peripheral benzodiazepine receptors and their endogenous ligands during rat sciatic nerve degeneration and regeneration: a role for PBR in neurosteroidogenesis. *Brain Res* 815:70–80
794. **Garcia-Ovejero D, Azcoitia I, DonCarlos LL, Melcangi RC, Garcia-Segura LM** 2005 Glia-neuron crosstalk in the neuroprotective mechanisms of sex steroid hormones. *Brain Res Rev* 48:273–286
795. **Kassiou M, Meikle SR, Banati RB** 2005 Ligands for peripheral benzodiazepine binding sites in glial cells. *Brain Res Rev* 48:207–210
796. **Vowinckel E, Reutens D, Becher B, Verge G, Evans A, Owens T, Antel JP** 1997 PK11195 binding to the peripheral benzodiazepine receptor as a marker of microglia activation in multiple sclerosis and experimental autoimmune encephalomyelitis. *J Neurosci* 17:345–353
797. **Leonelli E, Yague JG, Ballabio M, Azcoitia I, Magnaghi V, Schumacher M, Garcia-Segura LM, Melcangi RC** 2005 Ro5–4864, a synthetic ligand of peripheral benzodiazepine receptor, reduces aging-associated myelin degeneration in the sciatic nerve of male rats. *Mech Ageing Dev* 126:1159–1163
798. **Ryu JK, Choi HB, McLarnon JG** 2005 Peripheral benzodiazepine receptor ligand PK11195 reduces microglial activation and neuronal death in quinolinic acid-injected rat striatum. *Neurobiol Dis* 20:550–561
799. **Mills CD, Bitler JL, Woolf CJ** 2005 Role of the peripheral benzodiazepine receptor in sensory neuron regeneration. *Mol Cell Neurosci* 30:228–237
800. **Veiga S, Azcoitia I, Garcia-Segura LM** 2005 Ro5–4864, a peripheral benzodiazepine receptor ligand, reduces reactive gliosis and protects hippocampal hilar neurons from kainic acid excitotoxicity. *J Neurosci Res* 80:129–137
801. **Galiegue S, Tinel N, Casellas P** 2003 The peripheral benzodiazepine receptor: a promising therapeutic drug target. *Curr Med Chem* 10:1563–1572
802. **Azarashvili T, Krestinina O, Yurkov I, Evtodienko Y, Reiser G** 2005 High-affinity peripheral benzodiazepine receptor ligand, PK11195, regulates protein phosphorylation in rat brain mitochondria under control of Ca(2+). *J Neurochem* 94:1054–1062
803. **Stocco DM** 2001 StAR protein and the regulation of steroid hormone biosynthesis. *Annu Rev Physiol* 63:193–213
804. **Stocco DM, Wang X, Jo Y, Manna PR** 2005 Multiple signaling pathways regulating steroidogenesis and steroidogenic acute regulatory protein expression: more complicated than we thought. *Mol Endocrinol* 19:2647–2659
805. **Stocco DM** 2002 Clinical disorders associated with abnormal cholesterol transport: mutations in the steroidogenic acute regulatory protein. *Mol Cell Endocrinol* 191:19–25
806. **Haut T, Yao ZX, Bose HS, Wall CT, Han Z, Li W, Hales DB, Miller WL, Culty M, Papadopoulos V** 2005 Peripheral-type benzodiazepine receptor-mediated action of steroidogenic acute regulatory protein on cholesterol entry into Leydig cell mitochondria. *Mol Endocrinol* 19:540–554
807. **Sierra A, Lavaque E, Perez-Martin M, Azcoitia I, Hales DB, Garcia-Segura LM** 2003 Steroidogenic acute regulatory protein in the rat brain: cellular distribution, developmental regulation and overexpression after injury. *Eur J Neurosci* 18:1458–1467
808. **Sierra A** 2004 Neurosteroids: the StAR protein in the brain. *J Neuroendocrinol* 16:787–793
809. **Luo L, Chen H, Zirkin BR** 2005 Temporal relationships among testosterone production, steroidogenic acute regulatory protein (StAR), and P450 side-chain cleavage enzyme (P450scc) during Leydig cell aging. *J Androl* 26:25–31
810. **Lavaque E, Sierra A, Azcoitia I, Garcia-Segura LM** 2006 Steroidogenic acute regulatory protein in the brain. *Neuroscience* 138:741–747
811. **Benmessahel Y, Troadec JD, Cadepond F, Guennoun R, Hales DB, Schumacher M, Groyer G** 2004 Downregulation of steroidogenic acute regulatory protein (StAR) gene expression by cyclic AMP in cultured Schwann cells. *Glia* 45:213–228
812. **Schlichter R, Rybalchenko V, Poisbeau P, Verleye M, Gillardin J** 2000 Modulation of GABAergic synaptic transmission by the non-benzodiazepine anxiolytic etifoxine. *Neuropharmacology* 39:1523–1535
813. **Hamon A, Morel A, Hue B, Verleye M, Gillardin JM** 2003 The modulatory effects of the anxiolytic etifoxine on GABA(A) receptors are mediated by the β subunit. *Neuropharmacology* 45:293–303
814. **Rupprecht R, di Michele F, Hermann B, Strohle A, Lancel M, Romeo E, Holsboer F** 2001 Neuroactive steroids: molecular mechanisms of action and implications for neuropsychopharmacology. *Brain Res Rev* 37:59–67
815. **Eser D, Schule C, Baghai TC, Romeo E, Uzunov DP, Rupprecht R** 2006 Neuroactive steroids and affective disorders. *Pharmacol Biochem Behav* 84:656–666
816. **Uzunov DP, Cooper TB, Costa E, Guidotti A** 1996 Fluoxetine-elicited changes in brain neurosteroid content measured by negative ion mass fragmentography. *Proc Natl Acad Sci USA* 93:12599–12604
817. **Pinna G, Costa E, Guidotti A** 2006 Fluoxetine and norfluoxetine stereospecifically and selectively increase brain neurosteroid content at doses that are inactive on 5-HT reuptake. *Psychopharmacology (Berl)* 186:362–372
818. **Griffin LD, Mellon SH** 1999 Selective serotonin reuptake inhibitors directly alter activity of neurosteroidogenic enzymes. *Proc Natl Acad Sci USA* 96:13512–13517
819. **Uzunova V, Sheline Y, Davis JM, Rasmusson A, Uzunov DP, Costa E, Guidotti A** 1998 Increase in the cerebrospinal fluid content of neurosteroids in patients with unipolar major depression who are receiving fluoxetine or fluvoxamine. *Proc Natl Acad Sci USA* 95:3239–3244
820. **Romeo E, Strohle A, Spalletta G, di MF, Hermann B, Holsboer F, Pasini A, Rupprecht R** 1998 Effects of antidepressant treatment on neuroactive steroids in major depression. *Am J Psychiatry* 155:910–913



821. **Dubrovsky B** 2006 Neurosteroids, neuroactive steroids, and symptoms of affective disorders. *Pharmacol Biochem Behav* 84:644–655
822. **Uzunova V, Sampson L, Uzunov DP** 2006 Relevance of endogenous 3α -reduced neurosteroids to depression and antidepressant action. *Psychopharmacology (Berl)* 186:351–361
823. **Smith RG, Betancourt L, Sun Y** 2005 Molecular endocrinology and physiology of the aging central nervous system. *Endocr Rev* 26:203–250
824. **Mendez P, Azcoitia I, Garcia-Segura LM** 2005 Interdependence of oestrogen and insulin-like growth factor-I in the brain: potential for analysing neuroprotective mechanisms. *J Endocrinol* 185:11–17
825. **Berchtold NC, Kessler JP, Pike CJ, Adlard PA, Cotman CW** 2001 Estrogen and exercise interact to regulate brain-derived neurotrophic factor mRNA and protein expression in the hippocampus. *Eur J Neurosci* 14:1992–2002
826. **Sitruk-Ware R** 2000 Progestins in hormonal replacement therapy and prevention of endometrial disease. In: Sitruk-Ware R, Mishell DR, eds. *Progestins and antiprogestins in clinical practice*. New York: Marcel Dekker; 269–287
827. **Sitruk-Ware R** 2006 New progestagens for contraceptive use. *Hum Reprod Update* 12:169–178
828. **Schindler AE** 2003 Differential effects of progestins on hemostasis. *Maturitas* 46(Suppl 1):S31–S37
829. **Sitruk-Ware R** 2000 Progestins and cardiovascular risk markers. *Steroids* 65:651–658
830. **Sitruk-Ware R, Plu-Bureau** 2004 Exogenous progestagens and the human breast. *Maturitas* 49:58–66

Endocrine Reviews is published by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.